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The forests of Western Ghats, an abode of novel and interesting microfungi*

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"Nahī jñānena sadnusham pavitramahi vidyayā"
 (=Nothing is more purer than knowledge)

- Lord Shri Krishna in *Bhagavad Gita*

I am deeply honoured to have been elected as President of Mycological Society of India. I am equally thankful to Thapar University, Patiala, for organizing the 34th Annual meeting of the Society along with a National Symposium.

Mycological Society of India

The Mycological Society of India was established in January 1973 in Chennai (formerly Madras), on the sidelines of International Symposium on Taxonomy of Fungi held at Centre for Advanced Studies in Botany, University of Madras. The main architect of the event was Professor C.V. Subramanian. I joined the Botany Department, at a slightly later date in the same year, for my post-M.Sc. Diploma in Mycology and Plant Pathology and subsequently for doctoral studies. I narrowly missed the opportunity of witnessing the birth of the Society. Nevertheless, I have been associated with the Society ever since, in various capacities which I considered as a privilege. The Society has completed 33 years and I am very glad that a large number of young students of mycology are now enrolling as Members.

Centre for Advanced Studies in Botany, University of Madras

I did my doctoral thesis under the supervision of Professor C.V. Subramanian and, subsequently also worked on a UGC-sponsored project entitled 'Fungus Flora of South India'. My stay at the Centre for Advanced Studies in Botany, University of Madras, for duration of six years was most rewarding. Invaluable guidance from Professor Subramanian and numerous mycological events and interactions that took place those days together provided me not only a foundation on fungal biology but everything that I know today.

Forests of Western Ghats, southern India (Fig.1)

The Western Ghats, hilly and mountainous terrain on the western side of peninsular India, about 1500 km stretch parallel to the coastline between 8° N to 22°5' N and 72°5' E to 76°5' E, extend from river Tapti southward through the States of Maharashtra, Goa, Karnataka, Kerala and Tamil Nadu, up to

Kanyakumari. These mountains are 30–80 km away from the sea coast. The escarpment is steep and precipitous along the western side and gently inclined on the east, the highest elevation is at Anamudi in Kerala. Many

short, fast flowing, seasonal streams and perennial rivers originate in the Western Ghats and flow down to the Arabian Sea. The ghats receive south-west monsoon rain from June to September; the downpour is heavy on the western side of the ghat ranging from 200 to 350 cm annually. The mean annual temperature ranges between 26°C to 37°C, which seldom falls below 16°C. The mean annual relative humidity is about 80%. Under these warm and humid conditions, luxuriant forests flourish in the windward western side of the ghats, which include wet-evergreens, sholas, moist-deciduous, dry-deciduous, scrub-jungles and plateau-grasslands. Diverse plant species flourish in the forests of Western Ghats (Pascal, 1989).

Fungi of the Western Ghats have been explored and documented to some extent (Subramanian, 1971; Rao and de Hoog, 1986; Subramanian and Bhat, 1987; Bhat and Kendrick, 1993; Sarbhoy *et al.*, 1986; Sarbhoy *et al.*, 1996; Jamaluddin *et al.*, 2004; Manoharachary *et al.*, 2005; Jacob, 2000; D'Souza, 2002; Nair, 2002; Keshav Prasad, 2003; Jalmi, 2006; Gawas, 2008). Despite the hitherto efforts, vast area of peninsular India still remained unexplored.

Encounters with micro-fungi

My doctoral studies on Hypocrealean fungi and subsequent project works on floristics of Hyphomycetes demanded extensive field work. In the last three decades, in search of microfungi, I wandered around numerous forests of the Western Ghats (Fig. 1), from Kanyakumari (Tamil Nadu) to Mahabaleshwar (Maharashtra) through the States of Kerala, Karnataka

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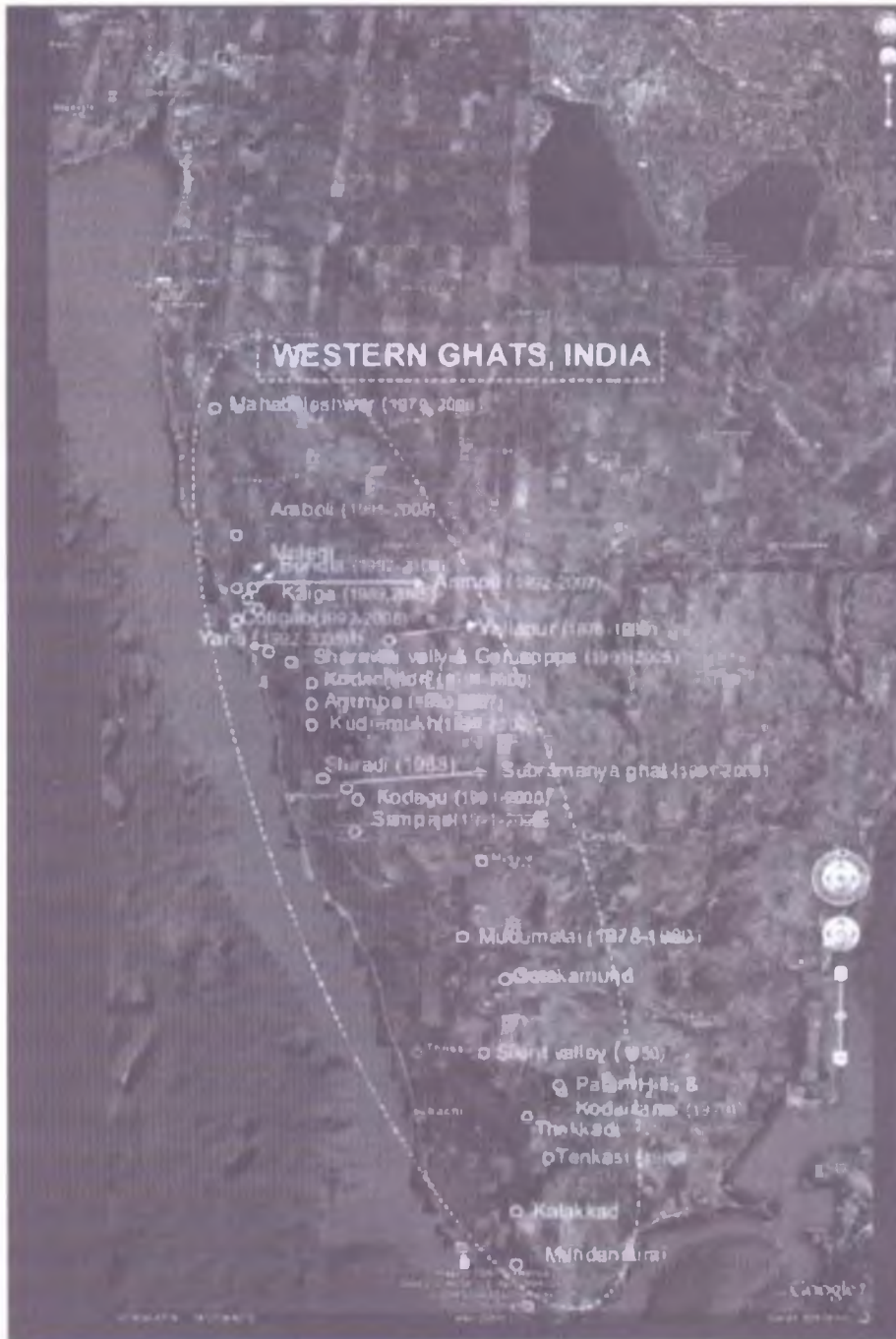


Fig. 1. Various localities in the forests of Western Ghats, India, visited by the author during 1976-2007

and Goa, and scanned several types of substrates which included decaying plant litter, freshwater foam, aerial plant parts, internal of fresh leaves and twigs, herbivore dung, dead insects, etc. This enduring search for microfungi provided me some insight on hyphomycete diversity of the region. In this address, I will highlight the excitements derived while encountering some of the new hyphomycetous fungi in the forests of Western Ghats.

Hyphomycetes from the forests of Western Ghats

It was in the monsoon of 1976, in search of Hypocrealean fungi, I set out on my first field trip into

the forests of Western Ghats in Coorg (now Kodagu), located in the State of Karnataka. Moist chamber incubated dead and decaying twigs of *Coffea Arabica* L., gathered from Abby Falls, near Madikeri, under stereoscope exhibited glistening, appendaged, colourless, spherical spores in masses on phialidic conidiophores. This was nothing to do with Hypocrealean fungi; yet, I showed the slides to Professor Subramanian. The mycology laboratory of Centre for Advanced Studies in Botany at Madras University then had total command on conidial fungi, at global level. Professor Subramanian was instant in declaring that the fungus might be an unknown one and I should examine it further. Continued

detailed studies in the next few months, resulted with description of *Bahusutrabeeja dwaya* Subram. et Bhat, a new species in a new genus (Subramanian and Bhat, 1977a). Though Hypocrealean fungi were my focus, in my inner-self, I developed special interest on hyphomycetous fungi. It was my firm conviction then that 'knowledge gaining exercises will surpass everything else'.

A phialidic hyphomycete, *Putagraivam sundaram* Subram. et Bhat (Subramanian and Bhat, 1977b), anamorphic phase of a hypocrealean *Peethambara sundara* Subram. et Bhat (Subramanian and Bhat, 1977c), that I gathered from my native village, Darbhe (Irde) located in the foothills of Western Ghats, Karnataka State, was an accidental discovery. Closely and curiously watching my fungus collecting methods, my father who didn't even have any formal education, brought a head-load of decaying twigs of *Macaranga indica* Wright which had bright-coloured stromatic ascocarps on the surface, from the nearby forests. The ascocarps of *Peethambara sundara*, a new hypocrealean fungus, had accompanying conidial synnemata on the substrate. Single ascospore in culture also produced the synnematosus conidial phase. The study ended up in the description of hitherto unknown teleomorph-anamorph connection of the new fungus. *Putagraivam sundaram* is now a synonym of *Didymostilbe sundara* (Seifert, 1985).

My two visits to Silent Valley in Kerala during March-April and December 1980 for collection of fungi were most memorable. Led by the Botanical Survey of India, I was part of a big team set out to study the extent of biodiversity of the region. I collected samples of fungi from all over the forests and streams over a period of 6 weeks. Together with Professor Subramanian, I examined every bit of the samples and documented the fungi (Subramanian, 1992; Subramanian and Bhat, 1987). This was a major fungal biodiversity study of the region. There is no exaggeration if I state that the then proposed plan of construction of a major hydro-electric dam across Kunthi puzza river in Silent Valley was permanently scrapped because of the evidences provided by the high biodiversity of the region, especially the fungi.

As a project staff in the 'Fungus Flora of South India' programme, I collected samples of fungi from different forests of Karnataka, Kerala and Tamilnadu, during May-December 1980. In one of the field trips, near Hassan, Karnataka State, our vehicle was hit by a huge vulture and the van fell into a 20 m deep ditch. Miraculous as it was, we escaped with very minor injuries. Nothing of the sort deterred us from continuing investigations on fungi. I do remember my first visit to Goa (Dudhsagar Falls), during November

1980. We traveled in a train engine to the Falls from Londa railway station and I had an hour-long swim in the pond adjacent to the railway track. I had no idea those days that I would continue my quest on fungi, subsequently from Goa University.

A brief with fungi of Ethiopia

During 1981-88, I had worked in Asmara University, Ethiopia (presently Eritrea). Positioned in Asmara, I had a distinct opportunity to survey the highland forests of southern Ethiopia. From fungal diversity study point of view, Ethiopia is equally rich and, undoubtedly, those were some of my very productive mycological study period (Bhat, 1983, 1984, 1985, 1986; Bhat and Sutton, 1984, 1985, 1986; Bhat and Chien, 1990, 1992).

Return to the Western Ghats

My interest on hyphomycetes grew steadily and progressively, on return to India. In the subsequent years, while working in Mangalore (1988-89) and Goa Universities (1990-2007), along with my students and colleagues, I surveyed many forests and streams of the Western Ghats and collected fungal samples from everywhere. We trekked long distances; stayed overnights in deep jungles, nearby bus stands and railway platforms. Collection and description of fungi were sheer excitements. Our mycofloristic investigations were not confined to the forests of Western Ghats alone. We also surveyed the forests of Andaman Islands (Bhat and Kendrick, 1993; D'Souza *et al.*, 2002) and north-eastern highlands of Arunachal Pradesh, Assam and Nagaland in India (Bhat, 2006).

I had invaluable support from Professor K.M. Kaveriappa, Vice Chancellor, Mangalore University, Professors K. Natarajan and B.P.R. Vittal, Centre for Advanced Studies in Botany, University of Madras; Dr B. Sutton, International Mycological Institute, UK; Professor B. Kendrick, University of Waterloo, Canada; and Dr. Kevin D. Hyde, University of Hong Kong (now in Mae Fah Luang University, Thailand), in my mycological journey. They not only accommodated me in their work places for a while but also supplied needful literature on fungi. Major fungal haul came from the wet-evergreen forests of Western Ghats in Silent Valley in Kerala State (Subramanian and Bhat, 1981, 1987-88), Agumbe, Sampaje, Kodachadri hills and Sharavati valley in Karnataka State (Bhat and Kendrick, 1993) and Bondla, Molem and Cotigao in Goa State (Bhat *et al.*, 2009). Not only many new but also numerous common, rare, interesting microfungi were discovered by us. The hitherto described novel fungi from the forests of Western Ghats are listed below, with details on their substrate, locality and time of collection (Table 1).

Table-1. New hyphomycetes discovered from the forests of Western Ghats:

Fungus name	Substrate	Locality	Year of collection	Reference
<i>Acrodictys malabarica</i> Subram. & Bhat	Dead Bamboo	Silent Valley, Kerala	1980	Subramanian and Bhat, 1987
<i>Anavirga vermiformis</i> Bhat & Kendrick	Unidentified dead twigs	Kodachadri Hills, Karnataka	1991	Bhat and Kendrick, 1993
<i>Arthrinium mytilomorphum</i> Bhat et Kendrick	Dead leaves of <i>Andropogon</i> sp.	Kodachadri Hills, Karnataka	1991	Bhat and Kendrick, 1993
<i>Aquaphila ramdayata</i> Maria & Bhat	Leaves of <i>Caryota urens</i>	Cotigao, Goa	1999	D'Souza and Bhat, 2001
<i>Bahusutrabeeya dwaya</i> Subram. & Bhat	Decaying coffee twigs	Abby Falls, Karnataka	1976	Subramanian and Bhat, 1977
<i>B. dubhashii</i> Bhat	Leaf Litter of <i>Pandanus fascicularis</i>	Molem, Goa	1992	Bhat, 1994
<i>B. globosa</i> Bhat & Kendrick	Unidentified Dead Twig	Kudremukh, Karnataka	1989	Bhat and Kendrick, 1993
<i>B. manoharachari</i> Pratibha & Bhat	Leaves of <i>Bridelia scandens</i>	Chorla ghat, Goa	2004	Pratibha and Bhat, 2004
<i>Beltrania circinata</i> Bhat & Kendrick	Dead leaves of <i>Terminalia</i> sp.	Kodachadri Hills, Karnataka	1989	Bhat and Kendrick, 1993
<i>Benjpalia sundara</i> Subram. & Bhat	Dead petiole of <i>Calamus</i> sp.	Silent Valley, Kerala	1980	Subramanian and Bhat, 1987
<i>Bharatheeya goanensis</i> D'Souza & Bhat	Unidentified dead Leaves	Morpirla, Goa	1993	D'Souza and Bhat, 2002
<i>B. mucoides</i> D'Souza & Bhat	Dry leaves of <i>Calamus thwaitii</i>	Bondla, Goa	1999	D'Souza and Bhat, 2002
<i>Blastocatena pulneyensis</i> Subram. & Bhat	Unidentified decaying twigs	Kodaikanal, Tamil Nadu	1980	Subramanian and Bhat, 1987
<i>Brachysporiellina pulneyensis</i> Subram. & Bhat	Unidentified dead twig	Kodaikanal, Tamil Nadu	1981	Subramanian and Bhat, 1987
<i>Catenularia kalakadensis</i> Subram. & Bhat	Unidentified decaying twigs	Kalakad, Tamil Nadu	1980	Subramanian and Bhat, 1987
<i>C. malabarica</i> Subram. & Bhat	Unidentified dead twig	Silent vally, Kerala	1980	Subramanian and Bhat, 1987
<i>Ceevesubramaniomyces litseae</i> Pratibha, Hyde & Bhat	On leaves of <i>Litsea</i> sp.	Yana, Karnataka	2005	Pratibha and Bhat, 2004
<i>Cerasosporella pulneyensis</i> Subram. & Bhat	Unidentified dead twig	Kodaikanal, Tamil Nadu	1980	Subramanian and Bhat, 1987
<i>Chalara indica</i> Pratibha, Gawas, Hyde & Bhat	Live leaves of <i>Areca catechu</i>	Kesarval, Goa	2003	Pratibha and Bhat, 2005
<i>Cheiropolyschema ghaticum</i> Bhat & Kendrick	Dead decaying leaves	Edakumeri, Karnataka	1989	Bhat and Kendrick, 1993
<i>Craspedodidymum fimbriatum</i> Bhat & Kendrick	Unidentified dead twigs	Shiradi ghat, Karnataka	1988	Bhat and Kendrick, 1993
<i>Craspedodidymum hyalosporum</i> Bhat & Kendrick	Unidentified dead twigs	Edakumeri, Karnataka	1989	Bhat and Kendrick, 1993

Fungus name*	Substrate	Locality	Year of collection	Reference
<i>C.pulneyensis</i> Subram. & Bhat	Unidentified dead twig	Kodaikanal, Tamil Nadu	1980	Subramanian and Bhat, 1987
<i>Cryptophiale apicalis</i> Bhat & Kendrick	Dead leaves of <i>Vateria indica</i>	Shiradi ghat, Karnataka	1989	Bhat and Kendrick, 1993
<i>Cryptophiale verrucosa</i> Bhat & Kendrick	Dead leaves of <i>Vateria indica</i>	Sampaje ghat, Karnataka	1989	Bhat and Kendrick, 1993
<i>Dendrospora yessemreddyi</i> Nair & Bhat	Fresh water stream	Bondla, Goa	1999	Nair and Bhat, 2002
<i>Dictyochaeta ciliata</i> (Onofri & Rambelli) Bhat & Kendrick	Dead leaves of <i>Cinnamomum</i> sp.	Kudremukh, Karnataka	1989	Bhat and Kendrick, 1993
<i>Dichotomophthoropsis safeulaensis</i> Bhat, Rasheed & Sheeba	Dead leaves of <i>Oryza sativa</i> Konaje	Karnataka	1989	Sheeba et al., 1989
<i>Didymobotryum spirillum</i> D'Souza & Bhat	Dead culm of <i>Dendrocalamus strictus</i>	Molem, Goa	2000	D'Souza and Bhat, 2002
<i>Digitaranspora tambdisurlensis</i> Pratibha, Raghuk & Bhat	Unidentified Dead twig	Tambdi surla, Goa	2008	Pratibha et al., 2009
<i>Dimastigosporium yanense</i> Yadav & Bhat	Decomposed cattle dung	Yana, Karnataka	2008	Yadav and Bhat, 2009
<i>Diplospora indica</i> Nair & Bhat	Unidentified submerged litter	Cotigao, Goa	1999	Nair and Bhat, 2001
<i>Echinospaeria macrospora</i> Puja, Hyde & Bhat	Endophyte of <i>Centella asiatica</i>	Colem, Goa	2005	Puja et al., 2006
<i>Eptendis</i> Dharg. & Bhat	Endophyte of Rachis of <i>Pteris vittata</i>	Gersoppa, Karnataka	2007	Dhargalkar and Bhat, 2009
<i>Fusichalara goanensis</i> Bhat & Kendrick	Unidentified dead twig	Cotigao, Goa	1991	Bhat and Kendrick, 1993
<i>Gangliostilbe malabarica</i> Subram & Bhat	Unidentified dead twig	Silent valley, Kerala	1980	Subramanian and Bhat, 1987
<i>Gliocladium sampajense</i> Subram. & Bhat	Dead bark of <i>Theobroma Cocoa</i>	Sampaje, Karnataka	1979	Subramanian and Bhat, 1987
<i>Gonatobotryum binorphosporum</i> Jacob & Bhat	Endophyte of <i>Carrisa carandas</i>	Taleigao plateau, Goa	1998	Jacob and Bhat, 2000
<i>Helminthosporium belgaumense</i> Subram. & Bhat	Dead twig of unidentified plant	Upper Anmod, Goa	1980	Subramanian and Bhat, 1987
<i>Hyphopolynema stilboideum</i> Bhat & Kendrick	Dead leaves of <i>Syzigium</i> sp.	Kodachadri hills, Karnataka	1991	Bhat and Kendrick, 1993
<i>Idniella malabarica</i> Subram. & Bhat	Unidentified leaf litter	Silent Vally, Kerala	1980	Subramanian and Bhat, 1987
<i>Kumanasamuha kalakadensis</i> Subram. & Bhat	Unidentified dead twig	Kalakad, Tamil Nadu	1980	Subramanian and Bhat, 1987
<i>Kumbhamaya goanensis</i> Maria & Bhat	Dead leaves of <i>Floucourtia montana</i>	Cotigao, Goa	1999	Bhat and Kendrick, 1993
<i>K. jalapriya</i> Nair & Bhat	Live roots of <i>Hopea ponga</i>	Molem, Goa	2000	Nair and Bhat, 2002
<i>Kumbhamaya indica</i> Miriam & Bhat	Endophyte of <i>Carissa carandas</i>	Verna plateau, Goa	1997	Jacob and Bhat, 2000

Fungus name*	Substrate	Locality	Year of collection	Reference
<i>Microsporium appendiculatum</i> Miriam & Bhat	On goat dung	Verna, Goa	1996	Miriam and Bhat, 1998
<i>Natarajania indica</i> Pratibha & Bhat	Dead leaf of <i>Antiaris toxicaria</i>	Netravali, Goa	2006	Pratibha and Bhat, 2005
<i>Parahelminthosporium malabaricum</i> Subram. & Bhat	Dead Rachis of <i>Calamus sp.</i>	Oochipara, Silent vally, Kerala	1980	Subramanian and Bhat, 1987
<i>Phalangispora bharathensis</i> K.Prasad & Bhat	Leaf litter of <i>Holigama arotiana</i>	Cotigao, Goa	1999	Prasad and Bhat, 2002
<i>Phialosporostibe setosa</i> Bhat & Kendrick	On decaying leaves of <i>Ochlandra sp.</i>	Kudremukh, Karnataka	1989	Bhat and Kendrick, 1993
<i>Pleurothecium magnum</i> Subram. & Bhat	Unidentified rotten wood	Kodaikanal, Tamil Nadu	1981	Subramanian and Bhat, 1987
<i>P.pulneyense</i> Subram. & Bhat	Unidentified Dead twig	Kodaikanal, Tamil Nadu	1980	Subramanian and Bhat, 1987
<i>Piricaudiopsis appendiculata</i> Bhat & Kendrick	Unidentified dead twigs	Kudremukh, Karnataka	1989	Bhat and Kendrick, 1993
<i>Porosubramaniana malabarica</i> Subram. & Bhat	Unidentified Dead twig	Thekkadi, Kerala	1977	Subramanian and Bhat, 1987
<i>Pseudotonia helica</i> Subram. & Bhat	Unidentified Dead twig	Kodaikanal, Tamil Nadu	1981	Subramanian and Bhat, 1987
<i>Putagraivam sundaram</i> Subram. & Bhat	Dead twigs of <i>Macaranga indica</i>	Darbhe (Irde), Karnataka,		Subramanian and Bhat, 1977
<i>Rattania setuifera</i> Prabhugaonkar & Bhat,	Endophyte of <i>Calamus thwaitesii</i>	Dhoodhsagar, Goa	2008	Prabhugaonkar and Bhat, 2009
<i>Sorocybe indicus</i> Gawas, Hyde & Bhat	Hanging dead twig	Amboli, Maharashtra	2003	Pratibha <i>et al.</i> , 2005
<i>Spagazzinia subramaninii</i> Bhat	Hyperparasite on <i>Helminthosporium velutinum</i>	Kodachadri hills, Karnataka	1989	Bhat, 1994
<i>Speiopsis rogergoosensis</i> K.Prasad & Bhat	Dead leaves of <i>Artocarpus hirsutus</i> Lam.	Subramanya, Karnataka	2001	Prasad <i>et al.</i> , 2002
<i>Spondylocladiopsis aseptata</i> Pratibha, Raghuk. & Bhat	Unidentified leaf litter	Calicut, Kerala	2008	Pratibha and Bhat, 2009
<i>Sporidesmina malabarica</i> Subram & Bhat	Dead petiole of <i>Curcuma sp.</i>	Silent Vally, Kerala	1980	Subramanian and Bhat, 1987
<i>Sporidesmiopsis malabarica</i> Subram & Bhat	Dead petiole of <i>Curcuma sp.</i>	Silent Vally, Kerala	1980	Subramanian and Bhat, 1987
<i>S. gounensis</i> Bhat & Kendrick	Unidentified dead twigs	Kodachadri hills, Karnataka	1991	Bhat and Kendrick, 1993
<i>Sporoschisma uniseptatum</i> Bhat & Kendrick	Unidentified decaying twig	Morphirla, Goa	1991	Bhat and Kendrick, 1993
<i>Stellomyces kendrickii</i> K.Prasad & Bhat	Decaying twigs of <i>Hopea pongu</i>	Molem, Goa	2001	KeshavPrasad and Bhat, 2002
<i>Tetraploa circinata</i> Pratibha & Bhat	Dead bamboo twig	Mahabaleshwar, Maharashtra	2006	Pratibha and Bhat, 2008
<i>Trichobotrys ramosa</i> D'Souza & Bhat	Dead leaves of <i>Dendrocalanus strictus</i>	Molem, Goa	1999	Maria and Bhat, 2001

Fungus name*	Substrate	Locality	Year of collection	Reference
<i>Trinacnum indica</i> Soosamma, Lekha, Sreekala & Bhat	Decaying leaf litter of <i>Coffea arabica</i>	Somvarpet, Karnataka	1998	Soosamma <i>et al.</i> , 2001
<i>Uberispora tropicalis</i> Bhat & Kendrick	Unidentified dead twigs	Donigalu, Karnataka	1989	Bhat and Kendrick, 1993
<i>Vamsapriya indica</i> Gawas & Bhat	Dead bamboo twig	Yellapur, Karnataka	2005	Gawas and Bhat, 2005
<i>V. mahabaleshwariensis</i> Pratibha & Bhat	Unidentified dead twig	Mahabaleshwar, Maharashtra	2006	Pratibha and Bhat, 2008
<i>Vanakripa gigaspora</i> Bhat, Kendrick & Nag Raj	Unidentified dead twigs	Molem, Goa	1991	Bhat and Kendrick, 1993
<i>V. parva</i> Bhat et Kendrick & Nag Raj	Unidentified dead twigs	Cotigao, Goa	1991	Bhat and Kendrick, 1993
<i>Vermiculariopsiella elegans</i> K Prasad, D'Souza & Bhat	Leaf litter of <i>Saraca asoka</i>	Bondla, Goa	1999	Prasad <i>et al.</i> , 2003
<i>V. endophytica</i> Puja, Shenoy, Hyde & Bhat	Endophyte of <i>Centella asiatica</i>	Colem, Goa	2005	Puja <i>et al.</i> , 2006
<i>V. indica</i> K.Prasad, D'Souza & Bhat	Endophyte of <i>Saraca asoka</i>	Bondla, Goa	1999	Prasad <i>et al.</i> , 2003
<i>V. pteridis</i> Dharg. & Bhat	Endophyte of rachis of <i>Pteris vittata</i>	Gersoppa, Karnataka	2007	Dhargalkar and Bhat, 2009
<i>Vermisporu cauvenana</i> Rajashekar, Bhat & Kaveriappa	Dead leaves of <i>Ficus religiosa</i>	Srirangapatana, Karnataka	1991	Rajashekar <i>et al.</i> , 1991
<i>Xenoheteroconium bicolor</i> Bhat, Kendrick & Nag Raj	Unidentified decaying leaves	Agumbe, Karnataka	1989	Bhat and Kendrick, 1993
<i>Zygosporium tuberculatum</i> Subram. & Bhat	Unidentified dead twig	Silent Vally, Kerala	1980	Subramanian and Bhat, 1987

*New genera are in bold

Establishment of a fungus culture repository in Goa University

Professor Subramanian had always advocated culturing and maintenance of fungi in the repositories. I did numerous single-spore isolations of hypocrealean and hyphomycetous fungi, while in Madras, for the Fungus Flora project. Along with my students and colleagues, I continued the exercise in Goa University. While dried herbarium specimens satisfy mandatory taxonomic obligations of description of novel species, culturing and maintaining the fungi in repositories is a potentially optimistic approach of not only slowing down the loss of bioresources from our forests but also a meaningful way of organizing sustainable and utilizable fungal gene banks. We set up an in-house such facility for fungi in the Department of Botany, Goa University, and I had Professor Subramanian to switch on the programme, in 1998.

Evolution of Hyphomycete taxonomy

Fries (1821-1832) was considered as the starting point for most fungi. For conidial fungi or

Deuteromycotina which embodied the Hyphomycetes and Coelomycetes, Persoon (1801) was taken as the starting point. The International Code of Botanical Nomenclature (ICBN) provides rules and recommendations with regard to application of names at different taxonomic hierarchy in fungi. Presently, St. Louis Code published in 2000 is in use. According to Article 59.1. of the Code, for ascomycete and basidiomycete fungi with asexual and sexual morphs, the name of the holomorph (species with all its morphs), typified by the teleomorph, is the earliest legitimate name to be applied.

In the treatment of Hawksworth *et al.* (1983), Eumycota included five subdivisions: Mastigomycotina (Chytridiomycetes, Hyphochytriomycetes, Oomycetes), Zygomycotina (Zygomycetes, Trichomycetes), Ascomycotina, Basidiomycotina and Deuteromycotina. Kirk *et al.* (2001) accommodated 'true fungi' in 4 phyla, viz. Chytridiomycota, Zygomycota, Ascomycota and Basidiomycota. The Deuteromycotina is not accepted as a formal taxonomic group as they are polyphyletic (Seifert *et*

al., 2000; Shenoy *et al.*, 2006, 2007). Recently, Hibbett *et al.* (2007) proposed a higher-level phylogenetic classification for fungi based on morphology, molecular biology and phylogenetic relationships which did not treat the sexual and asexual morphs separately.

Notwithstanding what is said above, conidial fungi are rarely encountered in their full form, as holomorph, in nature. Either they exist in their sexual phase (teleomorph) which reproduce by meiospores resulted from sexually differentiated cells as in Ascomycotina or Basidiomycotina or in asexual forms (anamorph) which are propagated by mitospores, as in Deuteromycotina. In nature, sexual and asexual phases of fungi are often separated in space and time; that is, separate entities of anamorphic and teleomorphic fungi exist. More often, an anamorph remains unconnected with a teleomorph, or the latter may be totally absent. Since, the teleomorph holds nomenclatural priority over anamorphic phase and as and when more and more anamorph-teleomorph connections are established, provisions are made in the Code to accommodate conidial fungi with binomials as 'form-genera' and 'form-species', that is 'anamorph-genera' and 'anamorph-species'.

Though all anamorphs are expected to get connected to their teleomorphs, sexual phase of a number of conidial fungi are not yet known. Absence of sexual phase does not imply that the fungus lacks sex. If the fungus is totally asexual, it wouldn't get the advantages of sexual reproduction such as generation of novel genotypes capable of living in changing environments, withstanding the deleterious effects of mutations, escaping from pathogens, etc. It is possible that substrate- and habitat-wise, seasonal and geographic separation of compatible mating types might be limiting the sexual reproduction in fungi (O'Gorman *et al.*, 2008). It is also intriguing that 22% of the reported fungi do not have sexual morphs (Shenoy *et al.*, 2007). This includes many of the industrially important strains and human pathogenic fungi. Though 'anamorphy' will continue to remain as an independent entity in the taxonomy of fungi at least for some time, in the future we will have a single system of classification of fungi based on DNA-sequence data supplemented by other available biological data.

Future of taxonomy of Hyphomycetes

Molecular genetic data of fungi provide value addition in taxonomic studies. Advanced methods applied and data generated serve as molecular signatures which on comparison with similar data from other isolate/s facilitate confirmation of relatedness of

the two. Sequencing of the ITS, 18S and 28S rRNA gene regions for fungi is employed to distinguish isolates at generic and species level and to study their phylogenetic affinities and relatedness (Hibbett *et al.*, 2007). Major drawback in general application of this technology, however, is the high cost of instrumentation, chemicals and lab-wares, especially in the fungi-rich developing countries such as ours.

Nevertheless, there are renewed efforts made to unravel the mysteries associated with refusal of asexual fungi to reproduce sexually. O'Gorman *et al.* (2008) studied the mating behaviour and sexuality of genetically variable 12 isolates of an opportunistic human pathogen, *Aspergillus fumigatus*, in a range of media and temperature regimes. In one of these crosses, after 6 months of incubation, they discovered fruiting bodies characteristic of a sexual phase of an aspergillus, *Neosartorya fumigata*. Examination of segregation of five genetic markers in ascospore progeny from different crosses confirmed that meiotic recombination did take place. The discovery of sexual cycle in this species offers not only a useful tool for investigations into classical genetic analyses and studies on genetic basis of pathogenicity and fungicidal resistance in *A. fumigatus* but also opens up avenues to improve the methods of control of aspergillosis. These results propose insights into the potential of sexual reproduction in similar supposedly 'asexual' fungi.

Sex determination in fungi is controlled by a small, specialized region of the genome. Different gene combinations reside at these mating-type (MAT) loci and confer sexual identity. The MAT loci have been characterized for a single monophyletic clade of fungi, the Dikarya (Ascomycota and Basidiomycota), but the ancestral state and evolutionary history of these loci have so far remained a mystery. Applying bioinformatic and genetic mapping tools to identify the sex-determining region in *Phycomyces blakesleeanus* (Zygomycota), which represents an early branch within the fungi, Idnurm *et al.* (2008) found out that each sex allele contains a single gene that encodes for a high mobility group (HMG)-domain, implicating that the HMG-domain proteins are an earlier form of fungal MAT loci.

Eighty years after Alexander Fleming's discovery of penicillin by a strain of *Penicillium notatum*, Hoff *et al.* (2008) discovered another strain of *Penicillium* with hyper antibacterial activity. Using a heterologous PCR approach, they showed that these strains are of opposite mating types and that both have retained transcriptionally expressed pheromone and pheromone receptor genes required for sexual

reproduction. This discovery gives us a clue why so many of the fungi still remain as asexual.

Epilogue

I believe that the basics lie, at least for the present, in the morphological diagnoses of the fungi. Every type or reference taxon needs to be correctly identified morphologically before its sequence forms a reference for comparison with other isolates. Morphology-based identification is a quick, pragmatic and inexpensive method for diagnosis of the fungi. Often, these are found to be phylogenetically natural (Seifert and Gams, 2001). I appeal fellow mycologists, young and elder, to simultaneously venture on morpho- and molecular-based systematics of fungi. This will provide a stable single classification and reliable identification system for fungi.

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Fig. 1: Extracted from Google Earth™ mapping service