A Thesis Entitled

SYNTHETIC STUDIES IN OXYGEN HETEROCYCLES

Submitted to Goa University for the Award of the Degree of

DOCTOR OF PHILOSOPHY

In

CHEMISTRY

By

Ms. MAYURI MADHUKER NAIK

M. Sc.

Under the Guidance of

PROF. VIJAYENDRA P. KAMAT

Department of Chemistry

and

PROF. SANTOSH G. TILVE

Department of Chemistry

GOA UNIVERSITY

Taleigao Plateau, Goa 403 206

INDIA

APRIL 2018

A Thesis Entitled

SYNTHETIC STUDIES IN OXYGEN HETEROCYCLES

Submitted to Goa University for the Award of the Degree of

DOCTOR OF PHILOSOPHY

In

CHEMISTRY

By

Ms. MAYURI MADHUKER NAIK

M. Sc.

Under the Guidance of

PROF. VIJAYENDRA P. KAMAT

Department of Chemistry

and

PROF. SANTOSH G. TILVE

Department of Chemistry

GOA UNIVERSITY

Taleigao Plateau, Goa 403 206

INDIA

APRIL 2018

DEPARTMENT OF CHEMISTRY

CERTIFICATE

This is to certify that the thesis entitled, "**Synthetic Studies in Oxygen Heterocycles**" submitted by Ms. **MAYURI MADHUKER NAIK**, is a record of research work carried out by the candidate during the period of study under our supervision and that it has not previously formed the basis for the award of any degree or diploma or other similar titles.

Goa University April 2018 **Prof. Vijayendra P. Kamat** Research Guide Department of Chemistry Goa University **Prof. Santosh G. Tilve** Research Guide Department of Chemistry Goa University

DECLARATION

I hereby declare that the work embodied in the thesis entitled "**Synthetic Studies in Oxygen Heterocycles**" is the result of investigations carried out by me under the guidance of **PROF. VIJAYENDRA P. KAMAT** and **PROF. SANTOSH G. TILVE** at Department of Chemistry, Goa University and that it has not previously formed the basis for the award of any degree or diploma or other similar titles.

In keeping with the general practice of reporting scientific observations, due acknowledgement has been made wherever the work described is based on the findings of other investigators.

Goa University April 2018 Ms. Mayuri M. Naik Ph. D. Student Department of Chemistry Goa University

ACKNOWLEDGEMENT

Ph. D. is a degree for which to achieve one has to go through several barriers in life. Also it cannot be complete without the direct or indirect help and support from various people. So, at the final stage of my Ph. D. course I would like to take the opportunity to thank all the people who helped me at different stages and without whom it would have not been possible to complete the thesis.

At the outset I would like to express my deep sense of gratitude to both my supervisors **Prof. V. P. Kamat** (former registrar, Goa University) and **Prof. S. G. Tilve** (former Head, Department of Chemistry, Goa University) for considering me and giving an opportunity to work as a research student under their valuable guidance. Their constant support, encouragement and useful suggestions helped me to work in right direction and inculcated broad research thinking. I really appreciate their immense knowledge, patience and the freedom given to me to do research work as well as to write research articles which helped me to develop myself throughout this period. I will be always grateful to them.

The role played by **Prof. S. G. Tilve** in my research career cannot be expressed in words. He has always been a supportive guide through his valuable guidance, suggestions, motivation and unique ideas throughout the Ph. D. course.

I would like to sincerely thank **Prof. Varun Sahni** (Vice-chancellor, Goa University), **Prof. Y. V. Reddy** (Registrar, Goa University), **Prof. S. Shetye** (former Vice-chancellor, Goa University) and **Prof. V. P. Kamat** (former Registrar, Goa University) for their valuable support by allowing me to carry out my research work in this institute.

I would also like to express my kind gratitude to **Prof. B. R. Srinivasan** (Head, Department of Chemistry, Goa University), **Prof. Gourish M. Naik** (Dean, Natural Sciences, Goa University), **Prof. J. A. E. Desa** and **Prof. A. V. Salkar** (Former Deans) for providing necessary facilities for my research work.

I sincerely acknowledge my subject experts, **Dr. Jayant Umarye** (Head, R & D, Godrej Agrovet Ltd., Mumbai & former group leader, Syngenta Biosciences Pvt. Ltd., Goa) and **Dr. Sandesh Bugde** (Assistant Professor in Chemistry, Parvatibai Chowgule college of Arts and Science) for providing the much needed insights into my subject. I also acknowledge **Prof. V. S. Nadkarni** and **Prof. B. R. Srinivasan** for their expert advice whenever required.

I would like to deeply acknowledge **Human Resource Development Group-Council of Scientific and Industrial Research (HRDG-CSIR)**, New Delhi for providing me financial assistance through NET - Junior and Senior Research Fellowships for 5 years.

I express my sincere gratitude towards all my teachers **Prof. K. S. Rane, Prof. J. B.** Fernandes, Prof. S. P. Kamat, Prof. S. G. Tilve, Prof. V. P. Kamat, Prof. A. V. Salkar, Prof. B. R. Srinivasan, Prof. V. S. Nadkarni, Dr. V. M. S. Verenkar, Dr. R. N. Shirsat,
Dr. S. N. Dhuri and Mrs. Siddhali Rajadhyaksha for their valuable teaching and motivational talks from which I could learn a lot. Also, I thank the present faculty members
Dr. Mahesh Majik, Dr. Kanchanmala, Dr. Kashinath, Dr. Pranay, Dr. Bhanudas, Dr.
Prachi and Dr. Sonia for their help and goodwill. I also thank former faculty Dr. Gururaja for his helpful discussions on NMR spectra. I also thank Dr. J. K. Kirtany and Dr. Lisette
Dsouza for their valuable suggestions during my Ph. D. course. I also express thanks to my
B. Sc. Chemistry teachers Dr. Beena Vernekar and Mrs. Jaqueline Fernandes for their teaching method, encouragement and developing interest in chemistry.

I am also thankful to **Dr. Gopakumar** (Librarian, Goa University) for his assistance in the plagiarism check of thesis. I also acknowledge the **staff members of Library** and **administrative members** Goa University for being kind and helpful. I also thank all the **non-teaching staff**, Department of Chemistry, Goa University for helping at numerous times.

I am thankful to **Prof. Sanjeev Ghadi** (Department of Biotechnology, Goa University) and **Dr. Surya Nandan Meena** for performing anti-diabetic studies of my samples.

I acknowledge Indian Institute of Science (IISc), Bangalore and Syngenta Biosciences Pvt., Ltd., Goa for providing the HRMS facility. I also thank Sophisticated Analytical Instrument Facility-Indian Institute of Technology (SAIF-IIT), Bombay for providing ICP-MS data.

I immensely thank my seniors **Dr. Kashinath**, **Dr. Sandesh** and **Dr. Prachi** for helping me and providing tremendous support in all the possible ways at difficult times during this course. I also appreciate their funny acts, joyful moments, fruitful research discussions and motivational talks which helped me to enjoy as well as boost up my research work. I also express my sincere thanks to all my group members **Dr. Mahesh**, **Dr. Rupesh**, **Dr. Prakash**, **Dr. Reshma**, **Dr. Sonia**, **Dr. Prachi**, **Dr. Chinmay**, **Dr. Hari**, **Dr. Sagar**, **Dr. Prajesh**, **Durga** and **Pratibha** for their constant help, encouragement, valuable discussions, suggestions and enjoyable moments. I cannot express in words the knowledge I gained and things I learnt from my group members for which I will always be grateful to them. I also acknowledge my junior group members **Ketan**, **Shashank** and **Lima** for pleasing working environment and **Dr. Sumit** for fruitful discussion.

I thank **Dr. Santosh Shetgaonkar**, **Dr. Mahesh Majik**, **Dr. Amit Vernekar**, **Dipesh Harmalkar** and **Dr. Siddhi** for providing the required references. Also, I am thankful to **Dr. Santosh** for helping me in providing LCMS data of some of my samples. I also offer my thanks to **Dr. Chinmay**, **Dr. Kashinath**, **Dr. Sandesh**, **Dr. Hari**, **Dr. Prajesh** and **Dr. Sagar** for providing great help by recording NMR spectra of my samples in the beginning of my

research work. I am greatful to **Dr. Kashinath**, **Dr. Prachi**, **Durga**, **Dr. Sandesh** and **Ashwini** for helping me in editing my thesis.

I would like to extend my thanks to all my senior and present colleagues **Dr. Vidhya**, **Dr. Nitya**, **Dr. Rajesh**, **Dr. Shrikant**, **Dr. Lactina**, **Dr. Vinod**, **Dr. Priyanka**, **Dr. Savia**, **Dr. Sulaksha**, **Dr. Jose**, **Dr. Satish**, **Dr. Rohan**, **Dr. Umesh**, **Dr. Kiran**, **Dr. Mithil**, **Rita**, **Savita**, **Dr. Diptesh**, **Dattaprasad**, **Satu**, **Shambhu**, **Dr. Madhavi**, **Mira**, **Kedar**, **Visha**, **Abhijeet**, **Daniel**, **Dr. Celia**, **Pratik**, **Apurva**, **Prajyoti**, **Chandan**, **Sarvesh**, **Madhavi**, **Johnross**, **Vishnu**, **Sudarshana**, **Pooja**, **Neha**, **Sudesh**, **Rahul** and **Amarja** for making days enjoyable at university. Also, I thank my friends **Roma**, **Samrudhi**, **Belinda**, **Sejal**, **Bindiya**, **Harshada**, **Pratiksha**, **Poonam**, **Nitesh**, **Madhukar**, **Manjunath**, **Dr. Dinesh**, **Anupa**, **Palmira**, **Shilpa**, **Sonam**, **Neclin**, **Mahesh**, **Santosh**, **Dr. Kiran** and **Dr. Pradnyesh** for their warm friendship.

I am also very thankful to **Dr. V. J. Pissurlekar** (The principal, P. E. S's. R. S. N. college of arts and science), **Dr. Anita S. Tilve** (M. Sc. co-ordinator) and all my colleagues of chemistry department for their support during my working period.

Also I am very much thankful to **Dr. Nandkumar N. Sawant** (The principal, Parvatibai Chowgule college of arts and science) and all my senior colleagues of chemistry department from the presently working college for their help and support.

I am deeply thankful to my father late **Mr. Madhuker M. Naik** for raising me with love, affection, good moral values and constant encouragement for higher studies. Also my mother **Mrs. Madhavi M. Naik** enabled me to study further by her extreme hard work, motivation, freedom and I thank her for always being a pillar of support. I am at loss of words to express their love and support offered to me for which I will be always grateful to them. Without them I would not have been able to reach at this stage of my career. The funny acts of my elder brother **Mandar** helped me to overcome the tensions and he also encouraged to work hard with his kind words. Also I thank my other **family members** from the bottom of my heart for constantly encouraging me and providing love, care and support throughout my life. I also thank my best friend **Suhel Narvekar** for understanding me and standing by my side during my ups and downs which motivated me all the time.

Last but not the least, I thank Almighty for giving me good health, strength and support to overcome all the barriers in my life and become a good human being by continuously showering his kind blessings.

Ms. Mayuri M. Naik

Dedicated

То Му

Beloved Parents

TABLE OF CONTENT

General Remarks	i
Abbreviations	ii
Abstract of thesis	V
Publications and conferences	viii & ix

	A short review on the recent advancements in the				
Chapter 1	synthesis of heterocycles using molecular iodine as a	1-21			
	catalyst or reagent				
1.1	Introduction	1			
1.2.1	Iodine as a sole catalyst	2			
1.2.2	Iodine as a sole reagent	12			
1.3	Conclusion	19			
1.4	References	19			
Chanter 2	Synthetic studies of chromans using molecular iodine	22-85			
	catalyst	00			
2.1	Introduction	22			
2.2	Occurrence	22			
2.3	Literature synthetic methods	29			
2.4	Results and Discussion	49			
2.5	Conclusion	61			
2.6	Experimental	62			
2.7	References	63			
	NMR spectra	70			
	Synthetic studies of flavones using pyrrolidine and				
Chapter 3	molecular iodine catalysts and their anti-diabetic	86-171			
	activity				
3.1	Introduction	86			
3.2	Occurrence	86			
3.3	Literature synthetic methods	89			

3.4	Results and Discussion	112
3.5	Anti-diabetic activity	131
3.6	Conclusion	137
3.7	Experimental	138
3.8	References	140
	NMR spectra	150
Chapter 4	Synthetic studies of coumestans using Cu(OAc) ₂	172-318
4.1	Introduction	172
4.2	Occurrence	172
4.3	Literature synthetic methods	185
4.4	Results and Discussion	213
	Results and Discussion	215
4.5	Conclusion	252
4.5 4.6	Conclusion Experimental	213 252 252
4.5 4.6 4.7	Conclusion Experimental References	252 252 257

GENERAL REMARKS

1) The compound numbers, figure numbers, scheme numbers and reference numbers given in each chapter refer to that particular chapter only.

2) All melting points and boiling points were recorded using Thiele's tube and are uncorrected.

3) Commercial reagents were used without further purification.

4) All solvents were distilled prior to use and then dried using standard procedure.

5) All reagents were prepared using literature methods.

6) Chromatographic purification was conducted by column chromatography using silica gel (60 - 120 mesh size) or by flash chromatography using silica gel (200-400 mesh size).

7) Thin layer chromatography (TLC) was carried out on silica gel 60 F254 aluminium plates purchased from Merck.

8) The IR spectra were recorded on Shimadzu FT-IR spectrophotometer.

9) ¹H NMR (400 MHz) and ¹³C NMR (100 MHz) spectra were recorded on a Brucker AVANCE 400 instrument and the multiplicities of carbon signals were obtained from DEPT experiment. Chemical shifts are expressed in δ relative to tetramethylsilane (TMS) which is expressed in ppm.

10) The high resolution mass spectra (HRMS) were recorded on MicroMass ES-QTOF mass spectrometer.

11) The absorbance values were obtained from UV-Vis spectrophotometer (Shimadzu-1800) at 517 nm.

DEFINITION OF ABBREVIATIONS

g	Gram/s	ee	Enantiomeric excess	
mg	Milligram/s	dr	Diastereomeric ratio	
μg	Microgram/s	conc.	Concentrated	
mol	Mole/s	aq.	Aqueous	
mmol	Millimole/s	0	Ortho	
mL	Milliliter/s	т	Meta	
mm	Millimeter/s	р	Para	
nm	Nanometre/s	MS	Molecular sieves	
m.p.	Melting point	psi	Pounds per square inch	
b.p.	Boiling point	cat.	Catalytic	
ev	Electron volt	atm.	Atmospheric	
lit.	Literature	et al.	Et alia (and others)	
d	Day/s	TLC	Thin layer chromatography	
h	Hour/s	sat.	Saturated	
min	Minute/s	MW	Microwave	
sec	Second/s	anhyd.	Anhydrous	
μΜ	Micromolar	°C	Degree Celcius	
nM	Nanomolar	rt	Room temperature	
ppm	Parts per million	Ζ	Zussamen (together)	
hv	Irradiation	Ε	Eentegegen (opposite)	
%	Percentage	equiv	Equivalent	
R	Rectus	wt./wt.	Weight per unit weight	
S	Sinister	Å	Ångström	

General Abbreviations

Compound Abbreviations

Ac	Acetyl	dppp	1,3-Bis(diphenylphosphino)propane
acac	Acetylacetone	DTDB	Di-tert-butyl peroxide
AcOH	Acetic acid	EtOAc	Ethyl acetate
Ac ₂ O	Acetic anhydride	EtOH	Ethanol
AgOTf	Silver	Et ₂ O	Diethyl ether
	trifluoromethanesulfonate		
AIBN	2,2'-Azobisisobutyronitrile	Et ₃ N	Triethylamine
BF ₃ .OEt ₂	Boron trifluoride diethyl etherate	Fe(OTf) ₃	Iron(III) trifluoromethanesulfonate
binap	(2.2'-	Ga(OTf) ₃	Gadolinium(III)
1	Bis(diphenylphosphino)-	()5	trifluoromethanesulfonate
	1,1'-binaphthyl)		
Bi(OTf) ₃	Bismuth(III)	IBX	2-Iodoxybenzoic acid

bipy 2,2'-Bipyridine In(OTf)3 Indium(III) trifluoromethanesulfonate BMIM 1-Butyl-3- methyimidazolium KOr-Bu Potassium <i>i</i> -butoxide Bn Benzyl LDA Lithium diisopropylanide BnBr Benzyl bromide LTA Lead tetraacaetate Boc <i>i</i> -Butyloxycarboayl McNO2 Nitromethane <i>n</i> -BuBu normal (primary) Butyl MOM Methoxymethyl ether <i>n</i> -BuLi <i>n</i> -Butyl lithium NBS <i>N</i> -Bromosuccinimide <i>i</i> -BuLi <i>n</i> -Butyl lithium NIS <i>N</i> -Chlorosuccinimide <i>i</i> -BuOH <i>i</i> -Butyl alcohol NMP <i>N</i> -Methyl-2-pyrrolidone <i>i</i> -BuOK Potassium tertiary butoxide PCC Pyidinium chlorochromate KTBT Potassium tertiary butoxide PCC Pyidinium chlorochromate COD Cycloocta-1,5-diene Phen/1,10- 1,10-Phenanthroline <i>m</i> -CrBA <i>m</i> -Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OAc) ₂ Copper(II) PIFA Polyphosphino-2',6'- Cu(COC) ₂ Copper(I) hiophene-2- PA Polyphosphinic Cu Copper(I) thiophene-2- PA Polyphosphinic acid Cy Cyclohexyl cabodiimide TBAC Tetra-tutyl ammonium chlo		trifluoromethanesulfonate			
Brite of generalization Brite of generalization BMIM 1-Butyl-3- methyimidazolium For Bu Potassium <i>t</i> -butoxide Bn Benzyl LDA Lithium diisopropylamide BaBr Benzyl bromide LTA Lead tetraacaetate Boc <i>t</i> -Butyloxycarbonyl McNO Nitromethane <i>n</i> =BuW normal (primary) Butyl MOM Methoxymethyl ether <i>n</i> =BuU <i>n</i> =Butyl lithium NBS <i>N</i> =Bronosuccinimide <i>r</i> =Butyl <i>r</i> -Butyl lithium NIS <i>N</i> -Iodosuccinimide <i>r</i> =Butyl <i>r</i> -Butyl lithium NIS <i>N</i> -Iodosuccinimide <i>r</i> =Butyl <i>r</i> -Butyl lithium NIS <i>N</i> -Iodosuccinimide <i>r</i> =Butyl r-Butyl lithium NIS <i>N</i> -Iodosuccinimide <i>r</i> =Butyl <i>r</i> -Butyl lithium NIS <i>N</i> -Iodosuccinimide <i>r</i> =Butyl relation NCC Pyridine <i>r</i> -Butyl <i>r</i> =Butyl Rolyphenzition Pityl Pitylonion <i>r</i> -Butylon <i>c</i> -Butylon Polopertylonion <i>c</i> -Core <i>c</i> -Diocolon	bipy	2.2'-Bipyridine	In(OTf) ₃	Indium(III)	
BMIM 1-Butyl-3- methylimidazolium KOr-Bu Potassium t-butoxide Bn Benzyl LDA Lithium diisopropylamide BnBr Benzyl bromide LTA Lead tetraacaetate Boc t-Butyloxycarbonyl MeNO2 Nitromethane n-Bu/Bu normal (primary) Butyl MOM Methoxymethyl ether n-Bu/Li n-Butyl lithium NBS N-Bromosuccinimide r-Bu/Li t-Butyl lithium NIS N-Iodosuccinimide r-BuU t-Butyl lithium NIS N-Iodosuccinimide r-BuOK Potassium tertiary butoxide PCC Pyridinium chlorochromate KTBT Potassium tertiary butoxide Pd/C Palladium on activated charcoal COD Cyclocta-1,5-diene Pher/1,10- 1,10-Phenanthroline me-Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OAc)2 Copper(II) rtifuoromethanesulfonate e) Cu(OTc) Copper(I) thophene-2- carboxylate Cy Cyclohexyl abodiimide TBA Phenylodine(III)bis(trifluoroacetat e)	- 15		(- 75	trifluoromethanesulfonate	
methylimidazolium methylimidazolium Bn Benzyl LDA Lithium diisopropylamide BnBr Benzyl bromide LTA Lead tetraacaetate Boc <i>t</i> -Butyloxycarbonyl McNO ₂ Nitromethane n-Bu/Li n-Bu/Li <i>n</i> -Bu/Li <i>n</i> -Butyl lithium NBS <i>N</i> -Bromosuccinimide <i>t</i> -Bu <i>t t</i> -Butyl lithium NIS <i>N</i> -Chlorosuccinimide <i>n</i> -Bu/Li <i>t</i> -Bu/Li <i>t</i> -Butyl lithium NIS <i>N</i> -Iodosuccinimide <i>n</i> -Bu/Li <i>t</i> -Bu/Li <i>t</i> -Butyl lithium NIS <i>N</i> -Methyl-2-pyrrolidone <i>P</i> -Bu/Li <i>t</i> -BuOK / Potassium tertiary butoxide PCC Pyridinium chlorochromate <i>P</i> -Bu/Li <i>t</i> -BuOK / Potassium tertiary butoxide Pd/C Palladium on activated charcoal COD Cycloocta-1,5-diene Phen/LiO- phen/LiO- phen/LiO- <i>m</i> -CHoroperbenzoic acid PhyO Diphenyl ether Cu(OAc)2 Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2', 6'- CuTC Copper(I) phenen- Phenyliodime(III)bis(BMIM	1-Buty1-3-	KOt-Bu	Potassium <i>t</i> -butoxide	
Bn Benzyl LDA Lithium diisopropylamide BnBr Benzyl bromide LTA Lead tetraacaetate Boc <i>t</i> -Butyloxycarbonyl MeNO2 Nitromethane <i>n</i> -Bu/Bu normal (primary) Butyl MOM Methoxymethyl ether <i>n</i> -BuLi <i>n</i> -Butyl lithium NBS <i>N</i> -Bromosuccinimide <i>t</i> -Butyl <i>t</i> -Butyl lithium NIS <i>N</i> -Chlorosuccinimide <i>t</i> -BuOK <i>t</i> -Butyl lithium NIS <i>N</i> -Othorosuccinimide <i>t</i> -BuOK Potassium tertiary butoxide PCC Pyridinium chlorochromate <i>t</i> -BuOK Potassium tertiary butoxide PCC Pyridinium chlorochromate <i>t</i> -BuOK Potassium tertiary butoxide PCC Pyridinium chlorochromate <i>CH</i> 3SO3H Methane sulfonic acid PM/C Palladium on activated charcoal COD Cyclocat-1,5-diene Phe/D 1,10-Phenanthroline <i>m</i> -Chloroperbenzoic acid PhyO Diphenyl ether Cu(OTf)2 Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2*,6*- CuT Copper(I) thiophene-2-		methyimidazolium			
BnBr Benzyl bromide LTA Lead tetraacaetate Boc t-Butyloxycarbonyl MeNO2 Nitromethane n-Bu/Bu normal (primary) Butyl MOM Methoxymethyl ether n-BuLi n-Butyl lithium NBS N-Bromosuccinimide t-BuLi t-Butyl lithium NIS N-Chorosuccinimide t-BuCi t-Butyl lithium NIS N-Chorosuccinimide t-BuCK Potassium tertiary butoxide PCC Pyridinium chlorochromate TBUCH Potassium tertiary butoxide PCC Pallalium on activated charcoal COD Cycloocta-1,5-diene Phen/1,10- 1,10-Phenanthroline m-Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OTc)2 Copper(II) copper(I) PhayO Diphenyl ether Cu(OTf)2 Copper(I) thiphene-2- carboxylate PPA Polyphosphoric acid Cy Cyclohexyl PPh3 Triphenylphosphine DD DBU 1,8- Dichlorobenzene Sc(OTf)3 Scandium(III) trifluoromethanesulfonate	Bn	Benzyl	LDA	Lithium diisopropylamide	
Boc <i>I</i> -Butyloxycarbonyl MeNO2 Nitromethane <i>n</i> -Bu/Bu normal (primary) Butyl MOM Methoxymethyl ether <i>n</i> -BuLi <i>n</i> -Butyl lithium NBS <i>N</i> -Bromosuccinimide <i>t</i> -Bu <i>t</i> -Butyl lithium NIS <i>N</i> -Chlorosuccinimide <i>t</i> -BuOK <i>t</i> -Butyl alcohol NMP <i>N</i> -Methyl-2-pyrrolidone <i>t</i> -BuOK / Potassium tertiary butoxide PCC Pyridinium chlorochromate <i>t</i> -BuOK / Potassium tertiary butoxide PCC Palladium on activated charcoal COD Cycloocta-1,5-diene Phen/1,10- 1,10-Phenanthroline <i>m</i> -CPBA <i>m</i> -Chloroperbenzoic acid Ph2O Diphenyl ether Cu(OAc)2 Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2',6'- CuTC Copper(I) thophene-2- PPA Polyphosphoric acid e) CuTC Copper(I) thophene-2- PPA Polyphosphoric acid e) DBU 1,8- Dicklorobenzene Sc(OTf)3 Scandium(III) trifluoromethanesulfonate DCC Dicyclohexylcabodiimide	BnBr	Benzyl bromide	LTA	Lead tetraacaetate	
n-Bu/Bu normal (primary) Butyl MOM Methoxymethyl ether n-Bu/Li n-Butyl lithium NBS N-Bromosuccinimide t-Bu t-Butyl NCS N-Chlorosuccinimide t-Bu t-Butyl lithium NIS N-Iodosuccinimide t-BuOH t-Butyl alcohol NMP N-Methyl-2-pyrrolidone t-BuOK / Potassium tertiary butoxide PCC Pyridinium chlorochromate KTBT C Palladium on activated charcoal COD Cycloocta-1,5-diene Phen/1,10- phen/1,10- phen phen phen phen m-CPBA m-Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OAc) ₂ Copper(II) actate S-Phos 2-Dicyclohexylphosphino-2', 6'-dimethoxybiphenyl Cu(OTf) ₂ Copper(I) trifluoromethanesulfonate e) Phenyliodine(III)bis(trifluoroacetat Cy Cyclohexyl PPh ₃ Triphenylphosphine Phenyliodine(III)bis(trifluoroacetat DBU 1,8- Dizabicyclo[5.4.0]undec-7-cne Sc(OTf) ₃ Scandium(III) <	Boc	<i>t</i> -Butyloxycarbonyl	MeNO ₂	Nitromethane	
n-BuLi n-Butyl lithium NBS N-Bromosuccinimide t-Bu t-Butyl NCS N-Chlorosuccinimide t-BuOH t-Butyl lithium NIS N-Iodosuccinimide t-BuOH t-Butyl alcohol NMP N-Methyl-2-pyrrolidone t-BuOK / Potassium tertiary butoxide PCC Pyridinium chlorochromate KTBT C Palladium on activated charcoal COD Cycloocta-1,5-diene Phen/1,10- phen 1,10-Phenanthroline m-CPBA m-Chloroperbenzoic acid Ph_2O Diphenyl ether Cu(OTc)_ Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2',6'- dimethoxybiphenyl Cu(OTf)_ Copper(I) thiophene-2- carboxylate PPA Polyphosphoric acid Cy Cyclohexyl PPh_3 Triphenylphosphine DBU 1,8- Dizazbicyclo[5.4.0]undec-7- ene Py Pyridine DCC Dicyclohexylcabodiimide TBAC1 Tetra-n-butylammonium fluoride DCL Dicyclohexylcabodiimide TBAC1 Tetra-n-butylammonium fluoride DCC Dicyclohexylcabodiimide	<i>n</i> -Bu/Bu	normal (primary) Butyl	MOM	Methoxymethyl ether	
A DitalA ProbabilityA Part of the second se	<i>n</i> -BuLi	<i>n</i> -Butyl lithium	NBS	N-Bromosuccinimide	
t-BuLi t-Butyl lithium NIS N-Iodosuccinimide t-BuOH t-Butyl alcohol NMP N-Methyl-2-pyrrolidone t-BuOK / Potassium tertiary butoxide PCC Pyridinium chlorochromate KTBT Pd/C Palladium on activated charcoal COD Cyclocat-1,5-diene Phen/1,10- 1,10-Phenanthroline m-CPBA m-Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OAc)2 Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2',6'-dimethoxybiphenyl Cu(OTf2 Copper(II) inophene-2- carboxylate PPA Polyphosphoric acid CuTc Copper(I) thiophene-2- carboxylate PPA Polyphosphoric acid Cy Cyclohexyl PPh3 Triphenylphosphine DBU 1,8- Diazabicyclo[5.4.0]undcc-7- ene Py Pyridine O-DCB o-Dichlorobenzene Sc(OTf)3 Scandium(III) trifluoromethanesulfonate DCC Dicyclohexylcabodiimide TBACI Tetra-n-butylammonium fluoride DCE 1,2-Dichlorobtane TBAF Tetra-n-butylammonium fluoride DCM Dichlorobethane TBHP tert-Butyl hydroperoxide	<i>t</i> -Bu	t-Butyl	NCS	<i>N</i> -Chlorosuccinimide	
1 Note1 Note1 Noter-BuOHr-Butyl alcoholNMPr-BuOKPotassium tertiary butoxidePCCPyridinium chlorochromateKTBTPotassium tertiary butoxideCH ₅ SO ₃ HMethane sulfonic acidPd/CPalladium on activated charcoalCODCycloocta-1,5-dienephenPhen/1,10-phen1,10-Phenanthrolinem-CPBAm-Chloroperbenzoic acidPh ₂ ODiphenyl etherCu(OAc)2Copper(II) acetateCopper(I)S-Phostrifluoromethanesulfonatee)CuTcCopper(I) thiophene-2- carboxylateCyCyclohexylPPAPolyphosphoric acidDBU1,8- Diazabicyclo[5.4.0]undec-7- eneo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACTetra-n-butylammonium chlorideDCE1,2-DichlorobaneDCE2,3-Dichloro5,6- dicyanobenzoquinoneDEADDiethyl azodicarboxylateDEADDiethyl acodicarboxylateDIADDiisopropyl azodicarboxylateDIADDiisopropyl azodicarboxylateDIADDimethylactamideDIADDimethylactamideDIADDimethylactamideDIADDimethylactamideDIADDimethylactamideDIADDimethylactamideDIADDimethylactamideDIADDimethylactamideDIADDimethylactamide<	t-BuLi	<i>t</i> -Butyl lithium	NIS	<i>N</i> -Iodosuccinimide	
ParticlePotasylationNMPPARCUPT2-pyrionitonicCBuOK /Potassium tertiary butoxidePCCPyridinium chlorochromateKTBTPCCPalladium on activated charcoalCDCycloocta-1,5-dienePhen/1,10- phen1,10-Phenanthrolinem-CPBAm-Chloroperbenzoic acidPh_2ODiphenyl etherCu(OAc)2Copper(II) acetateS-Phos2-Dicyclohexylphosphino-2',6'- dimethoxybiphenylCu(OT)2Copper(II) trifluoromethanesulfonatePIFAPhenyliodine(III)bis(trifluoroacetat e)Cu(OT)2Copper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh_3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3 Scandium(III) trifluoromethanesulfonateScandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBAC1 Tetra-n-butylammonium chlorideDCE1,2-DichloroethaneTBHP tert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFA Trifluoroacetic acidTrifluoroacetix acidDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMADimethylantininopyridineTMOFTrimethyl orthoformate		t Putyl alaohal	NMD	N Mothyl 2 pyrrolidono	
Product / KTBTPortastum fertiary butoxidePCCPyridinum enforcementateKTBTPd/CPalladium on activated charcoalCCh_SO_3HMethane sulfonic acidPd/CPalladium on activated charcoalCODCycloocta-1,5-dienePhen/1,10- phen1,10-Phenanthrolinem-CPBAm-Chloroperbenzoic acidPh_2ODiphenyl etherCu(OAc)2Copper(II) acetateS-Phos2-Dicyclohexylphosphino-2',6'- dimethoxybiphenylCu(OTf)2Copper(II) trifluoromethanesulfonatePIFAPhenyliodine(III)bis(trifluoroacetat e)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh_3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetra-n-butylammonium chlorideDCE1,2-DichloroethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetiz acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetiz acidDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMFN,N-Dimethylfo				N-Methyl-2-pytrondone	
CH ₃ SO ₃ H Methane sulfonic acid Pd/C Palladium on activated charcoal COD Cycloocta-1,5-diene Phen/1,10- phen 1,10-Phenanthroline m-CPBA m-Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OAc) ₂ Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2',6'- dimethoxybiphenyl Cu(OT) ₂ Copper(II) triplene-2- carboxylate PIFA Phenyliodine(III)bis(trifluoroacetat c) CuTc Copper(I) thiophene-2- carboxylate PPA Polyphosphoric acid DBU 1,8- Diazabicyclo[5.4.0]undec-7- ene Py Pyridine o-DCB o-Dichlorobenzene Sc(OTf) ₃ Scandium(III) trifluoromethanesulfonate DCC Dicyclohexylcabodiimide TBACI Tetrabutylammonium filoride DCE 1,2-Dichloroethane TBAF Tetra-n-butylammonium filoride DCM Dichloromethane TBHP tetra-Butyl hydroperoxide DDQ 2,3-Dichloro-5,6- dicyanobenzoquinone TEBAC Benzyltriethylammonium chloride DIAD Diisopropyl azodicarboxylate TFA Trifluoroacetic acid DIH 1,3-Diio	t-BuOK / KTBT	Potassium tertiary butoxide	PCC	Pyridinium chlorochromate	
CODCycloocta-1,5-dienePhen/1,10- phen1,10-Phenanthrolinem-CPBAm-Chloroperbenzoic acidPh2ODiphenyl etherCu(OAc)2Copper(II) acetateS-Phos2-Dicyclohexylphosphino-2',6'- dimethoxybiphenylCu(OTf)2Copper(II) acetatePIFAPhenyliodine(III)bis(trifluoroacetat e)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBAC1Tetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIADDiisopropyl azodicarboxylateTFAPO2,2,2-Trifluoroacetic acidDMADimethylacetamideTFE2,2,2-TrifluoroacetanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMADimethylacetamideTFFTetramethylofuranDMADimethyloformamideTMOFTrimethyl orthoformate	CH ₃ SO ₃ H	Methane sulfonic acid	Pd/C	Palladium on activated charcoal	
m-CPBAm-Chloroperbenzoic acidPhenm-CPBAm-Chloroperbenzoic acidPh2ODiphenyl etherCu(OAc)2Copper(II) acetateS-Phos2-Dicyclohexylphosphino-2',6'-dimethoxybiphenylCu(OTf)2Copper(II) trifluoromethanesulfonatePIFAPhenyliodine(III)bis(trifluoroacetat e)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- 	COD	Cycloocta-1,5-diene	Phen/1,10-	1,10-Phenanthroline	
m-CPBA m-Chloroperbenzoic acid Ph ₂ O Diphenyl ether Cu(OAc) ₂ Copper(II) acetate S-Phos 2-Dicyclohexylphosphino-2',6'-dimethoxybiphenyl Cu(OTf) ₂ Copper(II) trifluoromethanesulfonate PIFA Phenyliodine(III)bis(trifluoroacetat e) CuTc Copper(I) thiophene-2-carboxylate PPA Polyphosphoric acid Cy Cyclohexyl PPh ₃ Triphenylphosphine DBU 1,8- Py Pyridine Diazabicyclo[5.4.0]undec-7-ene PQ Pyridine o-DCB o-Dichlorobenzene Sc(OTf) ₃ Scandium(III) trifluoromethanesulfonate DCC Dicyclohexylcabodiimide TBACI Tetra-n-butylammonium chloride DCE 1,2-Dichloroethane TBHP tert-Butyl hydroperoxide DDQ 2,3-Dichloro-5,6- TEBAC Benzyltriethylammonium chloride DIAD Diisopropyl TFA Trifluoroacetic acid azodicarboxylate TFA Trifluoroacetic acid DIAD Diisopropyl TFA Trifluoroacetic acid DIAD Diisopropyl TFA Trifluoroacetic acid azodicarboxylate TFA			phen		
Cu(OAc)2Copper(II) acetateS-Phos2-Dicyclohexylphosphino-2',6'- dimethoxybiphenylCu(OTf)2Copper(II) trifluoromethanesulfonatePIFAPhenyliodine(III)bis(trifluoroacetat e)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium chlorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTHFTetrahydrofuranDMFN,N-DimethylformanideTMOFTrimethyl orthoformate	<i>m</i> -CPBA	<i>m</i> -Chloroperbenzoic acid	Ph ₂ O	Diphenyl ether	
Cu(OTf)2Copper(II) trifluoromethanesulfonatePIFAPhenyliodine(III)bis(trifluoroacetat e)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium chlorideDCCDicyclohexylcabodiimideTBAFTetra-n-butylammonium chlorideDCE1,2-DichloroethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimeth	Cu(OAc) ₂	Copper(II) acetate	S-Phos	2-Dicyclohexylphosphino-2',6'-	
Cu(OTf)2Copper(II) trifluoromethanesulfonatePIFAPhenyliodine(III)bis(trifluoroacetat e)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBHPtert-Butyl hydroperoxideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylbydantoinTFATrifluoroacetic acidDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTHF <td></td> <td></td> <td></td> <td>dimethoxybiphenyl</td>				dimethoxybiphenyl	
trifluoromethanesulfonatee)CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDIADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetic acidDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylaminopyridineTfOHTriflic acidDMADimethylaminopyridineTHFTetrahydrofuranDMADimethylaminopyridineTMOFTrimethyl orthoformate	Cu(OTf) ₂	Copper(II)	PIFA	Phenyliodine(III)bis(trifluoroacetat	
CuTcCopper(I) thiophene-2- carboxylatePPAPolyphosphoric acidCyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanbenzoquinoneTEBACBenzyltriethylammonium chlorideDIADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetic acidDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylaminopyridineTfOHTriflic acidDMADimethylaminopyridineTHFTetrahydrofuranDMADimethylaminopyridineTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane		trifluoromethanesulfonate		e)	
CyCyclohexylPPh3TriphenylphosphineDBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylbydantoinTFATrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2.2-TrifluoroethanolDMADimethylacetamideTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	CuTc	Copper(I) thiophene-2- carboxylate	PPA	Polyphosphoric acid	
DBU1,8- Diazabicyclo[5.4.0]undec-7- enePyPyridineo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBAClTetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- 	Су	Cyclohexyl	PPh ₃	Triphenylphosphine	
Diazabicyclo[5.4.0]undec-7- eneSc(OTf)3Scandium(III) trifluoromethanesulfonateo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBACITetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylaminopyridineTfOHTriflic acidDMFN,N-DimethylformanideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DBU	1,8-	Py	Pyridine	
eneSc(OTf)3Scandium(III) trifluoromethanesulfonateo-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBAC1Tetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylaminopyridineTfOHTriflic acidDMEDimethylgrmamideTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane		Diazabicyclo[5.4.0]undec-7-	5		
o-DCBo-DichlorobenzeneSc(OTf)3Scandium(III) trifluoromethanesulfonateDCCDicyclohexylcabodiimideTBAClTetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane		ene			
DCCDicyclohexylcabodiimideTBAClTetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFHTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	o-DCB	o-Dichlorobenzene	Sc(OTf) ₃	Scandium(III)	
DCCDicyclohexylcabodiimideTBAClTetrabutylammonium chlorideDCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane				trifluoromethanesulfonate	
DCE1,2-DichloroethaneTBAFTetra-n-butylammonium fluorideDCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DCC	Dicyclohexylcabodiimide	TBACl	Tetrabutylammonium chloride	
DCMDichloromethaneTBHPtert-Butyl hydroperoxideDDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DCE	1,2-Dichloroethane	TBAF	Tetra- <i>n</i> -butylammonium fluoride	
DDQ2,3-Dichloro-5,6- dicyanobenzoquinoneTEBACBenzyltriethylammonium chlorideDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DCM	Dichloromethane	TBHP	tert-Butyl hydroperoxide	
dicyanobenzoquinoneTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DDQ	2,3-Dichloro-5,6-	TEBAC	Benzyltriethylammonium chloride	
DEADDiethyl azodicarboxylateTEMPO2,2,6,6-Tetramethylpiperidin-1-ylDIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane		dicyanobenzoquinone			
DIADDiisopropyl azodicarboxylateTFATrifluoroacetic acidDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DEAD	Diethyl azodicarboxylate	TEMPO	2,2,6,6-Tetramethylpiperidin-1-yl	
azodicarboxylateDIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DIAD	Diisopropyl	TFA	Trifluoroacetic acid	
DIH1,3-Diiodo-5,5- dimethylhydantoinTFATTrifluoroacetyltriflateDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane		azodicarboxylate			
dimethylhydantoindimethylhydantoinDMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DIH	1,3-Diiodo-5,5-	TFAT	Trifluoroacetyltriflate	
DMADimethylacetamideTFE2,2,2-TrifluoroethanolDMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane		dimethylhydantoin			
DMAP4-DimethylaminopyridineTfOHTriflic acidDMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DMA	Dimethylacetamide	TFE	2,2,2-Trifluoroethanol	
DMEDimethoxyethaneTHFTetrahydrofuranDMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DMAP	4-Dimethylaminopyridine	TfOH	Triflic acid	
DMFN,N-DimethylformamideTMOFTrimethyl orthoformateDMSODimethyl sulfoxideTMSTetramethylsilane	DME	Dimethoxyethane	THF	Tetrahydrofuran	
DMSO Dimethyl sulfoxide TMS Tetramethylsilane	DMF	<i>N</i> , <i>N</i> -Dimethylformamide	TMOF	Trimethyl orthoformate	
	DMSO	Dimethyl sulfoxide	TMS	Tetramethylsilane	

dppb	1, 4-	Tol-BINAP	(S)-(-)-2,2'-Bis(di- <i>p</i> -
	Bis(diphenylphosphino)buta		tolylphosphino)-1,1'-binaphthyl
	ne		
dppf	1,1'-	p-TsOH/p-	<i>p</i> -Toluene sulfonic acid
	Bis(diphenylphosphino)ferr	TSA	
	ocene		
DPPH	2,2-Diphenyl-1-	Zn(OTf) ₂	Zinc trifluoromethanesulfonate
	picrylhydrazyl		

Spectroscopic Abbreviations

IR	Infrared	XRD	X-ray diffraction
v_{max}	Frequency maximum	ppm	Parts per million
cm ⁻¹	Frequency in wavenumber	ppb	Parts per billion
UV	Ultra violet	δ	Delta (Chemical shift
			in ppm)
NMR	Nuclear magnetic resonance	MHz	Megahertz
CDCl ₃	Deuterated chloroform	Hz	Hertz
DMSO-d ₆	Deuterated dimethyl sulfoxide	S	Singlet
acetone-d6	Deuterated acetone	d	Doublet
m	Multiplet	t	Triplet
dd	Doublet of doublet	q	Quartet
td	Triplet of a doublet	J	Coupling constant
dt	Doublet of a triplet	DEPT	Distortionless
			Enhancement by
			Polarization Transfer
br s	Broad singlet	HRMS	High Resolution Mass
			Spectrometry
M ⁺	Molecular ion	HPLC	High performance
			liquid chromatography
m/z	Mass to charge ratio	ICP-MS	Inductively coupled
			plasma mass
			spectrometry

ABSTRACT OF THESIS

TITLE: SYNTHETIC STUDIES IN OXYGEN HETEROCYCLES

Oxygen heterocyclic compounds occupy a prominent place in organic chemistry. Multifarious naturally occurring oxygen heterocycles exhibit various biological activities. The objective of this thesis was to develop short and efficient methodologies for the synthesis of selected oxygen heterocycles such as chromans, flavones and coumestans and evaluating some synthesized analogues for anti-diabetic activity. The thesis is divided into 4 chapters.

The first chapter provides a short review on the molecular iodine mediated recent developments in the synthesis of diverse heterocycles. The role of molecular iodine either as a sole catalyst or reagent and/or in presence of oxidant has been presented. The importance and versatility of iodine in organic synthesis can be understood from the documented one pot or two step one pot synthesis of several nitrogen, oxygen and sulphur heterocycles having simple to complex structures with diverse functional groups.

The second chapter describes a one pot method for the synthesis of chromans *via* [3+3] cyclocoupling of phenols with allylic alcohols using molecular iodine as a catalyst in refluxing chloroform or methanol solvent (Scheme I). The usefulness of this method is presented by synthesizing 13 chroman derivatives including naturally occurring dihydrolapachenole. Also, the synthesis of naturally occurring chromenes *viz*. precocene II and lapachenole has been demonstrated by conventional dehydrogenation of corresponding chromans.





The third chapter discusses a one pot method for the synthesis of flavones from 2'hydroxyacetophenones and aromatic aldehydes using pyrrolidine as a base and iodine as an oxidant in DMSO solvent at 150 °C (Scheme II). The wide substrate scope was demonstrated by synthesizing 18 derivatives of flavones in good yields. The flavone formation involves domino aldol-Michael-dehydrogenation sequence. Inexpensive catalysts, use of simple substrates which eliminates the preparation and isolation of chalcone or flavanone intermediate, good substrate generality, absence of metal catalysts and high yields of products without any side reactions make this method more advantageous.



Also, this chapter includes the anti-diabetic activity studies of flavones. Among several synthesized flavones, derivative **A** (Figure 1) showed highest inhibition of α -glucosidase enzyme. Dose dependent inhibition of α -glucosidase by **A** ranged from $8.4\pm 0.37\%$ at 1 µg/mL to 99.3± 0.26% at 7.6 µg/mL. Thus, it is active in very low concentration as compared to the standard anti-diabetic drug acarbose which shows maximum inhibition at 400 µg/mL. Kinetic study of **A** showed non-competitive type of inhibition whereas molecular docking study showed that the **A** occupy the allosteric site and is involved in the hydrogen bonding with amino acid Lys373 of α -glucosidase enzyme.



Figure 1

The fourth chapter deals with a two step synthesis of coumestan from 3-(2-hydroxyphenyl)coumarin which in turn was prepared by the condensation of salicylaldehydes and 2-coumaranone or 2-hydroxyphenylacetic acids (Scheme III). The oxidative cyclization of 3-(2-hydroxyphenyl)coumarin to produce coumestans was achieved using 1 equiv of anhydrous Cu(OAc)₂ in diphenyl ether under refluxing condition.



The broad scope of this method was demonstrated by synthesizing 23 coumestans with electron donating as well as electron releasing functionalities. These derivatives include dimethyl ether of naturally occurring coumestrol and sativol and trimethyl ether of lucernol obtained in 53-80 % yields. This efficient methodology was also successfully applied for the direct synthesis of hydroxyl substituted natural coumestans *viz*. coumestrol and 4'-*O*-methylcoumestrol (Figure 2) without any protection making this method superior.



Figure 2

Use of economical $Cu(OAc)_2$, absence of any additional reagent/additive, simple reaction procedure, large substrate scope, effortless product isolation & good yields are several advantages of this method. Additional advantages includes stepwise one pot synthesis, large scale preparation, and possible use of catalytic amount of $Cu(OAc)_2$. The probable mechanism is discussed by performing additional control experiments.

LIST OF PUBLICATIONS

 Molecular iodine catalyst promoted synthesis of chromans and 4-aryl-3,4dihydrobenzopyran-2-ones via [3+3] cyclocoupling, Naik, M. M.; Kamat, D. P.; Tilve, S. G.; Kamat, V. P. *Tetrahedron* 2014, *70*, 5221-5233. https://doi.org/10.1016/j.tet.2014.05.093

2. Pyrrolidine and iodine catalyzed domino aldol-Michael-dehydrogenative synthesis of flavones, **Naik, M. M.;** Tilve, S. G.; Kamat, V. P. *Tetrahedron Lett.* **2014**, *55*, 3340-3343.

https://doi.org/10.1016/j.tetlet.2014.04.051

3. Graphite as an effective catalyst for Michael addition of indoles to nitroolefins in aqueous alcoholic solution, Parsekar, S. B.; Naik, S.; Naik, M. M.; Tilve, S. G. *Monatsch. chem.* **2015**, *146*, 691-696.

https://doi.org/10.1007/s00706-014-1347-x

4. Molecular iodine, Naik, M. M. Iran. J. Catal. SPOTLIGHT 2015, 5, 383-385.

5. Copper-mediated synthesis of coumestans *via* C(sp2)-H functionalization: Protective group free route to coumestrol and 4'-*O*-methylcoumestrol, **Naik**, **M. M.;** Kamat, V. P.; Tilve, S. G. *Tetrahedron* **2017**, *73*, 5528-5536. https://doi.org/10.1016/j.tet.2017.07.057

6. A comprehensive review on coumestan synthesis, Naik, M. M.; Kamat, V. P.; Tilve, S.G. (*Manuscript under preparation*).

7. α -Glucosidase inhibition activity & *in silico* study of synthetic flavone derivative as a potential anti-diabetic drug nominee

Meena, S. N.; Kumar, U.; Naik, M. M.; Ghadi, S. S.; Kamat, V. P.; Tilve, S. G. (*Manuscript under preparation*).

Posters presented at National/International conferences:

1. Presented poster entitled "Molecular iodine catalyst promoted cyclocoupling of phenols and allylic alcohols to Chromans" at Second UK-India MedChem Congress 2013 symposium in CSIR Indian Institute of Chemical Technology, Hyderabad (22nd – 23rd March 2013).

2. Presented poster entitled "Pyrrolidine and iodine catalyzed one pot synthesis of flavones from 2'-hydroxyacetophenone and aromatic aldehydes" at 16^{th} CRSI National Symposium in Chemistry (NSC-16) in Department of Chemistry, IIT Bombay $(7^{th} - 9^{th}$ February 2014).

3. Presented poster entitled "Synthesis of coumestans *via* C-H functionalization of 3-(2-hydroxyphenyl)-2*H*-chromen-2-one using Cu(OAc)₂" at National Conference on New Frontiers in Chemistry-From Fundamentals to Applications (NFCFA) in Department of Chemistry, Birla Institute of Technology and Science Pilani, K. K. Birla Goa ($18^{th} - 19^{th}$ December 2015).

Conferences attended:

1. Participated in Junior-National Organic Symposium Trust (J-NOST) Conference for Research Scholars held in Indian Institute of Science Education and Research Bhopal (IISER) Bhopal, Madhya Pradesh ($4^{th} - 6^{th}$ December 2013).

 Participated in Conference on Chemical Industrial Disaster Management (CIDM): Emergency Planning and Disaster Management in Chemical, Petroleum, Petrochemical and Pharmaceutical Industry at Cidade De Goa, Goa (29th September – 1st October 2014).

3. Participated in National Symposium on Transcending Frontiers in Organic Chemistry (TFOC) at CSIR-National Institute for Interdisciplinary Science and Technology (NIIST), Trivandrum, Kerala $(9^{th} - 11^{th} \text{ October 2014})$.

Participated in International Conference on Green Chemistry: Catalysis, Energy and Environment (ICGC) at Chemistry Seminar Hall, Faculty Block E, Goa University, Goa (22nd – 24th January 2015).

<u>CHAPTER 1</u>

A short review on the recent

advancements in the synthesis of

heterocycles using molecular

iodine as a catalyst or reagent



1.1: Introduction

Molecular iodine in the recent years has emerged as an attractive tool in the field of synthetic organic chemistry. Being an environmentally benign chemical, it has been extensively employed in organic synthesis either as a catalyst or as a reagent. A number of reviews on its wide applications have been well documented in the literature.¹ Some of the recent reviews focus on C-N bond formation through redox reaction,^{1a} synthesis of heterocycles *via* electrophilic cylization of alkynes,^{1b} oxidative coupling reactions utilizing C-H and X-H as nucleophiles,^{1c} iodine catalysis as a green alternative to transition metals in organic chemistry and technology,^{1d} recent advances in organic reactions,^{1e} its use in monomer and polymer designing,^{1f} multicomponent reactions,^{1g} five and six membered heterocycles synthesis,^{1h} iodination and protection-deprotection of functional groups¹ⁱ and synthesis of chromone type compounds.^{1j} Also the synthetic applications of hypervalent iodine compounds have been recently reviewed.² Our group has also explored its applications in the synthesis of diverse heterocycles.³

Various advantages associated with molecular iodine include low cost, less-toxicity, ready availability and easy removal after reaction by simply washing with reducing agent such as sodium thiosulphate solution. In addition, its stability to air and moisture makes it easy to handle. The amount of iodine used varies from catalytic to stoichiometric or even excess for some reactions. Reactions employing iodine provides mild operating conditions usually allowing large substrate scope, high yields, regio/stereoselectivities, short reaction time and simple work up procedure. Solvent-less reactions and solid-supported iodine adds to the advantages of iodine. The mild Lewis acidity of iodine has enhanced its use in the organic synthesis. In many reactions it acts as a substitute to costly and hazardous acid/metal catalysts and/or reagents thus enabling the elimination of hazardous metals and providing a "metal free" approach. This makes iodine an eco-friendly alternative to acid/metal based reagents/catalysts which pose serious health and safety problems. As a result, it has gained considerable attention in the past decades and several green methodologies are being continuously developed using iodine. The present mini review highlights the recent applications of environmentally benign molecular iodine either as a sole catalyst or reagent in the synthesis of heterocycles.

1.2.1: Iodine as a sole catalyst

Yang *et al.*⁴ devised a metal free regio-divergent approach for the synthesis of α and δ -carbolines from a common indolyldihydrochalcone oxime ester substrate. Several derivatives of δ -carbolines were synthesized using catalytic iodine in DME at 100 °C. However, α -carbolines were obtained when the reaction was carried out in presence of DDQ (Scheme 1).



Scheme 1

A wide range of 2-aryl substituted benzimidazoles have been synthesized by Ravi *et al.*⁵ from aryl alkyl ketones and 2-amino anilines using molecular iodine. This one step strategy involves consecutive C-N bond formation and $C(sp^2)$ –C bond cleavage giving products in moderate to good yields (Scheme 2).





Sagir *et al.*⁶ developed an iodine catalyzed green methodology for the synthesis of multisubstituted pyrazolo-pyrido-pyrimidines and its spiro analogues in high yields. It involves a one pot four-component reaction of hydrazine, ethyl acetoacetate, 6-amino-1-methyl uracil and isatin or aldehyde in water as solvent. The method is operationally simple allowing products to obtain by filtration without any chromatography or recrystallization (Scheme 3).



Scheme 3

Wang *et al.*⁷ synthesized iodine catalyzed quinazolin-4-(1*H*)-one scaffolds by reacting different types of 1,2-dicarbonyl compounds with 2-aminobenzamides in 1-butyl-3-methylimidazolium bromide [BMIm]Br ionic liquid. Its wide scope is demonstrated by synthesizing library of 39 compounds and the recyclability of [BMIm]Br makes it environmentally benign (Scheme 4).



Scheme 4

Yi and Xi⁸ presented the synthesis of 2*H*-indazoles *via* aerobic-oxidative C-H functionalization of *o*-alkylazoarenes with iodine as catalyst, CuI as additive, pyridine as base and oxygen as terminal oxidant. Both electron rich and electron deficient azozrens were successfully converted to corresponding 2*H*-indazoles and the reaction can also be scaled up to gram level (Scheme 5).



Scheme 5

Sun *et al.*⁹ presented a convenient iodine catalyzed synthesis of C-4 sulfenylated pyrazoles. It is a domino multicomponent reaction involving 1,3-diketones, hydrazines and thiols which undergoes cyclocondensation and direct C-H bond sulphenylation reaction. Two new C-N bonds and one C-S bond are generated simultaneously in this protocol leading to target molecule (Scheme 6).



Scheme 6

Iyer and co-workers¹⁰ prepared aryl 4H-3,1-benzoxazin-4-ones using 2-aminobenzoic acid and aryl aldehydes. It is catalyzed by iodine in presence of oxone as an oxidant. This straight forward metal free approach employs readily available starting materials using environmentally benign oxidant (Scheme 7).



Scheme 7

An iodine catalyzed synthesis of 2-azaarenyl benzimidazoles and 2-azaarenyl benzothiazoles has been achieved by Yaragorla and Babu¹¹ from 2-methylazaarenes and *o*-phenylenediamine/2-aminothiophenol in DMSO. The umpolung behaviour of the methyl group of 2-methylazaarenes serving as an electrophile is executed. This oxidative $C(sp^3)$ –H functionalization of 2-methylazaarenes provides a broad substrate scope and high yields under open air conditions (Scheme 8).





Dighe *et al.*¹² presented the synthesis of 1-aroyl- β -carbolines by oxidative Pictet-Spengler reaction and terminal alkynes serving as the 2-oxoaldehyde surrogate mediated by iodine in DMSO. The broad substrate scope is well explored by synthesizing variety of derivatives, some of which were utilized for the total synthesis of naturally occurring Fascaplysin, Eudistomins Y1 and Y2 (Scheme 9).



Scheme	9
--------	---

Further applications includes the synthesis of pyrrolo[1,2-*a*]-quinoxaline and indolo[1,2-*a*]quinoxaline derivatives from respective pyrrole- and indole-based substrates. Interestingly, *N*-substituted indoles with no substituent at C-3 reacted with aromatic alkynes to form 3-thiomethylated indolo[1,2-*a*]quinoxalines. However on blocking C-3 position of indole with methyl group, desired product was formed. Also further utility of 1-aroyl- β -carbolines was shown by synthesizing thiomethylated and 4-aryl substituted canthin-6-ones derivatives (Scheme 10).



Harnying *et al.*¹³ employed catalytic iodine as low as 0.5-5 mol% in Prins reaction giving *rac*-1,3-dioxanes from substituted styrenes and aliphatic aldehydes. Prior to this report, Yadav *et al.*¹⁴ had explored the application of molecular iodine (1 equiv) in Prins reaction. However the amount of iodine was greatly reduced from stoichiometric to catalytic in the present work in presence of pyridinium bis(trifluoromethanesulfonyl)imide (TFSI) salt in 1:1 ratio to iodine. Diasteroselectivity of **A**:**B** up to 82:18 was observed by the isomerization of 1,3-dioxane products to the thermodynamically favoured one **A** (Scheme 11).



Scheme 11

Deb *et al.*¹⁵ presented an I_2/H_2O_2 promoted synthesis of 1,3-oxazines in moderate to good yields from 1-(aminoalkyl)-2-naphthols or 2-(aminoalkyl)phenols. This reaction involving an oxidative intramolecular C–O bond formation by catalytic iodine and H_2O_2 oxidant smoothly occurs at room temperature (Scheme 12).





Buduma *et al.*¹⁶ developed a simple and efficient synthesis of 2-pyridones using iodine catalyst in refluxing ethanol from 4H-pyrans. Both aromatic as well as hetero aromatic ring containing 4H-pyrans were successfully converted to the respective 2-pyridones. The analogues synthesized were tested for in vitro antiproliferative activity (Scheme 13).



Scheme 13

An $I_2/TBHP$ mediated efficient route to the synthesis of 1,3,5-trisubstituted 1,2,4-triazoles was described by Chen *et al.*¹⁷ from hydrazones and aliphatic amines. The substrate scope is well demonstrated by synthesizing a library of analogues in moderate to excellent yields. It also allows the synthesis of products in gram scale (Scheme 14).



Scheme 14

Zhang and co-workers¹⁸ synthesized 1,2,4-triarylpyrroles using a novel iodine mediated approach in moderate to good yields. It involved a cascade condensation-cyclization of simple substrates such as aryl methyl ketones and anilines avoiding any intermediate preparation (Scheme 15).



Scheme 15

Iodine catalyzed reductive redox cyclization of *o*-nitro-*t*-anilines with formic acid into fused tricyclic or 1,2-disubstituted benzimidazoles was successfully established by Nguyen *et al.*¹⁹ Broad substrate scope has been demonstrated. Also 1,2-disubstituted benzimidazole hydriodide derivatives were prepared by using 0.5 equiv of iodine thus playing dual role as catalyst as well as iodide source (Scheme 16).



Scheme 16

Sun *et al.*²⁰ reported iodine catalyzed route to new pyrazolo-[1,5-*a*]pyrimidin-4-ium sulfonates *via* regioselective bicyclization reaction of β -ketonitriles with sulfonyl hydrazides which under alkaline medium delivered densely functionalized pyrazolo[1,5-*a*]pyrimidines. Unprecedented direct C(sp²)–H bond bifunctionalization of these compounds on treatment with sulfonyl hydrazides provided fully substituted pyrazolo-[1,5-*a*]pyrimidines (Scheme 17).





Feng *et al.*²¹ developed a convenient iodine catalyzed route to 6-oxo-5,6dihydrodibenzo[b,h][1,6]naphthyridine-11-carboxamide derivatives. 2-Aminobenzamides and mucobromic acid on refluxing in THF undergoes a domino-type mechanism involving double elimination of hydrogen bromide (Scheme 18).



Scheme 18

Reddy *et al.*²² described an efficient iodine catalyzed route to indolizine-1-carboxylates from 2-pyridyl acetates and alkynes/alkenes. It comprises of oxidative C-C and C-N bond formations in absence of any metal, oxidant or base. The exact reaction mechanism is not proposed, however control experiments suggests a radical pathway (Scheme 19).



Scheme 19

A methodology for quinazolines preparation was developed by Tiwari and Bhanage²³ starting from 2-aminobenzylamines and with or without *N*-substituted benzylamines. The method employs iodine catalyst in presence of oxygen as an oxidant under solvent-free and additive-free conditions (Scheme 20).



Scheme 20

Inturi *et al.*²⁴ presented an I₂-TBHP catalyzed 4,3-fused 1,2,4-triazoles synthesis from *N*-tosylhydrazones and aromatic *N*-heterocycles. It occurs through generation of azomethine imine which undergoes regioselective 1,3-dipolar cycloaddition with variety of aromatic *N*-heterocycles. Large substrate scope has also been accomplished by synthesising variety of derivatives (Scheme 21).



Scheme 21

Also, the same group designed a two step one pot procedure to introduce R as linear alkyl groups in the product. Accordingly, the aldehyde substrates were treated with 4-methylbenzenesulfonohydrazide in CH_2Cl_2 at room temperature followed by treatment with 3-methylpyridine in presence of catalytic iodine and TBHP. This allowed the synthesis of triazole derivatives with aliphatic linkage (Scheme 22).





An *et al.*²⁵ prepared 3-nitro-2-arylimidazo[1,2-*a*]pyridine derivatives by an iodine/TBHP/Py mediated approach. It occurs *via* a Michael addition and oxidative coupling tandem reaction (Scheme 23).



Scheme 23

Yang *et al.*²⁶ developed an iodine catalyzed synthetic methodology for 2-heteroaryl quinazolinones from azaarenes through oxidative benzylic C-H bond amination. Several derivatives synthesized up to 95% yield exhibits wide substrate scope of this method (Scheme 24).





Xi *et al.*²⁷ synthesized 3-acylbenzothiadiazine 1,1-dioxides from different acetophenones and various 2-aminobenzenesulfonamides using molecular iodine (Scheme 25). However when 2-aminobenzenesulfonamides with an alkynyl group were treated under similar reaction condition, the triple bond was oxidized to *o*-diketone group along with the formation of 3-acylbenzothiadiazine 1,1-dioxide ring (Scheme 26).



Scheme 25



Scheme 26

Synthesis of chromeno[2,3-*b*]indoles was achieved by Deb *et al.*²⁸ from 3-(α , α -diarylmethyl)indoles through intramolecular dehydrogenative coupling reaction to form C-O bond. I₂/TBHP has been found to be an efficient medium for this conversion (Scheme 27).





Le *et al.*²⁹ developed a method to synthesize 2-methylquinolines by the condensation reaction of substituted anilines and vinyl ethers using iodine catalyst. The dual role of iodine species is explored wherein molecular iodine acts as an oxidant and hydrogen iodide acts as an activating agent by activating vinyl ether. Thus, this redox reaction allows molecular iodine to be employed in catalytic amount (Scheme 28).





Wei *et al.*³⁰ designed a methodology for the synthesis of [2,3]-fused indoline tetrahydropyridazines *via* direct functionalization of the $C(sp^3)$ -H bond. It employs iodine catalyst in presence of TBHP oxidant and *N*-Boc glycine as an acidic additive. It is a three component reaction of substituted indoles, hydrazines and acetophenones which undergoes dearomative oxidative coupling to furnish desired products in moderate to good yields (Scheme 29).



Scheme 29

1.2.2: Iodine as a sole reagent

Qu and co-workers³¹ described an iodine mediated synthesis of cyclic *N*,*O*-acetals through regio- and diastereoselective oxidation of the secondary α –C-H bonds of 2-functionalized pyrrolidines (Scheme 30). The methodology was also applied for the total synthesis of naturally occurring (±)-preussin and its C(3) epimer (Figure 1) from (±)-Boc pyrrolidin-3-ol in nine steps (22% overall yield).



Scheme 30



Figure 1: Structures of (±)-preussin and C(3)-epi-(±)-preussin

An efficient and scalable synthesis of 5-amino and 3,5-diamino substituted 1,2,4-thiadiazoles has been developed by Wang *et al.*³² Molecular iodine as an exclusive oxidant is responsible for the newly formed N-S bond. Its broad substrate scope is well demonstrated by synthesizing numerous derivatives from various imidoyl and guanyl substrates (Scheme 31).



Scheme 31

Hu *et al.*³³ developed a methodology for the synthesis of *N*-protected benzimidazoles by a sequential one pot two step process. The simple *o*-phenylenediamines and aromatic/aliphatic/cinnamic aldehydes condense to form imine intermediates which without any purification on iodine treatment under basic condition led to 1,2-disubstituted benzimidazoles. Authors exhibited broad scope of this intramolecular C–H amidation methodology by synthesizing large number of derivatives including *N*-unsubstituted benzimidazoles. Also the method works well on gram scale (Scheme 32).



Scheme 32

Wu *et al.*³⁴ have prepared 2-acylquinolines from methyl ketones, arylamines and 1,4-dithiane-2,5-diol as an ethylene surrogate *via* an iodine promoted formal [4+2] cycloaddition. Arylamine substrate also promoted 1,4-dithiane-2,5-diol participation in Povarov reaction. The mechanistic studies showed the mechanism operates *via* an iodination-Kornblumoxidation-Povarov-aromatization process. This interesting reaction exhibits the use of 1,4dithiane-2,5-diol in the synthesis of *N*-hetrocycles rather than *S*-heterocycles through desulfurization (Scheme 33).



Singh and co-workers³⁵ designed an efficient regioselective methodology for the synthesis of β -carboline *N*-fused imidazoles from natural α -amino acids/benzyl amines and 1-formyl pyrido[3,4-*b*]indoles *via* molecular iodine promoted decarboxylative amination. Several derivatives were synthesized using aliphatic, aromatic and heteroaromatic α -amino acids up to 91% yield. The plausible mechanism is also well explained (Scheme 34).



Scheme 34

Miao *et al.*³⁶ presented an iodine mediated one pot two step process to synthesize dihydrofurans and furans *via* formal [3 + 2] cycloaddition of 1,3-dicarbonyl compounds with enones. Firstly, the substrates undergo Michael addition on refluxing 4-hydroxycoumarin with benzyl triethyl ammonium chloride in water followed by iodine mediated cyclization. The reaction product depends on the type of base employed in the second step wherein I₂/DMAP furnishes dihydrofurans with high trans diastereoselectivity whereas I₂/DBU leads to furans (Scheme 35). Similarly, other 1,3-dicarbonyl compounds reacted to furnish respective products but in low yields (Scheme 36).



Scheme 36

Zou *et al.*³⁷ synthesized polyfunctionalized pyrroles from furfurylamines and ynones involving a two step one pot approach. The reaction between furfurylamines and ynones in methanol generated *N*-furfuryl- β -enaminones *in situ* which led to polyfunctionalized pyrroles through iodine mediated oxidative annulation reaction (Scheme 37).



Scheme 37

Kale *et al.*³⁸ developed a one pot methodology involving iodine-dimethyl sulfoxide for the synthesis of pyrido-fused imidazo[4,5-*c*]quinolines. The substrates pyridoimidazole arylamines and carbonyl compounds undergoes oxidative cross coupling followed by intramolecular cyclization to deliver products in high yields. A series of compounds were synthesized using different substrates and iodine concentration (Scheme 38).



Scheme 38

Volvoikar and Tilve^{3g} (our group) developed an iodine/TBHP procedure for the synthesis of 5,11-dialkylindolo[3,2-*c*]quinoline salts and 5,7-dimethylindolo[2,3-*c*]quinoline salts *via* intramolecular dehydrogenative coupling. Control experiments were performed to support the hypoiodite mediated mechanism for the annulation followed by aromatization sequence. The application of the methodology was demonstrated by synthesizing naturally occurring isocryptolepine hydroiodide, an indoloquinoline alkaloid (Scheme 39).



Scheme 39

Synthesis of 1,2,5-trisubstituted imidazoles *via* Radziszewski-type reaction between methyl ketones, anilines and tosylmethyl isocyanide was reported by Zhang *et al.*³⁹ employing molecular iodine. It occurs *via* Radziszewski-type reaction mechanism wherein methyl ketones play a dual role as α -dicarbonyl compounds and aldehyde (Scheme 40).





The mechanistic pathway involves the reaction of methyl ketone with iodine to form α iodoketone which then with the liberation of HI forms phenylglyoxal *via* Kornblum oxidation in DMSO. The aniline substrate reacts with the aldehyde group of phenylglyoxal to form Cacylimine which on reaction with *in situ* formed amine from tosylmethyl isocyanide and HI results in the formation of intermediate *via* an *in situ* cross-trapping process. Lastly, the cyclocondensation of intermediate formed with another phenylglyoxal molecule provides the target molecule (Scheme 41).




Likhar and co-workers⁴⁰ achieved iodo-substituted dihydrofurans regioselectively from amine substituted alkynols *via* iodocyclization in presence of excess of iodine, base and solvent (Scheme 42). As a part of further application, the products obtained with iodo substituent were functionalized by performing different C-C and C-N coupling reactions.





Azimi and Azizian⁴¹ described a mild, one pot three component, high yielding synthetic protocol for 2,3-dihydroquinazolin-4(1H)-ones preparation by employing benzyl alcohols, isatoic anhydride and primary amines as substrates using iodine under basic condition (Scheme 43). *In situ* oxidation of benzyl alcohols to benzaldehydes allows avoiding the use of unstable aldehydes. Author claims this work as the first report using benzyl alcohols instead of benzaldehydes in dihydroquinazolins synthesis.





An iodine-mediated synthesis of highly functionalized arylazopyrazoles was achieved by Pandit *et al.*⁴² from β -ketoesters and arylhydrazines (2 equiv) in presence of catalytic AgNO₃. The product formation occurs *via* α -iodination of β -ketoesters, formation of pyrazol-3-one followed by substitution with a nitrogen nucleophile and finally oxidation followed by enolization. Also 2-arylpyrazol-3-ones were reacted with arylhydrazines (1 equiv) to prepare arylazopyrazoles with two different arylhydrazine entities (Scheme 44).



Scheme 44

Ramesha *et al.*⁴³ reported an iodine-mediated approach for the synthesis of 3-sulfenylimidazo[1,5-a]pyridines through C–H functionalization in good yields. It makes use of dithioesters, 2-methylaminopyridines and sulfonyl hydrazides as substrates. Also a series of imidazopyridines have been synthesized in good yields from dithioesters, 2-methylaminopyridines (Scheme 45).



Scheme 45

1.3: Conclusion

The present mini review has included the recent developments in diverse heterocycles synthesis using molecular iodine either as a sole catalyst or reagent and/or in presence of oxidant. Numerous nitrogen heterocycles and a few of oxygen and sulphur heterocycles synthesis with a wide range of functional group diversity comprising of simple to complex structural skeleton in one pot or two step one pot method suggests the importance and versatility of iodine in organic synthesis. Thus from future point of view, an efficient, green, economic and very useful molecular iodine will be explored further as catalyst/reagent to accomplish the synthesis of many more diversified complex heterocycles.

1.4: References

- For recent iodine reviews: a) Nguyen, T. B. Asian J. Org. Chem. 2017, 6, 477. b) Aggarwal, T.; Kumar, S.; Verma, A. K. Org. Biomol. Chem. 2016, 14, 7639. c) Liu, D.; Lei, A. Chem. Asian J. 2015, 10, 806. d) Yusubov, M. S.; Zhdankin, V. V. Resour. Effic. Technol. 2015, 1, 49. e) Zhao, J.; Gao, W.; Chang, H.; Li, X.; Liu, Q.; Wei, W. Chin. J. Org. Chem. 2014, 34, 1941. f) Moulay, S. Des. Monomers Polym. 2014, 17, 501. g) Ren, Y-M.; Cai, C.; Yang, R. C. RSC Adv. 2013, 3, 7182. h) Parvatkar, P. T.; Parameswaran, P. S.; Tilve, S. G. Chem. Eur. J. 2012, 18, 5460. i) Tekale, S. U.; Kauthale, S. S.; Dake, S. A.; Sarda, S. R.; Pawar, R. P. Curr. Org. Chem. 2012, 16, 1485. j) Pinto, D. C. G. A.; Silva, A. M. S. Curr. Org. Synth. 2012, 9, 561.
- For recent hypervalent iodine reviews: a) Yoshimura, A.; Zhdankin, V. V. Chem. Rev. 2016, 116, 3328. b) Wang, L.; Liu, J. Eur. J. Org. Chem. 2016, 2016, 1813. c) Arnold, A. M.; Ulmer, A.; Gulder, T. Chem. Eur. J. 2016, 22, 1. d) Yoshimura, A.; Yusubov, M. S.; Zhdankin, V. V. Org. Biomol. Chem. 2016, 14, 4771. e) Dohi, T.; Kita, Y. Curr. Org. Chem. 2016, 20, 580. f) Romero, R. M.; Woeste, T. H.; Muniz, K. Chem. Asian J. 2014, 9, 972. g) Zheng, Z.; Zhang-Negrerie, D.; Du, Y.; Zhao, K. Sci. China Chem. 2014, 57, 189. h) Brown, M.; Farid, U.; Wirth, T. Synlett 2013, 24, 424. i) Parra, A.; Reboredo, S. Chem. Eur. J. 2013, 19, 17244. j) Kajiyama, D.; Saitoh, T.; Nishiyama, S. Electrochemistry 2013, 81, 319. k) Yusubov, M. S.; Zhdankin, V. V. Curr. Org. Synth. 2012, 9, 247.
- a) Parvatkar, P. T.; Parameswaran, P. S.; Tilve, S. G. J. Org. Chem. 2009, 74, 8369. b) Menezes, M. J.; Manjrekar, S.; Pai, V.; Patre, R. E.; Tilve, S. G. Indian J. Chem. 2009, 48B, 1311. c) Kamat, D. P.; Tilve, S. G.; Kamat, V. P. Tetrahedron Lett. 2012, 53, 4469. d) Parvatkar, P. T.; Ajay, A. K.; Bhat, M. K.; Parameswaran, P. S.; Tilve, S. G. Med. Chem. Res. 2013, 22, 88. e) Naik, M. M.; Tilve, S. G.; Kamat, V. P. Tetrahedron Lett. 2014, 55, 3340. f) Naik, M. M.; Kamat, D. P.; Tilve, S. G.; Kamat, V. P. Tetrahedron 2014, 70, 5221. g) Volvoikar, P. S.; Tilve, S. G. Org. Lett. 2016, 18, 892.

- Yang, T-H.; Kuo, C-W.; Kavala, V.; Konala, A.; Huang, C-Y.; Yao, C-F. *Chem. Commun.* 2017, 53, 1676.
- 5. Ravi, O.; Shaikh, A.; Upare, A.; Singarapu, K. K.; Bathula, S. R. J. Org. Chem. 2017, 82, 4422.
- 6. Sagir, H.; Rai, P.; Rahila, Tiwari, S.; Siddiqui, I. R. J. Heterocycl. Chem. 2017, 54, 397.
- 7. Wang, J.; Zhang, M-M.; Wang, X-S. Res. Chem. Intermed. 2017, 43, 2985.
- 8. Yi, X.; Xi, C. Tetrahedron 2017, 73, 1311.
- Sun, P.; Yang, D.; Wei, W.; Sun, X.; Zhang, W.; Zhang, H.; Wang, Y.; Wang, H. *Tetrahedron* 2017, 73, 2022.
- 10. Munusamy, S.; Muralidharan, V. P.; Iyer, S. K. Tetrahedron Lett. 2017, 58, 520.
- 11. Yaragorla, S.; Babu, P. V. Tetrahedron Lett. 2017, 58, 1879.
- 12. Dighe, S. U.; Samanta, S. K.; Kolle, S.; Batra, S. Tetrahedron 2017, 73, 2455.
- 13. Harnying, W.; Neudörfl, J-M.; Berkessel, A. Synthesis 2017, 49, 269.
- Yadav, J. S.; Subba Reddy, B. V.; Hara Gopal, A. V.; Narayana Kumar, G. G. K. S.; Madavi, C.; Kunwar, A. C. *Tetrahedron Lett.* 2008, 49, 4420.
- 15. Deb, M. L.; Borpatra, P. J.; Saikia, P. J.; Baruah, P. K. Synlett 2017, 28, 461.
- Buduma, K.; Chinde, S.; Arigari, N. K.; Grover, P.; Srinivas, K. V. N. S.; Kumar, K. J. Bioorg. Med. Chem. Lett. 2016, 26, 2159.
- 17. Chen, Z.; Li, H.; Dong, W.; Miao, M.; Ren, H. Org. Lett. 2016, 18, 1334.
- 18. Xu, H.; Wang, F-J.; Xin, M.; Zhang, Z. Eur. J. Org. Chem. 2016, 2016, 925.
- 19. Nguyen, T. B.; Ermolenko, L.; Al-Mourabit, A. Green Chem. 2016, 18, 2966.
- 20. Sun, J.; Qiu, J-K.; Jiang, B.; Hao, W-J.; Guo, C.; Tu, S-J. J. Org. Chem. 2016, 81, 3321.
- 21. Feng, B-B.; Lu, L.; Li, C.; Wang, X-S. Org. Biomol. Chem. 2016, 14, 2774.
- 22. Reddy, N. N. K.; Mohan, D. C.; Adimurthy, S. Tetrahedron Lett. 2016, 57, 1074.
- 23. Tiwari, A. R.; Bhanage, B. M. Org. Biomol. Chem. 2016, 14, 10567.
- 24. Inturi, S. B.; Kalita, B.; Ahamed, A. J. Org. Biomol. Chem. 2016, 14, 11061.
- 25. An, L.; Sun, X.; Lv, M.; Zhou, J.; Zhu, F.; Zhong, H. Z. Naturforsch. B. J. Chem. Sci. 2016, 71, 141.
- 26. Yang, L.; Shi, X.; Hu, B-Q.; Wang, L-X. Asian J. Org. Chem. 2016, 5, 494.
- 27. Xi, L-Y.; Zhang, R-Y.; Shi, L.; Chen, S-Y.; Yu, X-Q. Beilstein J. Org. Chem. 2016, 12, 1072.
- 28. Deb, M. L.; Pegu, C. D.; Deka, B.; Dutta, P.; Kotmale, A. S.; Baruah, P. K. Eur. J. Org. Chem. 2016, 2016, 3441.
- 29. Le, S. T.; Yasuoka, C.; Asahara, H.; Nishiwaki, N. Molecules 2016, 21, 827.
- 30. Wei, F.; Cheng, L.; Huang, H.; Liu, J.; Wang, D.; Liu, L. Sci. China Chem. 2016, 59, 1311.
- 31. Rong, H-J.; Yao, J-J.; Li, J-K.; Qu, J. J. Org. Chem. 2017, 82, 5557.
- 32. Wang, B.; Meng, Y.; Zhou, Y.; Ren, L.; Wu, J.; Yu, W.; Chang, J. J. Org. Chem. 2017, 82, 5898.

- 33. Hu, Z.; Zhao, T.; Wang, M.; Wu, J.; Yu, W.; Chang, J. J. Org. Chem. 2017, 82, 3152.
- 34. Wu, X.; Geng, X.; Zhao, P.; Zhang, J.; Gong, X.; Wu, Y-D.; Wu, A-X. Org. Lett. 2017, 19, 1550.
- Singh, D.; Kumar, V.; Devi, N.; Malakar, C. C.; Shankar, R.; Singh, V. Adv. Synth. Catal.
 2017, 359, 1213.
- 36. Miao, C-B.; Liu, R.; Sun, Y-F.; Sun, X-Q.; Yang, H-T. Tetrahedron Lett. 2017, 58, 541.
- 37. Zou, J.; Zeng, G.; Yang, R.; Yin, B. Synthesis 2017, 49, 2241.
- Kale, A.; Bingi, C.; Ragi, N. C.; Sripadi, P.; Tadikamalla, P. R.; Atmakur, K. Synthesis 2017, 49, 1603.
- 39. Zhang, J.; Gao, Q.; Wu, X.; Geng, X.; Wu, Y-D.; Wu, A. Org. Lett. 2016, 18, 1686.
- 40. Vijay, V.; Karkhelikar, M. V.; Sridhar, B.; Mirzadeh, N.; Bhargava, S.; Likhar, P. R. Org. Biomol. Chem. 2016, 14, 288.
- 41. Azimi, S. B.; Azizian, J. Tetrahedron Lett. 2016, 57, 181.
- 42. Pandit, R. P.; Kim, S. H.; Lee, Y. R. Org. Biomol. Chem. 2016, 14, 6996.
- 43. Ramesha, A. B.; Pavan Kumar, C. S.; Sandhya, N. C.; Kumara, M. N.; Mantelingu, K.; Rangappa, K. S. *RSC Adv.* **2016**, *6*, 48375.

CHAPTER 2

Synthetic studies of chromans

using molecular iodine catalyst



2.1: Introduction

Dihydrobenzopyran (chroman) ring system, more appropriately described as 3,4-dihydro-2H-1-benzopyran **1**, (Figure 1) is an oxygen heterocyclic compound having a benzene ring and a 3,4-dihydropyran ring fused together sharing a common C=C bond. It is a saturated analogue of benzopyran (chromene) constituting the basic unit in numerous naturally occurring scaffolds isolated from different sources.¹

2,2-Disubstituted chromans 2 (Figure 2) are of utmost importance as diversified analogues with a wide range of activities have been reported bearing such substitution pattern. Diverse range of compounds bearing chroman skeleton shows a broad range of biological activities. Hence it has attracted the attention of a large group of chemists. Chroman skeleton prevalently appears in important naturally occurring bioactive compounds such as vitamin E and its derivatives,² flavonoids,³ etc. In addition to biologically active natural products, several dihydrobenzopyrans are pervasive motifs in pharmaceutical drug targets exhibiting diverse activities.



Figure 1: Structure of chroman 1 with general numbering.



Figure 2: General structure of 2,2-disubstituted chromans 2.

2.2: Occurrence

Several naturally occurring biologically active dihydrobenzopyrans having substituent/s at 2-position have been isolated from diverse plant sources. Some of these chroman members along with their sources are represented below (Figures 3-6, Table 1-2).

Vitamin E **3** has been predominantly found in various sources such as corn, soybean, olive oil, palm, rice bran, and barley oils and also been isolated from *Vitis labrusca*, *Allium sativum* and *Garcinia kola*.⁴ It bears a chroman ring system with a hydrocarbon chain at 2-position. It constitutes a group of 8 naturally occurring compounds which are subdivided into four

tocopherols **3a** and four tocotrienols **3b** differing in the degree of saturation of the hydrocarbon chain. The tocopherols have fully saturated chain whereas tocotrienols have chain with 3 double bonds. Tocopherols and tocotrienols are collectively called as tocols. The 8 tocols include α -, β -, γ - and δ -tocopherols and α -, β -, γ - and δ -tocotrienols. It is well known for its role as antioxidant.^{4a} Also tocotrienols have been found to achieve potent anti-cancer effects.⁵ Tocotrienols possesses potent neuroprotective properties⁶ γ -Tocotrienol was found to be a blocker of signal transducer and activator of transcription 3 (STAT3) activation pathway and may have a potential in the treatment of cancers.⁷



Figure 3: Structures of Vitamin E 3: Tocopherols 3a and Tocotrienols 3b.



Figure 4: Structures of natural chromans 4-14.



Figure 5: Structures of natural chromans 15-26.

Table 1: Source	of isolation	of naturally	occurring	chromans 4-26
Table 1. Source	of isolation	of flaturally	occurring	Cinomans 4 -20.

No.	Name
	Source of isolation
4	δ -Garcinoic acid (δ -tocotrienolic acid) ^{4c,8}
	Garcinia kola, Clusia grandiflora, Tovomitopsis psychotriifolia, Clusia
	obdeltifolia
5	γ -Garcinoic acid (γ -tocotrienolic acid) ⁹
	Garcinia amplexicaulis
6	Garcinal ^{4c}
	Garcinia kola
7	δ -Tocotrienilic alcohol ^{8c}
	Clusia obdeltifolia
8	Polycerasoidin ¹⁰
	Polyalthia cerasoides
9	Polycerasoidol ¹⁰
	Polyalthia cerasoides
10	Racemosol ¹¹
	Bauhinia racemosa
11	de-O-Methylracemosol ^{11b}
	Bauhinia racemosa

Pseudoguttiaphenone-A ¹²
Garcinia pseudoguttifera
Vismiaguianone A ¹³
Vismia guianensis
4'-Methoxy-bavachromanol ¹⁴
Propolis
Δ^9 -Tetrahydrocannabinol ¹⁵
Cannabis sativa
(+)-Catechin ¹⁶
Streblus asper, Byrsonima crassa, Harpephyllum caffrum,
Cassia fistula, Acacia catechu, Terminalia fagifolia
3,4-Dihydromollugin ¹⁷
Rubia cordifolia
Dihydrolapachenole ¹⁸
Tabebuia chrysantha
<i>cis</i> -3,4-Dihydroxy-3,4-dihydromollugin ¹⁹
Pentas longiflora, Rubia cordifolia
trans-3,4-Dihydroxy-3,4-dihydromollugin ¹⁹
Pentas longiflora, Rubia cordifolia
Epoxymollugin ²⁰
Rubia cordifolia
Peperobtusin A ²¹
Peperomia clusiifolia, Peperomia obtusifolia
Clusifoliol/Isopeperobtusin A ^{21c-d}
Peperomia obtusifolia
3,4-Dihydro-5-hydroxy-2,7-dimethyl-8-(2"-methyl-2"-butenyl)-2-(4'-
methyl-1',3'-pentadienyl)-2H-1-benzopyran-6-carboxylic acid ^{21b,d}
Peperomia amplexicaulis, Peperomia obtusifolia
Rhododaurichromanic acid A ²²
Rhododendron dauricum
(-)-Siccanin A ²³
Helminthosposium siccans



Figure 6: Naturally occurring pyranocoumarins 27.

Table 2: Source	of isolation	of naturally	occurring	chromans	27a-r
Table 2. Source	of isofation	of naturally	occurring	cinomans	<i>⊿</i> /a-1

No.	Name	R ₁	\mathbf{R}_2
	Source of isolation		
27a	Hyuganin A ²⁴		-o <u> </u>
	Angelica furcijuga		
27b	Hyuganin B ²⁴	-0.	- 0
	Angelica furcijuga		0
27c	Hyuganin C ²⁴	-o	-0,
	Angelica furcijuga	Ö	
27d	Hyuganin D ²⁴	-o	-0, <u> </u>
	Angelica furcijuga	Ö	0 0
27e	(+)- <i>cis</i> -Khellactone ²⁴	—он	—он
	Angelica furcijuga		
27f	(-)- <i>cis</i> -3',4'-Diacetylkhellactone ²⁴	_o	-0
	Angelica furcijuga	ö	ö
27g	Anomalin ²⁴		-0 <u> </u>
	Angelica furcijuga		o O
27h	Pteryxin ^{24,25}	-o	- 0
	Angelica furcijuga, Pteryxia	Ö	
	terebinthina		
27i	Isopteryxin ²⁴		-0 <u>_</u>
	Angelica furcijuga		Ö
27ј	Isoepoxypteryxin ²⁴		-0
	Angelica furcijuga		Ö
27k	Praeroside II ²⁴	-O-Glc	—он
	Angelica furcijuga		
271	Praeroside IV ²⁴	-0-Glc	Η
	Angelica furcijuga		

27m	(+)-Lomatin ²⁴	—ОН	−H
	Angelica furcijuga		
27n	(+)-Acetyllomatin ²⁴	-0 <u> </u>	—Η
	Angelica furcijuga	Ö	
270	Suksdorfin ²⁴⁻²⁶	_0	-0 <u> </u>
	Angelica furcijuga, Lomatium	Ö	Ŏ
	suksdorfii, Arracacia nelsonii		
27p	(-)-Isosamidin ^{26b}	_0	-0
	Arracacia nelsonii	Ö	Ö
27q	Laserpitin ^{14,27}	—он	
	Propilis, Angelica keiskei, Musineon		
	divaricatum		Ū.
27r	Isolaserpitin ^{14,27}		—он
	Propilis, Angelica keiskei, Musineon		
	divaricatum		

 δ -Garcinoic acid **4** shows an antibacterial activity.^{8b} Also **4** and γ -garcinoic acid **5** are potent mPGES-1 inhibitors.⁹ δ -Garcinoic acid **4** and garcinal **6** are known for antioxidant activity.^{4c} Racemosol **10** and de-*O*-methylracemosol **11** exhibited antioxidant activities by using the scavenging of 2-2-diphenyl-1-picrylhydrazyl radical (DPPH) and FRAP total reduction capability methods wherein **11** was found to be active at all concentrations. These compounds also displayed partial cleavage of DNA.²⁸

Vismiaguianone A **13** displayed cytotoxic activity.¹³ 4'-Methoxy-bavachromanol **14** showed a strong soybean lipoxygenase inhibitory activity.¹⁴ Δ^9 -Tetrahydrocannabinol **15** showed antidepressant-like actions^{29a} and also found to induce mouse-killing behaviour (muricide).^{29b} (+)-Catechin **16** is a strong antioxidant^{16c,f} as well as possesses a potential agonist characteristic that can activate insulin receptor and peroxisome proliferator-activated receptor gamma, thus exhibiting the hypoglycemic effect of catechin.^{16d} 3,4-Dihydromollugin **17** exhibits high antioxidant activity in DPPH inhibition^{30a} and also possesses antiviral activity with an IC₅₀ value of 2.0 µg/mL in human hepatoma Hep3B cells.^{30b}

Epoxymollugin **21** isolated from *Rubia cordifolia* showed inhibition of DNA topoisomerases I and II and cytotoxic activity against MCF-7 cell line.²⁰ Peperobtusin A **22**, clusifoliol **23** and 3,4-dihydro-5-hydroxy-2,7-dimethyl-8-(2-methyl-2-butenyl)-2-(4-methyl-1,3-pentadienyl)-

2*H*-1-benzopyran-6-carboxylic acid **24** were obtained from *Peperomia obtusifolia* and exhibited potent trypanocidal activity.^{21d} Rhododaurichromanic acid A **25** is a potent anti-HIV

agent²² whereas (-)-Siccanin **26** is a potent antifungal agent.²³ Suksdorfin **270** was found to exhibit anti-HIV activity.^{26a} Also many chroman derivatives are used as antioxidants for fats and oils³¹ and some exhibit weak estrogenic activity.³²

Besides a large number of naturally occurring dihydrobenzopyrans, several synthetic analogues of chromans **28-41** have been well characterized and studied for their biological activities. Selected examples are given below (Figure 7, Table 3).



Figure 7: Structures of bioactive synthetic chromans 28-41.

Table 3:	Bioactive	synthetic	chromans	28-41 .
----------	-----------	-----------	----------	----------------

No.	Name		
28	α -Garcinoic acid (α -tocotrienolic acid) ⁹		
29	β -Garcinoic acid (β -tocotrienolic acid) ⁹		
30	6-Hydroxy-2,2-dimethyl-3,4-dihydro-2 <i>H</i> -benzo[<i>h</i>]chromene-5-		
	carboxylic acid ^{30a}		
31	Methyl 6-hydroxy-2-methyl-2-(4-methylpentyl)-3,4-dihydro-2 <i>H</i> -		
	benzo[h]chromene-5-carboxylate ^{30a}		

32	Methyl 6-hydroxy-2-phenyl-3,4-dihydro-2 <i>H</i> -benzo[<i>h</i>]chromene-5-	
	carboxylate ^{30a}	
33	Phenethyl 6-hydroxy-2,2-dimethyl-3,4-dihydro-2 <i>H</i> -benzo[<i>h</i>]chromene-	
	5-carboxylate ^{30a}	
34	Phenethyl 6-hydroxy-2-methyl-2-(4-methylpentyl)-3,4-dihydro-2 <i>H</i> -	
	benzo[h]chromene-5-carboxylate ^{30a}	
35	3',4'-Di-O-(-)-Camphanoyl-(+)-cis-Khellactone ³³	
36	Cromakalim ³⁴	
37	Trolox ³⁵	
38	MDL-73404 ³⁶	
39	Centchroman ³⁷	
40	Nebivolol ³⁸	
41	Troglitazone ³⁹	

a- and β -Garcinoic acids **28-29** are potent mPGES-1 inhibitors.⁹ Generally chromenes are more potent than their corresponding saturated analogues (chromans). Interestingly, Idhayadhulla *et al.*^{30a} synthesized different dihydromollugin analogues **30-34** which were found to exhibit little better antioxidant activity than their corresponding dehydro (chromene) compounds. Also, all these synthetic derivatives showed antibacterial activities. 3',4'-Di-*O*-(-)-camphanoyl-(+)-*cis*-khellactone **35** demonstrated anti-HIV activity.³³ Cromakalim **36** acts as an antihypertensive agent^{34a} and vasodilators.^{34b-c} It inhibits transmitter acetylcholine release by the activation of ATP-sensitive potassium channels from atrial parasympathetic nerves and cholinergic nerves in rat trachea on cells in the epithelial layer.^{34d-f} Trolox **37** is a synthetic water soluble analogue of vitamin E is widely known for its antioxidant activity.³⁵ MDL-73404 **38** is an antioxidant and also reduces myocardial infarct size.³⁶ Centchroman **39** is an antifertility agent and used as oral contraceptive pills.³⁷ Nebivolol **40** is an anti-hypertensive agent.³⁸ Troglitazone **41** inhibits serum-induced proliferation of human umbilical vascular endothelial cell (HUVEC) by suppressing casein kinase 2 activity.^{39a} It also inhibits insulin hypersecretion.^{39b-c}

2.3: Literature synthetic methods

Owing to the various biological benefits of chromans, they have attracted the attention of several chemists in recent years and numerous synthetic routes have been devised and reviewed.⁴⁰ The retrosynthetic analysis of these synthetic methods can be broadly divided into 5 categories as depicted below. These 5 approaches have been classified on the basis of i)

reaction of phenols with various electrophiles (Scheme 1, Route A), ii) intramolecular cyclization involving C-O or C-C bond forming reactions (Scheme 2, Routes B-D), iii) Diels Alder reaction between *ortho*-quinone methides intermediate and olefins (Scheme 3, Route E), iv) interconversion of functional group (Scheme 4, Route F), and v) miscellaneous routes (Scheme 5, Route G).



Scheme 1: Reaction of phenols with various electrophiles (Route A).



Scheme 2: Intramolecular cyclization involving C-O or C-C bond forming reactions (Routes B-D).



Scheme 3: Diels Alder reaction between *ortho*-quinone methides intermediate and olefins (Route E).



Scheme 4: Interconversion of functional group (Route F).



Scheme 5: Miscellaneous routes (Route G).

Diverse methodologies have been devised for chroman synthesis some of which involves the construction of the parent chroman. Asymmetric syntheses of several chroman molecules have been developed and reviewed.⁴¹ The various chroman syntheses known are discussed below.

2.3.1: Reaction of phenols with various electrophiles (Route A)

2.3.1.1: From phenols and 1,3-dienes

Most of the methodologies involves coupling of phenols and 1,3-dienes by using several homogenous or heterogenous acids as reagents/catalysts which is an atom economical way of constructing these heterocycles. These are included in the following table 4.⁴²⁻⁵⁷



Sr.	Reagent/catalyst	Reaction condition	Ref.
No.			
1)	Potassium phenoxide, AlCl ₃	benzene, rt	42
2)	PPA	xylene, 30-35 °C	43
3)	cat. ZnCl ₂ /HCl	acetic acid, 0 °C	44
4)	cat. Phosphoric acid	20 °C, 16 h	45
		or	
		xylene, 30-35 °C	
		or	
		light petroleum, 30-35 °C	
5)	cat. Aluminum phenoxide	110 °C	46
6)	cat. (+)-10-Camphorsulfonic	octane, reflux	47
	acid		
7)	cat. AlCl ₃ -[Bu ₄ N]I/AlCl ₃ -	octane/dibutyl ether, reflux	48
	$[(C_8H_{17})_4N]Br$		
8)	cat. Amberlyst 15	THF, 65-70 °C	49
9)	cat. Zeolite HSZ-360	chlorobenzene, 120 °C	50
10)	cat. [RhCl(COD)] ₂	i) cat. dppb, K ₂ CO ₃ , toluene, 110 °C	51
		ii) MeAlCl ₂ or <i>p</i> -TSA, hexane, 100°C	
11)	cat. AgOTf	ClCH ₂ CH ₂ Cl, 40 °C	52
12)	cat. AgOTf, <i>t</i> -BuCl	ClCH ₂ CH ₂ Cl, rt	53
13)	cat. Sc(OTf) ₃ –[bmim][PF ₆]	toluene, 60 / 100 °C	54
14)	cat. Cu(OTf) ₂	(bipy)/PPh ₃ , DCE, 50 °C	55
15)	cat. Bi(OTf) ₃	toluene, 40 °C	56
16)	cat. FeCl ₃ /AgBF ₄	DCE, 60 °C	57

 Table 4: Synthesis of chromans from phenols and 1,3-dienes using various reagents or catalysts.

Bolzoni *et al.*⁴² synthesized chroman in good yields by employing potassium phenoxide and AlCl₃ reagents in benzene at room temperature. Recently, Raju *et al.*⁴³ employed polyphosphoric acid in xylene to carry out this cyclization.

Several catalytic systems have also been reported for the annulation of phenols with 1,3dienes forming chromans. Smith *et al.*⁴⁴ synthesized chromans by dissolving phenol and isoprene in acetic acid in presence of dry HCl or ZnCl₂. Bader and Bean^{45a} explored phosphoric acid catalyst for the reaction of isoprene and phenol. However along with chroman product, other products like *o*- and *p*-3-methylcrotylphenols and γ -hydroxyisoamylphenols were also isolated. Later, polyhydroxyacetophenones were condensed with isoprene in phosphoric acid by Ahluwalia *et al.*^{45b-c} to form acetylchromans.

Dewhirst and Rust⁴⁶ isolated the mixture of alkenylphenols and chromans on carrying out a reaction between phenol and dienes in presence of catalytic amount of aluminium phenoxide. The catalytic activity of (+)-10-camphorsulfonic acid was investigated by Matsui and Yamamoto⁴⁷ to obtain chromans including α -tocopherol in 2 steps. However spiro compound was formed as the side product in small amounts.

Matsui and Yamamoto⁴⁸ explored the catalytic effect of AlCl₃ in presence of phase transfer catalyst such as tetraalkylammonium salts. These salts were found to accelerate Lewis acid catalyzed Friedel-Crafts alkylation reaction of phenol with 1,3-diene. Among the several tetraalkylammonium salts studied, tetrabutylammonium iodide-aluminium chloride and tetraoctylammonium bromide-aluminium chloride complexes effectively catalyzed the reaction between trimethylhydroquinone with myrecene followed by cyclization to form α -tocopherol. Kalena *et al.*⁴⁹ obtained chromans after slow addition of isoprene to the mixture of phenols and cation exchange resin amberlyst 15 in THF at 65-70 °C. However, the use of polyhydroxyphenols substrates resulted in a mixture of isomeric chroman products.

Zeolite HSZ-360 have been successfully utilized as catalyst to synthesize chromans by Bigi *et al.*⁵⁰ The one pot method selectively delivered chromans in chlorobenzene at 120 °C with traces of *o*-isopentenylphenols. However, the solid acid catalysis is also useful to obtain *o*-isopentenylphenols as major product by maintaining the temperature at 80 °C. Bienaymé *et al.*⁵¹ investigated the regioselective arylation of β -springene with trimethylhydroquinone using [RhCl(COD)]₂ catalyst followed by cyclization in presence of MeAlCl₂ or *p*-TSA resulting in chroman skeleton.

Recently, several environmentally sound catalytic systems have been reported. AgOTf was investigated to form C-C/C-O bond sequentially between phenols and dienes by Youn and Eom.⁵² The method is mild, efficient, and economical but dihydrobenzofuran ring compounds were also isolated in some cases along with the desired dihydrobenzopyrans. The hidden Brønsted acid catalyst HOTf generated from AgOTf in presence of co-catalyst *tert*-butyl chloride have been utilized for chroman synthesis by Dang *et al.*⁵³

Youn⁵⁴ developed reusable catalytic system $Sc(OTf)_3$ –[bmim][PF₆] to afford dihydrobenzopyrans along with dihydrobenzofuran ring systems. The ionic liquid [bmim][PF₆] plays dual role as an additive and an immobilizing agent for facilitating recycling of catalyst. Adrio and Hii⁵⁵ explained chroman formation from phenols to 1,3-dienes with recyclable Cu(OTf)₂ catalyst along with 2,2'-bipyridine (bipy) or PPh₃ ligands in

moderate to good yields. Both ligands were equally effective, however the cheap availability and higher resistivity of bipy to oxidation than PPh₃ makes it a preferred ligand. The method involves tandem hydroalkoxylation–rearrangement–hydroalkylation sequence. However few derivatives resulted in a mixture of dihydrobenzopyran and dihydrobenzofuran ring compounds.

Judd and Caggiano⁵⁶ explained the utility of Bi(OTf)₃ catalyst in the formation of chromans from substituted phenols and excess of isoprene. However, the products in the few examples demonstrated are obtained in low to moderate yields due to the unreacted phenol. Villani-Gale and Eichman⁵⁷ discussed the formation of regioselective prenylation of phenols followed by cyclization to chromans by employing catalytic mount of FeCl₃ and AgBF₄. The method allows employing isoprene directly and also avoids the use of stoichiometric Lewis/Bronsted acids.





Table 5: Synthesis of chromans from phenols and allylic alcohols/acetates using various reagents or catalysts.

Sr.	Reagent/Catalyst	Reaction condition	Ref.
No.			
1)	Formic acid	benzene reflux/100 °C	58
2)	BF ₃ /AlCl ₃	CH ₂ Cl ₂ , CH ₃ NO ₂	59
3)	TFA/H ₂ O	Ar, rt, 15-30 min	60
4)	ZnCl ₂	CH ₂ Cl ₂ , reflux	61
5)	cat. $(acac)_2$ Mo(SbF ₆) ₂	CH_2Cl_2, rt	62
6)	cat. $[Mo(CO)_4Br_2]_2$	CH ₂ Cl ₂ , rt	63
7)	cat. CpMoCl(CO) ₃ / <i>o</i> -chloranil	MW, 150 °C	64
8)	cat. <i>p</i> -TsOH	DCE, reflux	65
9)	cat. In(OTf) ₃	CH ₂ Cl ₂ /CH ₃ CN/CH ₃ NO ₂	66
10)	cat. CuAl-SBA-15	cyclohexane, reflux	67
11)	cat. CuAl-KIT-5-10	1,2-dichloroethane, 60-85 °C	68
12)	cat. Polyphosphoric acid	CH ₂ Cl ₂ , rt	69

13)	cat. BF ₃ .Et ₂ O	CH_2Cl_2, rt	70
14)	cat. PPh ₃ AuNTf ₂	toluene	71

Another route to chromans involves condensation of phenols with allylic alcohols/acetates in presence of Bronsted or Lewis acids and various catalysts (Table 5). Mamalis *et al.*^{58a} synthesized tocol from quinol monobenzyl ether or from quinol by adding phytol in refluxing benzene in presence of formic acid. Recently Harel *et al.*^{58b} achieved the total synthesis of encecalol angelate wherein the first step was the synthesis of chroman from resorcinol and 2-methylbut-3-en-2-ol in formic acid at 100 °C. Wehrli *et al.*⁵⁹ investigated the reaction of trimethylhydroquinone with isophytol in presence of BF₃ or AlCl₃ reagents. The reaction involved the formation of trimethylhydroquinone-boron trifluoride or trimethylhydroquinone-aluminium trichloride complex which on alkylation with isophytol followed by cyclization delivered chroman.

Ismail *et al.*⁶⁰ showed the utility of trifluoroacetic acid/H₂O combination for benzopyran synthesis. The presence of water in trifluoroacetic acid was found to accelerate the reaction. Jun and co-workers⁶¹ studied the condensation reaction of resorcinol with 2-methyl-3-buten-2-ol using several acid catalysts or acid solvents among which zinc chloride furnished chroman product in 70 % yield.

A large number of catalytic systems (Table 5) have also been devised for chroman synthesis from phenols and allylic alcohols/acetates. Various molybdenum complexes such as $(acac)_2Mo(SbF_6)_2$, $[Mo(CO)_4Br_2]_2$, and CpMoCl(CO)_3/o-chloranil have been explored. Chroman formation on treatment of prenyl alcohol or its isomer 2-methylbut-3-en-2-ol with *p*-cresol using $(acac)_2Mo(SbF_6)_2$ was studied by Malkov *et al.*⁶² However the product yields were poor. The same group also studied the catalytic activity of $[Mo(CO)_4Br_2]_2$ for the reaction between aliphatic allylic acetate or its isomer and phenol furnishing chroman.⁶³ The cyclocoupling of phenol with allylic alcohols was well explored by Yamamoto and Itonaga⁶⁴ using CpMoCl(CO)_3/o-chloranil catalytic system under microwave conditions. The chromans were obtained rapidly in moderate to good yields. The methodology was also applied for the synthesis of naturally occurring α -tocopherol derivatives and 3,4-dihydromollugin.

Several chroman derivatives were obtained by Ishino *et al.*⁶⁵ from phenols and unsaturated alcohols by employing catalytic amount of *p*-toluenesulfonic acid in 1,2-dichloroethane solvent. The cyclocoupling of phenols and allylic acetates was also achieved by Vece *et al.*⁶⁶ with indium triflate catalyst. The efficiency of catalyst can be easily identified as the reaction smoothly proceeded with just 1 mol% of the catalyst. It involves tandem allylation–

intramolecular hydroalkoxylation sequence. However noncyclised *o*- and *p*-allylated products and dihydrobenzofuran in some cases were also recovered along with the desired chroman. Varghese *et al.*⁶⁷ utilized mesoporous and hexagonally ordered CuAl-SBA-15 catalyst for chroman synthesis in good yields from phenols and allylic alcohols. The recyclable catalyst allows tandem C-C and C-O bond formation leading to several chroman derivatives including vitamin E. The same group⁶⁸ also showed the scope of CuAl-KIT-5-10 catalyst to obtain chromans from phenols and 2-methylbut-3-en-2-ol at 85 °C. However, a mixture of *o*prenylated phenol and chromans were obtained on decreasing the temperature to 60 °C.

Polyphosphoric acid was employed for the construction of benzopyran skeleton by Murthy *et al.*⁶⁹ It was further functionalized to obtain several novel compounds which were evaluated for antimicrobial activities. Madabhushi *et al.*⁷⁰ extended the scope of methodology employing BF₃.Et₂O catalyst for chroman synthesis wherein 2-methylbut-3-en-2-ol was treated with *p*-cresol to yield chroman in 80 %. However on changing the unsaturated alcohol to 1,1-diphenylprop-2-en-1-ol no product was isolated. Coutant *et al.*⁷¹ demonstrated cyclocoupling of phenols and allylic alcohols using catalytic amount of PPh₃AuNTf₂. The method involves a one-pot regioselective Friedel–Crafts allylation reaction followed by intramolecular hydroalkoxylation. A large range of derivatives have been synthesized.



Scheme 6

Miller and Wood⁷² synthesized 2,2-dimethylchroman derivatives by the treatment of 3,3dimethylallyl diphenyl phosphates with an excess of phenols at 120 °C in absence or presence of solid sodium hydrogen carbonate (Scheme 6).

2.3.1.3: From phenols and allylic halides



Table 6: Synthesis of chromans from phenols and allylic halides using various reagents or catalysts.

Sr.	Reagent/ Catalyst Reaction condition		Ref.
No.			
1)	BuLi	benzene, reflux	73

2)	Montmorillonite K-10	CCl ₄	74
3)	cat. $Ni(acac)_2$	90 °C	75
4)	cat. <i>p</i> -TSA	toluene, reflux	76

Chromans have also been obtained by the cyclization of phenols with allylic bromides/chlorides using some reagents/catalysts (Table 6). Cardillo *et al.*⁷³ obtained chromans by treating lithium salts of phenol obtained from phenol and butyllithium with prenyl bromide in refluxing benzene. The products were dehydrogenated further with DDQ to form chromenes. Similarly, Dintzner *et al.*⁷⁴ demonstrated cyclocondensation of substituted phenols with prenyl bromide to afford 2,2-dimethylbenzopyrans using montmorillonite K-10. However mixture of regioisomers was obtained in few cases. Camps *et al.*⁷⁵ carried out the cycloaddition of 4-methoxyphenol and prenyl chloride to produce 2,2-dimethyl-6-methoxychroman using Ni(acac)₂ catalyst at 90 °C. Jetter *et al.*⁷⁶ observed the reaction of 7-hydroxycoumarins with allyl halides, homoallyl halides or alcohols in presence of catalytic amount of *p*-toluenesulphonic acid. It delivered chromans/reduced derivatives of naturally occurring xanthyletin and seselin *via* a one step alkylation followed by cyclization.

2.3.1.4: From phenols and 1,3-dichloro-3-methylbutane





Camps *et al.*⁷⁵ also explored the effectiveness of $Ni(acac)_2$ catalyst to obtain chroman product from phenol and 1,3-dichloro-3-methylbutane. Different derivatives were obtained, some of which were accompanied by overalkylation (Scheme 7).

2.3.2: Intramolecular cyclization involving C-O or C-C bond forming reactions (Routes B, C and D)

2.3.2.1: From aryl prenyl ethers (Route B)



Table 7: Synthesis of chromans from aryl prenyl ethers using various reagents or catalysts.

Sr.	Reagent/Catalyst	Reaction condition	Ref.
No.			
1)	Montmorillonite clay	benzene, 50 °C, N ₂	77

2)	Calcined SiO ₂	neat, 60-70 °C or benzene, reflux	78
3)	cat. Mo(CO) ₆	toluene, 115 °C, Ar	79
4)	cat. Bi(OTf) ₃ ·4H ₂ O	toluene, 0-22 °C	80

Rearrangement of aryl prenyl ethers is an alternative route to obtain chromans (Table 7). Dauben *et al.*⁷⁷ carried out rearrangement of aryl prenyl ethers using montmorillonite clay *via* [1,3] shift of ethers followed by cyclization to synthesize chromans. However, some substrates delivered *o*-allyl phenols and coumarans. Similarly aryl prenyl ether was rearranged with calcined SiO₂ (300 °C, 5 h) by Pogrebnoi *et al.*⁷⁸ either under neat heating or refluxing benzene conditions.

Bernard *et al.*⁷⁹ showed the usefulness of $Mo(CO)_6$ catalyst in carrying out Claisen rearrangement of aryl prenyl ethers followed by cyclization leading to chromans. In some cases, the products were accompanied with small amounts of corresponding starting phenols. Later, Ollevier and Mwene-Mbeja⁸⁰ employed bismuth triflate catalyst for [1,3] rearrangement of aryl prenyl ethers to synthesize chromans. However, *p*-prenylphenols are isolated as side products.



Scheme 8

Barluenga *et al.*⁸¹ found the selective rearrangement of allylphenyl ethers followed by cyclization to 2-substituted 3-iodochromans using IPy_2BF_4 which promotes C-C bond formation and also serves as a source of iodonium ion. Also isomeric 4-substituted 3-iodochromans were obtained *via* an intramolecular cyclization reaction (Scheme 8).



Scheme 9

The tertiary ether was chemoselectively hydroborated with catecholborane in the presence of 2 % Wilkinson's catalyst by Trost and Toste⁸² which underwent electrophilic cycloalkylation to chroman upon activation of the primary alcohol as its triflate (Scheme 9).

2.3.2.2.1: From 2-(3-hydroxyalkyl)phenols/anisoles (Route C)

Table 8: Synthesis of chromans from 2-(3-hydroxyalkyl)phenols using various reagents or catalyst.



Sr.	Reagent/Catalyst	Reaction condition	Ref.
No.			
1)	DEAD, triphenylphosphine	dioxane, rt	83
2)	H ₃ PO ₄	toluene, reflux	84
3)	H_2SO_4	toluene	85
4)	cat. <i>p</i> -TsOH	benzene, reflux	86

Dehydration of substituted 2-(3-hydroxyalkyl)phenols also delivers chromans (Table 8). Aristoff *et al.*⁸³ carried out an intramolecular dehydration of substituted 2-(3-hydroxyalkyl)phenols to the corresponding extremely potent benzopyran prostacyclin mimics through an intramolecular Mitsunobu reaction as the key step. In addition, few structurally simple substrates on treatment with DEAD and triphenylphosphine in dioxane delivered chromans and dihydrobenzofurans in moderate to excellent yields. Only few examples reported suggest the limited scope of the method. Yus *et al.*⁸⁴ carried out acidic intramolecular dehydration of 2-(3-hydroxy-3-substituted)phenols to chromans using 85 % phosphoric acid in refluxing toluene. However substrate scope is limited. Erhardt and coworkers⁸⁵ used 15 % H₂SO₄ in toluene for dehydration of diol. Chroman obtained was further functionalized over several steps to accomplish synthesis of complex molecule. Tanaka *et al.*⁸⁶ obtained 3,4-dihydro-2,2,6-trimethyl-2*H*-1-benzopyran from the corresponding 2-(3-hydroxy-3-methylbutyl)-4-methylphenol using *p*-toluenesulfonic acid as catalyst.



Scheme 10

Demethylative cyclisation of substituted *o*-methoxy tertiary alcohols was carried out by Nilsson *et al.*⁸⁷ with HBr in acetic acid condition to furnish chromans (Scheme 10).

2.3.2.2.2: From 2-(3',3'-dialkylallyl)phenols/anisoles/phenyl acetates (Route C)

Table 9: Synthesis of chromans from 2-(3',3'-dialkylallyl)phenols using various reagents or catalysts.



Sr.	Reagent/Catalyst	Reaction condition	Ref.
No.			
1)	Phenol (excess)	100 °C	72
	diphenyl hydrogen phosphate		
	sodium hydrogen carbonate		
2)	HCl/AlC1 ₃	cyclohexane, rt	88
3)	cat. FeCl ₃	25 °C	89
4)	cat. Al(O <i>i</i> Pr) ₃	PhCl, 250 °C, MW	90
5)	cat. Fe-montmorillonite/ Bi-	dimethyl carbonate, 80 °C	91
	montmorillonite		

2-(3',3'-Dialkylallyl)phenols have also been converted to the respective chromans (Table 9). 2-(3',3'-Dimethylallyl)phenol, phenol, diphenyl hydrogen phosphate, and sodium hydrogen carbonate were heated at 100 °C by Miller and Wood⁷² to form chroman products. Later, Wang *et al.*⁸⁸ obtained chromans by the intramolecular cyclization of 2-(3-methyl-2buteny1)phenols along with 1,1-dimethyl-4-indanols. Also the substrate scope is limited. The cyclization of 2-(3',3'-dimethylallyl)phenol derivatives to the corresponding chromans using catalytic FeCl₃ was carried out by Macone *et al.*⁸⁹ The products were evaluated for cytotoxicity and pro-apoptotic activity.

Later, Schlüter *et al.*⁹⁰ converted 2-allylphenols to cyclic ethers *via* intramolecular hydroxylation using cat. aluminium isopropoxide under microwave irradiation. The scope of the catalyst was extended by synthesising chroman skeleton from 2-(3',3'-dimethylallyl)phenol. Recently, Francesco *et al.*⁹¹ employed recyclable heterogeneous catalyst Fe/Bi-Montmorillonite in dimethyl carbonate solvent to achieve chromans. In addition, a wide range of cyclic ethers were produced.



Scheme 11

Hurd and Hoffman⁹² prepared chroman from 2-(3',3'-dimethylallyl)phenol upon treatment with ketene and trace amount of sulfuric acid along with minor acetylation product *viz. o-(\gamma, \gamma*-dimethylallyl)phenyl acetate. However when the acetate product separately treated with hydrogen bromide, 2,2-dimethylchroman was produced (Scheme 11).



Scheme 12

Jiménez *et al.*⁹³ carried out irradiation of trans-2-cinnamylphenol using various conditions to deliver 2-phenylchroman along with cis-2-cinnamylphenol and 2-benzyl-2,3-dihydrobenzofuran (Scheme 12).



Scheme 13

Nicolaou *et al.*⁹⁴ employed a selenium-based methodology for the solid phase construction of resin bound 2,2-dimethylbenzopyrans wherein the loading step is a key ring-forming reaction. A library of derivatives has been synthesized (Scheme 13).



Scheme 14

Boltze and Dell⁹⁵ on heating 2-(3-methyl-2-butenyl)resorcinol dimethyl ether with pyridinium chloride at 210-220 °C for 5 h delivered the desired chroman skeleton (Scheme 14).



Scheme 15

Gopalakrishnan *et al.*⁹⁶ employed AlCl₃/EtSH reagent to carry out a tandem demethylationcyclisation reaction to form pyran rings (Scheme 15).

2.3.2.2.3: From substituted 2-(3-butenyl)phenols (Route C)



Scheme 16

Bravo and Ticozzi⁹⁷ converted the olefin starting to the epoxide using *m*-chloroperbenzoic acid which was converted to chroman by the attack of the phenolic oxygen on the quaternary epoxide carbon in epoxide intermediate during work up (Scheme 16).



Scheme 17

Chroman derivative was synthesized by Sato *et al.*⁹⁸ by treating the *o*-alkylphenol starting with boron trifluoride etherate followed by desulphurization with Raney nickel (Scheme 17).





Exo-methylene chromans were obtained by Grigg *et al.*⁹⁹ along with dihydrobenzofurans through cyclization of dienylmethyl phenols with *p*-toluenesulphonic acid and trace amount of TFA (Scheme 18).



Scheme 19

According to Stoltz and co-workers¹⁰⁰ 5 mol% of Pd(TFA)₂ catalyst in presence of pyridine and Na₂CO₃ using molecular oxygen as oxidant was effective in constructing chroman skeleton *via* aerobic oxidative cyclization of oxygen nucleophile onto olefin (Scheme 19).



Scheme 20

Pd-catalyzed carboetherification reactions between 2-(but-3-en-1-yl)phenols and aryl/alkenyl halides was described by Ward *et al.*¹⁰¹ Several 2-substituted and 2,2-disubstituted derivatives have been prepared (Scheme 20).

2.3.2.3: From 3-(2-bromoaryl)-1-propanols/3-aryl-1-propanols (Route D)



 Table 10: Synthesis of chromans from 3-(2-bromoaryl)-1-propanols using various reagents or catalysts.

Sr.	Reagent/Catalyst	Reaction condition	Ref.
No.			
1)	t-BuOK	DMSO, MW, 140 W	102
2)	cat. $Pd(OAc)_2$, Tol-	K ₂ CO ₃ /NaOt-Bu, toluene, 80-100 °C	103
	BINAP/DPPF		
3)	cat. Pd(OAc) ₂ , 2-(di- <i>tert</i> -	Cs ₂ CO ₃ , toluene, 50-80 °C	104
	butylphosphino)-1,1'-		
	binaphthyl/ 2-di-tert-		
	butylphosphinobiphenyl		
4)	cat. $Pd(dba)_2$, $Ph_5FcP(t-Bu)_2$	toluene, rt	105
5)	cat. CuI, 8-hydroxyquinoline	Cs ₂ CO ₃ , toluene, Ar atm, 110 °C	106
6)	cat. CuI, 2,2-bipyridyl	KOt-Bu, DMF, 120 °C	107

Intramolecular coupling of aryl bromides with alcohol is another way to obtain chromans (Table 10). Xu *et al.*¹⁰² investigated an efficient aromatic C–O bond forming reaction using potassium *tert*-butoxide under microwave irradiation. Diverse products were synthesized including chromans avoiding the use of transition-metal catalyst. The method is also useful for the synthesis of bioactive natural flavans.

Palladium catalysis in presence of various ligands is also well known for such cyclization reactions. Palucki *et al.*¹⁰³ studied the role of Pd(OAc)₂ catalyst in presence of (*S*)-(-)-2,2'- bis(di-*p*-tolylphosphino)-1,1'-binaphthyl (Tol-BINAP) and 1,1'-bis(diphenylphosphino)-ferrocene (DPPF) in the synthesis of chromans. Cyclization of intramolecular substrates was carried out in toluene in presence of NaOt-Bu or K₂CO₃ base. However, the reactions were accompanied by side products including dehalogenation of the aryl halides and oxidation of the alcohol to the ketone in case of substrates containing a secondary alcohol.

Later, to overcome the above problems, Torraca *et al.*¹⁰⁴ explored 2-(di-*tert*-butylphosphino)-1,1'-binaphthyl and 2-di-*tert*-butylphosphinobiphenyl as ligands, the former being more general, in presence of $Pd(OAc)_2$ catalyst. The products were obtained in good yields. Shelby *et al.*¹⁰⁵ successfully explored a combination of cat. $Pd(dba)_2$ with $Ph_5FcP(t-Bu)_2$ ligand which is effective at room temperature for aromatic C-O bond formation reactions. Two chromans were synthesized in good yields along with several acyclic ethers.

Copper catalysis is also effective to render chromans. Niu *et al.*¹⁰⁶ developed a methodology using copper (I) iodide catalyst, 8-hydroxyquinoline ligand and K₃PO₄ base. The method involves C-O coupling between aliphatic alcohols and aryl bromides to form acyclic ethers. Further intramolecular coupling resulted in chromans and benzofurans in presence of Cs₂CO₃ base. Suchand *et al.*¹⁰⁷ also reported copper catalyzed intramolecular C-O bond formation reactions of secondary and tertiary alcohol substrates in presence of 2,2-bipyridyl ligand. Several 2-substituted and 2,2-disubstituted chroman derivatives were synthesized in good yields.



Scheme 21

Furuyama and Togo¹⁰⁸ irradiated various 3-aryl-1-propanols with excess of 1,3-diiodo-5,5dimethylhydantoin (DIH) with tungsten lamp in presence of ethyl acetate or 1,2dichloroethane to produce chroman derivatives. Products were obtained in moderate to good yields (Scheme 21).



Scheme 22

Hamamoto *et al.*¹⁰⁹ utilized phenyliodine(III)bis(trifluoroacetate) (PIFA), a hypervalent iodine (III) reagent for the direct aromatic carbon–oxygen bond forming reaction of 3-aryl-1-propanols leading to chromans. It involves an aromatic cation radical pathway (Scheme 22).



Scheme 23



Figure 8: Structures of QuCN⁺ and Co(dmgBF₂)₂(CH₃CN)₂.

Recently, Zheng *et al.*¹¹⁰ demonstrated the scope of a dual catalytic system consisting of 3cyano-1-methylquinolinum as a photocatalyst and $Co(dmgBF_2)_2(CH_3CN)_2$ as a cocatalyst for the chroman synthesis. It allows the direct intramolecular alkoxylation of 3-aryl-1-propanols with the evolution of hydrogen gas as the only byproduct (Scheme 23, Figure 8).

2.3.3: Diels Alder reaction between *ortho*-quinone methides intermediate and olefins (Route E)



Scheme 24

Chiba *et al.*¹¹¹ carried out hetero-Diels–Alder reaction of *o*-quinomethanes and unactivated alkenes using montmorillonite catalyst in a LiClO₄–MeNO₂ solution (Scheme 24).



Scheme 25

Yadav *et al.*¹¹² reported an efficient scandium triflate catalyst for the cyclocondensation of *o*-hydroxybenzaldehydes with olefins in the presence of trimethyl orthoformate to form chromanes with high diastereoselectivity (Scheme 25). It involves *o*-quinone methides intermediate generated from salicylaldehyde and trimethyl orthoformate.



Scheme 26

Nakamura *et al.*¹¹³ synthesized chroman by heating 4H-1,2-benzoxazine with styrene in toluene. It involves Diels-Alder reaction between *in situ* formed *o*-benzoquinone methide intermediate with styrene (Scheme 26).



Scheme 27

Radomkit *et al.*¹¹⁴ developed a method for the preparation of chromans through generation of *ortho*-quinone methides intermediate which underwent $PtCl_4$ catalysed cycloaddition reaction with olefins. Also the conversion takes place in presence of *p*-TsOH immobilized on silica (PTS-Si) however in low yields (Scheme 27).

2.3.4: Interconversion of functional group (Route F)



Reagent/Catalyst Reaction condition Ref. Sr. No. 1) i) EtMgBr (excess) i) ether, reflux 115 ii) recrystallization i) MeMgI (excess) i) ether, rt 2) 116 ii) H_2SO_4/H_2O ii) reflux i) MeMgCl (excess) i) THF. 0 °C - rt 3) 117 ii) benzene, 105 °C ii) H₂SO₄/H₂O i) MeLi (excess) i) THF, 0 °C - rt 4) 118 ii) cat. TsOH ii) toluene, reflux

Table 11: Synthesis of chromans from coumarin using various reagents or catalysts.

Dihydrocoumarin has been converted to chromans (Table 11). Smith *et al.*¹¹⁵ treated dihydrocoumarin with Grignard reagent to form carbinol followed by recrystallization which resulted in the cyclization leading to chromans. Later, Fatope and Abraham¹¹⁶ and Teng *et al.*¹¹⁷ also exposed dihydrocoumarin to Grignard reagent forming tertiary alcohol which underwent cyclization in presence of aqueous H₂SO₄. Further, Bernier and Brückner¹¹⁸

converted dihydrocoumarin to chroman using MeLi followed by cyclodehydration with TsOH.



Scheme 28

Chroman has also been synthesized from chromanone by Clemmensen reduction¹¹⁹ (Scheme 28).



Anioł *et al.*¹²⁰ converted chromene to chroman by reducing with sodium, however in low yields as several other products were also obtained (Scheme 29).

2.3.5: Miscellaneous routes (Route G)



Scheme 30

Webb and Hall¹²¹ showed the formation of chroman in TFA from 2,6-diphenyl phenol and acetone. It involves the reversible formation of the phenol hemiacetal of acetone followed by its protonation and loss of water forming carbonium ion which then attacks either of the phenyl groups to form chroman (Scheme 30).



Scheme 31

Verhé *et al.*¹²² heated 2-chloro-2-(3-methyl-2-butenyl)-1,3-cyclohexanedione in DMF in presence of HCl at 140 °C which rearranged to the 4-chloro isomer. It then underwent HCl elimination to form 2-(3-methyl-2-butenyl)resorcinol which cyclized to produce 2,2-dimethyl-5-hydroxychroman (Scheme 31).



Bravo *et al.*¹²³ obtained chromans, however in low yields by reacting the phenolic substrate with dimethyloxosulphoxonium methylide in DMSO/THF. However, the reaction favoured the formation of 7-membered 3-hydroxy-2,3,4,5-tetrahydro-1-benzo-oxepin as major product (Scheme 32).



Scheme 33

De Renzi *et al.*¹²⁴ treated phenol with 1,1-dimethylallene in presence of platinum catalyst to afford a mixture of 2,2-dimethylchroman and *o*-isopentenylphenol. The amount of the products varied depending upon the temperature, reaction time and phenolic substrate. Chromans are formed *via* a regiospecific C-alkenylation followed by cyclization (Scheme 33).



Scheme 34

Larock *et al.*¹²⁵ carried out palladium catalyzed annulations of *o*-iodophenols with 1,4-dienes to produce chromans in good yields. It involves heating the starting materials with catalytic palladium acetate and base in presence or absence of PPh₃ for prolonged time (Scheme 34).



Scheme 35

Buchwald and co-workers¹²⁶ treated aryl bromide complex (dppf)Pd[o-C₆H₄(CH₂)₂C(Me)₂OH]Br with KH in THF at room temperature forming oxapalladacycle (dppf)Pd[o-C₆H₄(CH₂)₂C(Me)₂O] which produced chroman on thermolysis (Scheme 35).



Scheme 36

Knight and Little¹²⁷ treated the starting alcohol with *N*-iodosuccinimide to generate 8iodochroman *via* benzyne intermediate. Only few derivatives have been reported (Scheme 36).



Scheme 37

2,4-Dimethoxy-2-methylbenzopyrans were synthesized by Yadav *et al.*¹²⁸ by the cyclocondensation of *o*-hydroxybenzaldehydes with 2,2-dimethoxypropane at room temperature using scandium triflate catalyst. Several advantages associated with the methodology includes high yields, high diastereoselectivity, short reaction times, easy availability of starting materials, reusable catalyst and simple experimental/isolation procedures which makes it an useful method (Scheme 37).

2.4: Results and Discussion

Several syntheses of chromans have been reported as discussed above. However these methods make use of either hazardous or costly catalysts, associated with side products resulting in poor yields, requires additional preparation of substrates and lack of substrate scope which led us to develop a new synthetic methodology for chromans synthesis using an environmentally benign inexpensive catalyst. We envisioned that the Lewis acidity of iodine can be employed for its synthesis. Accordingly the retrosynthetic analysis of this one pot approach suggested simple substrates such as phenols **42** and unsaturated alcohols **43** (Scheme 38).



Scheme 38: Retrosynthetic analysis of chroman 2.

We began chroman synthesis from β -naphthol **42a** and prenyl alcohol **43a** and examined the effect of iodine catalyst (0.3 equiv) on its cyclization in chloroform at room temperature. After prolonged stirring it was pleasing to see a new spot which was further isolated as colorless solid and characterized as the desired chroman **2a**. On refluxing, the reaction time was reduced and product formation was observed within short period (1 h) (Scheme 39). The reaction was also accompanied by another very light spot (TLC) and was later characterized as **2a**'. The structures of **2a** and **2a**' were confirmed by the following spectral data.



Scheme 39: Reaction of β -naphthol with prenyl alcohol.

Spectral data of 3,3-dimethyl-2,3-dihydro-1H-benzo[f]chromene (2a)



colorless solid; m.p. 68-70 °C; lit.⁶⁴ 68-69 °C.

IR (**KBr**): $\tilde{v} = 2974$, 1620, 1597, 1236 cm⁻¹.

¹**H NMR** (**CDCl**₃, **400 MHz**): δ 1.38 (s, 6 H), 1.95 (t, *J* = 6.8 Hz, 2 H), 3.03 (t, *J* = 6.8 Hz, 2 H), 7.02 (d, *J* = 8.8 Hz, 1 H), 7.32 (ddd, *J* = 8.0, 7.6, 0.8 Hz, 1 H), 7.47 (ddd, *J* = 8.4, 6.8, 1.2 Hz, 1 H), 7.61 (d, *J* = 8.8 Hz, 1 H), 7.75 (d, *J* = 8.0 Hz, 1 H), 7.82 (d, *J* = 8.4 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 19.3 (CH₂), 26.6 (2 X CH₃), 32.7 (CH₂), 74.0 (Cq), 112.4 (Cq), 119.8 (CH), 121.9 (CH), 122.9 (CH), 126.2 (CH), 127.7 (CH), 128.4 (CH), 128.7 (Cq), 133.1 (Cq), 151.3 (Cq).

Spectral data of 2-iodo-3,3-dimethyl-2,3-dihydro-1H-benzo[f]chromene (2a')



colorless solid; m.p. 108-110 °C.

Rf: 0.56 (5 % ethyl acetate/petroleum ether).

IR (KBr): $\tilde{v} = 2974$, 1622, 1597, 1236 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.45 (s, 3 H), 1.57 (s, 3 H), 3.55-3.75 (m, 2 H), 4.51 (dd, *J* = 9.6, 6.0 Hz, 1 H), 6.96 (d, *J* = 9.2 Hz, 1 H), 7.29 (t, *J* = 7.2 Hz, 1 H), 7.43 (t, *J* = 7.2 Hz, 1 H), 7.60 (d, *J* = 9.2 Hz, 1 H), 7.67 (d, *J* = 8.8 Hz, 1 H), 7.71 (d, *J* = 8.0 Hz, 1 H).

¹³C NMR (DMSO, 100 MHz): δ 24.3 (CH₃), 26.6 (CH₃), 33.1 (CH₂), 34.3 (CH), 76.1 (Cq), 111.5 (Cq), 119.0 (CH), 121.9 (CH), 123.4 (CH), 126.6 (CH), 128.2 (2 X CH), 128.5 (Cq), 131.9 (Cq), 149.6 (Cq).

Anal. Calcd. For C₁₅H₁₅IO: C, 53.3; H, 4.4 %; Found: C, 53.4; H, 4.5 %.

Next, we went on to examine the stoichiometry of the substrates (Table 12). First, the stoichiometry of substrate **42a** was varied from 1-5 equivalents. It was observed that the yield goes on increasing with increasing β -naphthol concentration up to 4 equivalents (entries 1-4),

however, further increasing concentration to 5 equivalents resulted in decreased product yield (entry 5). Hence 4 equivalents of β -naphthol was found to be the optimum concentration to give maximum yield of 92 % after 4 h (entry 4). On the other side, increasing the concentration of prenyl alcohol did not show any increase in product yield, moreover it led to a complex mixture. Hence, further optimization was carried out with 4 equivalents of β -naphthol.

Table 12: Optimization of stoichiometry of starting materials 42a and 4	3a
---	----

OH +	\rightarrow	0.3 equiv iodine	0
40-	/OH	chloroform, reflux	
42a	40a		💛 2a

Sr. No.	42a (equiv)	43a (equiv)	Time (h)	Yield (%) ^{a,b}
1)	1	1	4	78
2)	2	1	4	80
3)	3	1	4	88
4)	4	1	4	92
5)	5	1	4	85

^a Isolated yields of **2a** based on substrate **43a**.

^b Trace amount of **2a'** was formed.

Further optimization of iodine concentration was studied by varying the amount of iodine (Table 13). No product was formed in the absence of iodine (entry 1). When 10 mol% iodine was used, **2a** was formed in 85 % yield (entry 2) and the product yield was found to increase with increase in catalyst loading and with reduction in time (entries 3-4). 30 Mol% of iodine in refluxing chloroform was found to be the optimum concentration (entry 4). Further increase in iodine concentration resulted in decreased product yield (entry 5). When the reaction was carried out with stoichiometric amount of iodine at room temperature with 1:1 ratio of **42a**:**43a**, the formation of mixture of 3,3-dimethyl-2,3-dihydro-1*H*-benzo[*f*]chromene **2a** and 2-iodo-3,3-dimethyl-2,3-dihydro-1*H*-benzo[*f*]chromene **2a**' was observed. As **2a**' was formed in appreciable amount, it was purified further to obtain as colorless solid and characterized at this stage.
+ 42a (4 equiv)	Hard Hard Hard Hard Hard Hard Hard Hard		
Sr. No.	Iodine (mol%)	Time (h)	Yield (%) ^a
1)	0	24	0
2)	10	24	85
3)	20	24	89
4)	30	4	92
5)	40	2	79
6) ^b	100	1.3	58, 31 ^c

Table 13: Optimization of iodine concentration in refluxing chloroform.

^a Isolated yields of **2a** based on substrate **43a**.

^bReaction carried out at room temperature.

^c% yield of **2a'**.

Thus the yield of the reaction product can be modulated by changing the concentration of iodine. Catalytic iodine furnished chroman **2a** as the major product with trace amount of **2a**' under chloroform reflux whereas stoichiometric amount of iodine resulted in increased quantity of **2a**' in chloroform solvent at room temperature along with **2a**.

Next, the effect of solvents was studied by carrying out the reaction in different solvents (Table 14). The product formation was seen in all the studied solvents (entries 1-9). The time required for product formation was more when refluxed in protic solvents like methanol and ethanol (entries 1-2). Chlorinated solvents such as chloroform and dichloromethane gave better results with less duration of time (entries 3-4), of which chloroform gave maximum product yield (entry 3). Other solvents like toluene, tetrahydrofuran, 1,4-dioxane, 1,2-dichloroethane and acetonitrile also showed product formation but in moderate yields (entries 5-9). Among all these solvents, chloroform gave highest product yield and hence was selected for further studies.

Table 14: Solvent screening.



Sr. No.	Solvent	Time (h)	Yield (%) ^a
1)	Methanol	24	50
2)	Ethanol	24	45
3)	Chloroform	4	92
4)	Dichloromethane	4	90
5)	Toluene	24	30
6)	Tetrahydrofuran	7	65
7)	1,4-Dioxane	7	60
8)	1,2-Dichloroethane	5	68
9)	Acetonitrile	24	25

^a Isolated yields based on substrate **43a**.

Table 15: Synthesis of various chroman derivatives **2a-j**, **18** using phenol derivatives **42** andprenyl alcohol **43a** under optimized reaction condition.



Sr.	Substituted Phenol	Chroman	Solvent	Time	Yield ^a
No.			(reflux)	(h)	(%)
1)	42a		CHCl ₃	4	92
		2a			
2)	ОН		CHCl ₃	4	48
	42b	2b			
3)	MeO OH MeO	MeO MeO	CHCl ₃	2	77
	42c	2c			
4)	МеО	MeO	CHCl ₃	4	58
	42d	2d			
	MeO	MeO			

5)	42e	2e	CHCl ₃	4	45
6)	O → OH 42f	2f	CHCl ₃	4	58
7)	ОН	MeO	CHCl ₃	2	60
	42g	18			
8)	HOUDH	HO	МеОН	4	46
	42h	2g			
9)	НО	HOHO	МеОН	4	33
	42i	2h			
10)	HOOHOH	HO O OH	МеОН	4	56
	42j	2i			
11)	CI	CI	CHCl ₃	24	20
	42k	2 j			

^a Isolated % yield of chroman product based on substrate **43a**.

Unreacted and excess phenol substrates were recovered.

Table 16: Synthesis of chroman derivatives **2a**, **k**, **l** using β -naphthol **42a** and unsaturated alcohols **43b-d** under optimized reaction conditions.

$\begin{array}{cccc} & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & $					
Sr.	Unsaturated alcohol	Chroman	Solvent	Time	Yield ^a
No.			(reflux)	(h)	(%)
1)	он 43b		CHCl ₃	4	87
		2a			

2)	43с	2k	CHCl ₃	4	76
3)	Ph OH 43d	2l	CHCl ₃	4	60

^a Isolated % yield of chroman product based on unsaturated alcohol substrate **43b-d**. Unreacted and excess β -naphthol substrate was recovered.

Once the optimum condition was established, the generality of this reaction was explored by subjecting various phenols to the optimum reaction condition (Table 15-16). The reaction of β -naphthol **42a** with prenyl alcohol **43a** gave chroman **2a** in 92 % yield. Electron rich phenols with methyl and methoxy substituents were converted to respective chromans **2b-e** in good to moderate yields. Similarly 3,4-methylenedioxyphenol underwent cyclization readily to afford chroman **2f**. Naturally occurring dihydrolapachenole **18**¹⁸ was synthesized from 4-methoxynaphthol in good yield. Dihydroxy compounds like resorcinol and quinol when refluxed in methanol due to lack of solubility in chloroform solvent, afforded respective chromans **2g** and **2h** in moderate yields. Interestingly, the resorcinol substrate which may result in mixture of isomeric chromans delivered exclusively one regioisomer **2g**. Further the trihydroxy compound phloroglucinol in refluxing methanol afforded chroman **2i** in 56 % yield. *p*-Chlorophenol was converted to corresponding chroman **5k** in low yield. The slow reaction and low yield is attributed to the electron withdrawing effect of chloro group.

Also, we examined the reaction of β -naphthol **42a** with different unsaturated alcohols such as 2-methylbut-3-en-2-ol **43b**, phytol **43c** and cinnamyl alcohol **43d** to afford desired chromans **2a**, **2k** and **2l** in 87, 76 and 60 % yields respectively.

Spectral data of chromans 6-Methyl-2,2-dimethyl-3,4-dihydro-2*H*-1-benzopyran (2b)

2b

yield (0.099 g, 48 %); colorless oil.⁶⁴ **IR (neat):** $\tilde{v} = 2976$, 1498, 1261, 1105 cm⁻¹. ¹H NMR (CDCl₃, 400 MHz): δ 1.23 (s, 6 H), 1.69 (t, J = 6.8 Hz, 2 H), 2.16 (s, 3 H), 2.64 (t, J = 6.8 Hz, 2 H), 6.60 (d, J = 8.0 Hz, 1 H), 6.78 (s, 1 H), 6.80 (d, J = 8.4 Hz, 1 H).
¹³C NMR (CDCl₃, 100 MHz): δ 19.4 (CH₃), 21.4 (CH₂), 25.8 (2 X CH₃), 31.8 (CH₂), 72.8 (Cq), 115.9 (CH), 119.5 (Cq), 126.9 (CH), 127.6 (Cq), 128.7 (CH), 150.6 (Cq).

6,7-Dimethoxy-2,2-dimethyl-3,4-dihydro-2*H*-chromene (2c)



yield (0.199 g, 77 %); colorless oil.¹²⁹

IR (neat): $\tilde{v} = 2974, 2933, 1620, 1514, 1122 \text{ cm}^{-1}$.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.24 (s, 6 H), 1.70 (t, *J* = 6.8 Hz, 2 H), 2.61 (t, *J* = 6.8 Hz, 2 H), 3.74 (s, 6 H), 6.30 (s, 1 H), 6.48 (s, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 22.1 (CH₂), 26.7 (2 X CH₃), 32.9 (CH₂), 55.8 (CH₃), 56.4 (CH₃), 73.9 (Cq), 101.2 (CH), 111.2 (Cq), 112.2 (CH), 142.6 (Cq), 147.6 (Cq), 148.3 (Cq).

6-Methoxy-2,2-dimethyl-3,4-dihydro-2H-1-benzopyran (2d)



yield (0.130 g, 58 %); colorless oil.⁵²

IR (neat): $\tilde{v} = 2972$, 1492, 1247 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.23 (s, 6 H), 1.69 (t, *J* = 6.8 Hz, 2 H), 2.66 (t, *J* = 6.8 Hz, 2 H), 3.65 (s, 3 H), 6.52 (d, *J* = 2.4 Hz, 1 H), 6.60-6.64 (m, 2 H).

¹³C NMR (CDCl₃, 100 MHz): δ 22.8 (CH₂), 26.8 (2 X CH₃), 32.8 (CH₂), 55.7 (CH₃), 73.8 (Cq), 113.4 (CH), 113.9 (CH), 117.8 (CH), 121.5 (Cq), 148.0 (Cq), 152.9 (Cq).

7-Methoxy-2,2-dimethyl-3,4-dihydro-2*H*-1-benzopyran (2e)



yield (0.101 g, 45 %); colorless oil.¹²⁴

IR (neat): $\tilde{v} = 2974$, 1620, 1504, 1151 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.25 (s, 6 H), 1.71 (t, *J* = 6.8 Hz, 2 H), 2.63 (t, *J* = 6.8 Hz, 2 H), 3.67 (s, 3 H), 6.28 (d, *J* = 2.4 Hz, 1 H), 6.36 (dd, *J* = 8.4, 2.4 Hz, 1 H), 6.87 (d, *J* = 8.4 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 21.8 (CH₂), 26.9 (2 X CH₃), 32.9 (CH₂), 55.2 (CH₃), 74.3 (Cq), 101.7 (CH), 106.9 (CH), 112.9 (Cq), 129.9 (CH), 154.7 (Cq), 159.1 (Cq).

6,6-Dimethyl-7,8-dihydro-6H-[1,3]dioxolo[4,5-g]chromene (2f)



yield (0.139 g, 58 %); colorless oil.⁵²

IR (neat): $\tilde{v} = 2974$, 1504, 1479, 1151 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.22 (s, 6 H), 1.67 (t, *J* = 6.8 Hz, 2 H), 2.59 (t, *J* = 6.8 Hz, 2 H), 5.77 (s, 2 H), 6.26 (s, 1 H), 6.42 (s, 1 H).

¹³C NMR (CDCl3, 100 MHz): δ 22.6 (CH₂), 26.6 (2 X CH₃), 32.8 (CH₂), 73.9 (Cq), 98.9 (CH), 100.7 (CH₂), 108.1 (CH), 112.2 (Cq), 140.9 (Cq), 146.4 (Cq), 148.4 (Cq).

6-Methoxy-2,2-dimethyl-3,4-dihydro-2*H*-benzo[*h*]chromene (18)



yield (0.169 g, 60 %); colorless solid; m.p. 74-75 °C; lit.⁶⁴ 77-78 °C.

IR (KBr): $\tilde{v} = 2974$, 1633, 1597, 1273 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.30 (s, 6 H), 1.79 (t, *J* = 6.8 Hz, 2 H), 2.73 (t, *J* = 6.8 Hz, 2 H), 3.82 (s, 3 H), 6.38 (s, 1 H), 7.33 (dt, *J* = 7.6, 1.6 Hz, 1 H); 7.37 (dt, *J* = 7.2, 1.6 Hz, 1 H); 8.05 (dd, *J* = 8.4, 1.6 Hz, 1 H); 8.08 (dd, *J* = 8.8, 1.6 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 23.4 (CH₂), 26.9 (2 X CH₃), 33.1 (CH₂), 55.8 (CH₃), 73.9 (Cq), 105.4 (CH), 113.4 (Cq), 121.4 (CH), 121.7 (CH), 125.0 (CH), 125.3 (Cq), 125.7 (CH), 126.5 (Cq), 142.5 (Cq), 148.5 (Cq).

7-Hydroxy-2,2-dimethyl-3,4-dihydro-2H-1-benzopyran (2g)



yield (0.096 g, 46 %); colorless solid; m.p. 68-70 °C; lit.^{45c} 72-73 °C.

IR (KBr): $\tilde{v} = 3383, 2974, 1593, 1149 \text{ cm}^{-1}$.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.24 (s, 6 H), 1.69 (t, *J* = 6.8 Hz, 2 H), 2.60 (t, *J* = 6.8 Hz, 2 H), 6.22 (s, 1 H), 6.28 (dd, *J* = 8.4, 2.0 Hz, 1 H), 6.81 (d, *J* = 8.4 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 20.7 (CH₂), 25.8 (2 X CH₃), 31.9 (CH₂), 73.5 (Cq), 102.7 (CH), 106.5 (CH), 112.2 (Cq), 129.1 (CH), 153.5 (Cq), 153.8 (Cq).

6-Hydroxy-2,2-dimethyl-3,4-dihydro-2H-1-benzopyran (2h)



yield (0.069 g, 33 %); colorless solid; m.p. 77-78 °C; lit.⁵⁰ 77-78 °C.

IR (KBr): $\tilde{v} = 3379, 2974, 1502, 1195 \text{ cm}^{-1}$.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.23 (s, 6 H), 1.69 (t, *J* = 6.8 Hz, 2 H), 2.63 (t, *J* = 6.8 Hz, 2 H), 6.48 (d, *J* = 2.8 Hz, 1 H), 6.52 (dd, *J* = 8.8, 2.8 Hz, 1 H), 6.57 (d, *J* = 8.4 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 21.6 (CH₂), 25.7 (2 X CH₃), 31.7 (CH₂), 72.9 (Cq), 113.4 (CH), 114.4 (CH), 116.8 (CH), 120.8 (Cq), 146.8 (Cq), 147.5 (Cq).

2,2-Dimethyl-3,4-dihydro-2H-chromene-5,7-diol (2i)



yield (0.127 g, 56 %); colorless solid; m.p. 158-160 °C; lit.^{45c,130} 163-164 °C.

IR (KBr): $\tilde{v} = 3375$, 1600, 1144, 1053 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.13 (s, 6 H), 1.56 (t, *J* = 6.4 Hz, 2 H), 2.39 (t, *J* = 6.4 Hz, 2 H), 5.21 (s, 2H), 5.81 (s, 2 H).

¹³C NMR (CDCl₃, 100 MHz): δ 16.4 (CH₂), 26.6 (2 X CH₃), 32.2 (CH₂), 74.4 (Cq), 94.7 (CH), 96.6 (CH), 100.9 (Cq), 154.6 (Cq), 155.0 (Cq), 155.5 (Cq).

6-Chloro-2,2-dimethyl-3,4-dihydro-2H-1-benzopyran (2j)



yield (0.046 g, 20 %); colorless oil.⁶⁴

IR (neat): $\tilde{v} = 2976$, 1479, 1261, 1122 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.31 (s, 6 H), 1.77 (t, *J* = 6.8 Hz, 2 H), 2.73 (t, *J* = 6.8 Hz, 2 H), 6.70 (d, *J* = 9.2 Hz, 1 H), 7.02 (d, *J* = 9.6 Hz, 1 H), 7.03 (s, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 22.4 (CH₂), 26.8 (2 X CH₃), 32.4 (CH₂), 74.5 (Cq), 118.6 (CH), 122.5 (Cq), 124.2 (Cq), 127.2 (CH), 128.9 (CH), 152.6 (Cq).

<u>3-Methyl-3-(4,8,12-trimethyltridecyl)-2,3-dihydro-1*H*-benzo[*f*]chromene (2k)</u>



yield (0.109 g, 76 %); pale yellow oil.

Rf: 0.50 (5 % ethyl acetate/petroleum ether).

IR (neat): $\tilde{v} = 2951$, 1625, 1598, 1234 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 0.80-0.87 (m, 12 H), 1.10-1.15 (m, 7 H), 1.24-1.67 (m, 14 H), 1.32 (s, 3 H), 1.89-1.99 (m, 2 H), 2.99 (t, *J* = 6.8 Hz, 2 H) 7.03 (d, *J* = 8.8 Hz, 1 H), 7.31 (t, *J* = 7.6 Hz, 1 H), 7.47 (t, *J* = 7.6 Hz, 1 H), 7.60 (d, *J* = 8.8 Hz, 1 H), 7.74 (d, *J* = 8.0 Hz, 1 H), 7.81 (d, *J* = 8.4 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 19.1 (CH₂), 19.7-19.8 (3 peak tops, CH₃), 21.2 (CH₂), 22.7-22.8 (2 peak tops, CH₃), 23.9 (CH₃), 24.5 (CH₂), 24.8-24.9 (2 peak tops, CH₂), 28.0 (CH), 30.8-30.9 (2 peak tops, CH₂), 32.7-32.8 (4 peak tops, CH), 37.3-37.6 (6 peak tops, CH₂), 39.4-39.6 (3 peak tops, CH₂), 76.1 (Cq), 112.6 (Cq), 119.9 (CH), 121.9 (CH), 122.9 (CH), 126.2 (CH), 127.7 (CH), 128.4 (CH), 128.7 (Cq), 133.1 (Cq), 151.4 (Cq).

Anal. Calcd. For C₃₀H₄₆O: C, 85.3; H, 10.9 %; Found: C, 85.6; H, 11.1 %; GCMS: *m/z* calcd for C₃₀H₄₆O [M]⁺: 422.68; found: 422.22.

3-Phenyl-2,3-dihydro-1*H*-benzo[*f*]chromene (2l)



yield (0.117 g, 60 %); colorless solid; m.p. 84-86 °C; lit.¹³¹ 86 °C.

IR (KBr): $\tilde{v} = 3062, 2924, 1597, 1236 \text{ cm}^{-1}$.

¹**H** NMR (CDCl₃, 400 MHz): δ 2.11-2.19 (m, 1 H), 2.28-2.33 (m, 1 H), 3.06-3.10 (m, 2 H), 5.04 (dd, J = 10.0, 2.0 Hz, 1 H), 7.07 (d, J = 8.8 Hz, 1 H), 7.23-7.43 (m, 7 H), 7.56 (d, J = 8.8 Hz, 1 H), 7.69 (d, J = 8.0 Hz, 1 H), 7.73 (d, J = 8.8 Hz, 1 H).

¹³C NMR (CDCl3, 100 MHz): δ 21.8 (CH₂), 29.7 (CH₂), 77.5 (CH), 113.6 (Cq), 119.2 (CH), 121.9 (CH), 123.3 (CH), 126.1 (2 X CH), 126.4 (CH), 127.8 (CH), 127.9 (CH), 128.5 (CH), 128.6 (2 X CH), 129.0 (Cq), 133.0 (Cq), 141.6 (Cq), 152.7 (Cq).

As a further application to our work, synthesis of naturally occurring precocene II 44 and lapachenole 45 was performed (Scheme 40). Compound 44 is an insect growth regulator, genotoxic and produces hepatic centrolobular necrosis in rats.¹³² Compound 45 has been used

as a fluorescent photoaffinity label¹³³ and also shows cancer chemopreventive activity.¹³⁴ Chroman **2c** was refluxed in benzene with DDQ for 6 h to give **44** in 60 % yield.¹³⁵ Similarly chroman **18** on refluxing for 1.3 h in benzene delivered **45** in 52 % yield.



Scheme 40: Synthesis of precocene II 44 and lapachenole 45.

Spectral data of chromenes

6,7-Dimethoxy-2,2-dimethyl-2H-chromene/Precocene II (44)



yield (0.059 g, 60 %); colorless solid; m.p. 46-48 °C; lit.¹³⁶ 45-46 °C.

IR (KBr): $\tilde{v} = 2974$, 1500, 1458, 1278, 1195 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.34 (s, 6 H), 3.75 (s, 3 H), 3.76 (s, 3 H), 5.41 (d, *J* = 10.0 Hz, 1 H), 6.17 (d, *J* = 10.0 Hz, 1 H), 6.34 (s, 1 H), 6.46 (s, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 26.6 (2 X CH₃), 54.9 (OCH₃), 55.5 (OCH₃), 74.9 (Cq), 99.9 (CH), 108.7 (CH), 112.0 (Cq), 120.9 (CH), 127.2 (CH), 142.0 (Cq), 146.2 (Cq), 148.6 (Cq).

6-Methoxy-2,2-dimethyl-2H-benzo[h]chromene/Lapachenole (45)



yield (0.051 g, 52 %); colorless solid; m.p. 63-64 °C; lit.¹³⁷ 63-64 °C.

IR (KBr): $\tilde{v} = 2974, 2933, 1641, 1597, 1276 \text{ cm}^{-1}$.

¹**H NMR (CDCl₃, 400 MHz):** δ 1.42 (s, 6 H), 3.89 (s, 3 H), 5.58 (d, *J* = 9.6 Hz, 1 H), 6.33 (d, *J* = 9.6 Hz, 1 H), 6.44 (s, 1 H), 7.34-7.42 (m, 2 H), 8.08 (d, *J* = 8.4 Hz, 2 H).

¹³C NMR (CDCl₃, 100 MHz): δ 27.6 (2 X CH₃), 55.8 (OCH₃), 76.2 (Cq), 102.5 (CH), 114.8 (Cq), 121.7 (CH), 121.8 (CH), 123.0 (CH), 125.5 (CH), 125.9 (CH), 126.0 (Cq), 128.3 (Cq), 129.9 (CH), 141.9 (Cq), 149.3 (Cq).



Scheme 41: Proposed mechanism for the formation of chromans.

The proposed mechanistic pathway for the formation of chroman is shown in scheme 41. Loss of alcoholic OH group takes place by coordination of iodine making it a better leaving group facilitated by allylic double bond. Intermolecular electrophillic attack by the phenolic arene ring on less hindered carbon atom of thus formed carbocation followed by intramolecular oxygen attack on more substituted carbon atom of double bond and subsequent aromatization delivered chroman. On the other hand if the double bond coordinates with iodine to form iodinium ion prior to the intramolecular oxygen attack, oxygen attack can take place either by path A or path B to give 2' or 2'' respectively. But the obtained product 2a' indicates the attack at more substituted carbon atom which is favoured when excess of iodine is used.

2.5: Conclusion

We have demonstrated the utility of molecular iodine as a mild Lewis acid for the synthesis of oxygen heterocycle chromans *via* [3+3] cyclocoupling of phenols and allylic alcohols. 30 Mol % of iodine was effective in chloroform or methanol solvent under refluxing condition. The usefulness of the method is demonstrated by synthesis of naturally occurring chroman dihydrolapachenole **18**.

Further these chromans can be used for the synthesis of natural chromenes by conventional dehydrogenation which is demonstrated by synthesizing precocene II **44** and lapachenole **45**.

2.6: Experimental



2.6.1: A general procedure for the synthesis of chromans 2a-l, 18: In a 50 mL round bottom flask, unsaturated alcohol 43 (1.16 mmol) was mixed with chloroform/methanol (10 mL). To it, substituted phenol 42 (4.65 mmol) and iodine (0.35 mmol) were added at room temperature. This reaction mixture was then subjected to reflux with stirring for the mentioned 2-24 h. It was then cooled to room temperature. Chloroform was directly washed with saturated solution of sodium thiosulphate whereas methanol was removed under reduced pressure followed by addition of chloroform solvent. Chloroform was then washed with dilute sodium hydroxide solution and then with water, dried over sodium sulphate and concentrated to furnish the crude product. This was then purified using 60-120 mesh silica gel column chromatography with petroleum ether-ethyl acetate as an eluent to give chromans 2a-l, 18 in 20-92 % yield.



2.6.2: A procedure for the synthesis of chroman 2a: In a 50 mL round bottom flask, prenyl alcohol 43a (0.1 g, 1.16 mmol) was mixed with chloroform (10 mL). To it, β -naphthol 42a (0.67 g, 4.65 mmol) and iodine (0.044 g, 0.35 mmol) were added at room temperature. This reaction mixture was then subjected to reflux with stirring for 4 h. It was then cooled to room temperature. Chloroform was directly washed with saturated sodium thiosulphate solution followed by washings with dilute sodium hydroxide solution and water, dried over sodium sulphate and concentrated to furnish the crude product. This was then purified using 60-120 mesh silica gel column chromatography to give chroman 2a (0.227 g, 92 %) with petroleum ether-ethyl acetate (9.8:0.2) as an eluent.



2.6.3: A procedure for the synthesis of chromenes 44, 45: To 3,4-dimethoxychroman 2c (0.1 g, 0.45 mmol) /dihydrolapachenole 18 (0.1 g, 0.41 mmol) in a 50 mL round bottom flask, was added DDQ (1 equiv) and was subjected to reflux in benzene (10 mL). After completion of reaction, benzene was removed by distillation and crude reaction mixture thus obtained was purified by column chromatography to furnish chromenes 44 (0.059 g, 60 %) and 45

(0.051 g, 52 %) with petroleum ether-ethyl acetate (9:1) and (9.5:0.5) respectively as an eluent.



2.6.4: A procedure for the synthesis of 2-iodo-3,3-dimethyl-2,3-dihydro-1*H*benzo[*f*]chromene 2a': A mixture of prenyl alcohol 43a (0.1 g, 1.16 mmol), β -naphthol 42a (1.16 mmol) and iodine (1.16 mmol) in chloroform (10 mL) was stirred for 1.3 h at room temperature. Chloroform was then directly washed with saturated solution of sodium thiosulphate followed by dilute sodium hydroxide solution. Finally chloroform layer was washed with water, dried over sodium sulphate and concentrated to furnish the crude mixture of 2a and 2a'. This was then purified using 60-120 mesh silica gel column chromatography to give pure 2a (0.143 g, 58 %) and 2a' (0.122 g, 31 %) with petroleum ether-ethyl acetate (9.8:0.2) and (9.95:0.05) respectively as an eluent.

2.7: References

- a) Hepworth, J. D.; Gabbutt, C. D.; Heron, M. B. Comprehensive Heterocyclic Chemistry II; Pergamon: New York, NY, **1996**; p 301. b) Geen, G. R.; Evans, J. M.; Vong, A. K. Comprehensive Heterocyclic Chemistry II; Pergamon: New York, NY, **1996**; p 469.
- a) Koufaki, M. *Expert Opin. Ther. Pat.* 2016, 26, 35. b) Schneider, C. *Mol. Nutr. Food Res.* 2005, 49, 7. c) Brigelius-Flohe, R.; Traber, M. G. *FASEB J.* 1999, 13, 1145.
- a) Ren,W.; Qiao, Z.;Wang, H.; Zhu, L.; Zhang, L. Med. Res. Rev. 2003, 23, 519. b) Middleton, E., Jr.; Kandaswami, C.; Theoharides, T. C. Pharmacol. Rev. 2000, 52, 673.
- a) Güder, A.; Korkmaz, H.; Gökce, H.; Alpaslan, Y. B.; Alpaslan, G. Spectrochim. Acta A Mol. Biomol. Spectrosc. 2014, 133, 378. b) Malik, M. N.; Fenko, M. D.; Shiekh, A. M.; Wisniewski, H. M. J. Agric. Food Chem. 1997, 45, 817. c) Terashima, K.; Shimamura, T.; Tanabayashi, M.; Aqil, M.; Akinniyi, J. A.; Niwa, M. Heterocycles 1997, 45, 1559.
- Loganathan, R.; Selvaduray, K. R.; Nesaretnam, K.; Radhakrishnan, A. K. Cell Prolif. 2013, 46, 203.
- a) Sen, C. K., Khanna, S., Roy, S., and Packer, L. J. Biol. Chem. 2000, 275, 13049. b) Sen, C. K., Khanna, S., and Roy, S. Ann. N. Y. Acad. Sci. 2004, 1031, 127.
- 7. Kannappan, R.; Yadav, V. R.; Aggarwal, B. B. J. Biol. Chem. 2010, 285, 33520.
- a) Delle Monache, F.; Marta, M.; Mac-Quhae, M. M.; Nicoletti, M. *Gazz. Chim. Ital.* 1984, *114*, 135. b) Setzer, W. N.; Green, T. J.; Lawton, R. O.; Moriarity, D. M.; Bates, R. B.; Caldera, S.; Haber, W. A. *Planta Med.* 1995, *61*, 275. c) Teixeira, J. S. R.; Moreira, L. de M.; Guedes, M. L. da S.; Cruz, F. G. J. Braz. Chem. Soc. 2006, *17*, 812.

- Alsabil, K.; Suor-Cherer, S.; Koeberle, A.; Viault, G.; Lavaud, A.; Temml, V.; Waltenberger, B.; Schuster, D.; Litaudon, M.; Lorkowski, S.; Vaumas, R.; Helesbeux, J-J.; Guilet, D.; Stuppner, H.; Werz, O.; Seraphin, D.; Richomme, P. *Planta Med.* 2016, 82, 1110.
- Gonzalez, M. C.; Serrano, A.; Zafra-Polo, M. C.; Cortes, D.; Rao, K. S. J. Nat. Prod. 1995, 58, 1278.
- a) Anjaneyulu, A. S. R.; Rahgava Reddy, A. V.; Reddy, D. S. K.; Cameron, T. S.; Roe, S. P. *Tetrahedron* 1986, 42, 2417. b) Palanisamy, P.; Gandhidasan, R.; Raman, P. V.; Krishnasamy, N. R.; Nanduri, S. *Phytochemistry* 1994, 36, 817.
- 12. Ali, S.; Goundar, R.; Sotheeswaran, S.; Beaulieu, C.; Spino, C. Phytochemistry 2000, 53, 281.
- Seo, E. K.; Wani, M. C.; Wall, M. E.; Navarro, H.; Mukherjee, R.; Farnsworth, N. R.; Kinghorn, A. D. *Phytochemistry* 2000, 55, 35.
- Kumazava, S.; Suzuki, S.; Ahn, M-R.; Kamihira, M.; Udagawa, Y.; Bang, K-S.; Nakayama, T. Food Sci. Technol. Res. 2006, 12, 67.
- a) Y. Gaoni, R. Mechoulam, J. Am. Chem. Soc. 1964, 86, 1646. b) Ahmed, S. A.; Ross, S. A.;
 Slade, D.; Radwan, M. M.; Zulfiqar, F.; ElSohly, M. A. J. Nat. Prod. 2008, 71, 536.
- a) Chen, H.; Li, J.; Wu, Q.; Niu, X-T.; Tang, M-T.; Guan, X-L.; Li, J.; Yang, R-Y.; Deng, S-P.; Su, X-J. *Fitoterapia* **2012**, *83*, 643. b) Maltarollo, V. G.; Sannomiya, M.; Honório, K. M. *J. Comput. Theor. Nanosci.* **2013**, *10*, 1385. c) Moodley, R.; Koorbanally, N. A.; Islam, M. D. S.; Jonnalagadda, S. B. *J. Environ. Sci. Health B* **2014**, *49*, 938. d) Pitchai, D.; Manikkam, R. *Med. Chem. Res.* **2012**, *21*, 2238. e) Hye, M. A.; Taher, M. A.; Ali, M. Y.; Ali, M. U.; Zaman, S. *J. Sci. Res.* **2009**, *1*, 300. f) Chaves, M. C. C. A. e M. H.; Rinaldo, D.; Júnior, W. V. e G. M. V. *Quim. Nova* **2009**, *32*, 1509.
- 17. Itokawa, H.; Qiao, Y.; Takeya, K. Phytochemistry 1989, 28, 3465.
- 18. Burnett, A. R.; Thomson, R. H. J. Chem. Soc. C 1968, 850.
- a) El-Hady, S.; Bukuru, J.; Kesteleyn, B.; Van Puyvelde, L.; Nguyen Van, T.; De Kimpe, N. J. Nat. Prod. 2002, 65, 1377. b) Itokawa, H.; Ibraheim, Z. Z.; Qiao, Y-F.; Takeya, K. Chem. Pharm. Bull. 1993, 41, 1869.
- Son, J. K.; Jung, S. J.; Jung, J. H.; Fang, Z.; Lee, C. S.; Seo, C. S.; Moon, D. C.; Min, B. S.; Kim, M. R.; Woo, M. H. *Chem. Pharm. Bull.* **2008**, *56*, 213.
- 21. a) Seeram, N. P.; Jacobs, H.; McLean, S.; Reynolds, W. F. *Phytochemistry* 1998, 49, 1389. b)
 Burke, S. J.; Jacobs, H.; Mclean, S.; Reynolds, W. F. *Magn. Reson. Chem.* 2003, 41, 145. c)
 Tanaka, T.; Asai, F.; Iinuma, M. *Phytochemistry* 1998, 49, 229. d) Mota, J. da S.; Leite, A.
 C.; Junior, J. M. B.; López, S. N.; Ambrósio, D. L.; Passerini, G. D.; Kato, M. J.; Bolzani, V.
 da S.; Cicarelli, R. M. B.; Furlan, M. *Planta Med.* 2009, 75, 620.
- Kashiwada, Y.; Yamazaki, K.; Ikeshiro, Y.; Yamagishi, T.; Fujioka, T.; Mihashi, K.; Mizuki, K.; Cosentino, L. M.; Fowke, K.; Morris-Natschke, S. L.; Lee, K-H. *Tetrahedron* 2001, *57*, 1559.

- 23. Ishibashi, K. J. J. Antibiot. Ser. A. 1962, 15, 161.
- Matsuda, H.; Murakami, T.; Nishida, N.; Kageura, T.; Yoshikawa, M. Chem. Pharm. Bull.
 2000, 48, 1429.
- 25. Willette, R. E.; Soine, T. O. J. Pharm. Sci. 1962, 51, 149.
- a) Lee, T. T-Y.; Kashiwada, Y.; Huang, L.; Snider, J.; Cosentino, M.; Lee, K-H. *Bioorg. Med. Chem.* **1994**, *2*, 1051. b) Delgado, G.; Garduño, J. *Phytochemistry* **1987**, *26*, 1139.
- 27. a) Akihisa, T.; Tokuda, H.; Ukiya, M.; Iizuka, M.; Schneider, S.; Ogasawara, K.; Mukainaka, T.; Iwatsuki, K.; Suzuki, T.; Nishino, H. *Cancer Lett.* 2003, 201, 133. b) Swager, T. M.; Cardellina II, J. H. *Phytochemistry* 1985, 24, 805.
- 28. Jain, R.; Yadav, N.; Jain, S. C. J. Chem. Pharm. Res. 2015, 7, 1032.
- a) El-Alfy, A. T.; Ivey, K.; Robinson, K.; Ahmed, S.; Radwan, M.; Slade, D.; Khan, I.; ElSohly, M.; Ross, S. *Pharmacol. Biochem. Behav.* 2010, 95, 434. b) Ueki, S.; Fujiwara, M.; Ogawa, N. *Physiol. Behav.* 1972, 9, 585.
- a) Idhayadhulla , A.; Xia , L.; Lee, Y. R.; Kim, S. H.; Wee, Y-J.; Lee, C-S. *Bioorg. Chem.* **2014**, *52*, 77. b) Ho, L-K. *J. Nat. Prod.* **1996**, *59*, 330.
- 31. Folkes, K.; Wolf, D. E. U. S. Patent 3,026,330, Mar. 20, 1962.
- 32. Ray, S.; Grover, P. K.; Kamboj, V. P.; Setty, B. S.; Kar, A. B.; Anand, N. J. Med. Chem. **1976**, *19*, 276.
- 33. Huang, L.; Kashiwada, Y.; Cosentino, M.; Fan, S.; Lee, K-H. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 593.
- 34. a) Ashwood, V. A.; Buckingham, R. E.; Cassidy, F.; Evans, J. M.; Faruk, E. A.; Hamilton, T. C.; Nash, D. J.; Stemp, G.; Willcocks, K. J. Med. Chem. 1986, 29, 2194. b) Asano, M.; Masuzawa-Ito, K.; Matsuda, T. Eur. J. Pharmacol. 1994, 263, 121. c) Gurpinar, T.; Gok, S. Swiss Med. Wkly. 2012, 142, w13558. d) Fabiani, M. E.; Story, D. F. Pharmacological Res. 1995, 32, 155. e) Fabiani, M. E.; Vlahos, R.; Story, D. F. Pharmacological Res. 1996, 33, 261. f) Vlahos, R.; Fabiani, M. E.; Story, D. F. Naunyn Schmiedeberg's Arch. Pharmacol 2003, 368, 256.
- 35. a) Terao, K.; Niki, E. J. Free Rad. Biol. Med. 1986, 2, 193. b) Albertini, R.; Abuja, P. M. Free Rad. Res. 1999, 30, 181. c) Huang, S-W.; Hopia, A.; Schwarz, K.; Frankel, E. N.; German, J. B. J. Agric. Food Chem. 1996, 44, 444.
- Grisar, J. M.; Petty, M. A.; Bolkenius, F. N.; Dow, J.; Wagner, J.; Wagner, E. R.; Haegele, K. D.; Jong, W. D. J. Med. Chem. 1991, 34, 257.
- a) Sankaran, M. S.; Prasad, M. R. N. Contraception 1974, 9, 279. b) Lal, J. Contraception 2010, 81, 275.
- 38. a) De Crée, J.; Geukens, H.; Leempoels, J.; Verhaegen, H. *Drug Dev. Res.* 1986, *8*, 109. b)
 Van de Water, A.; Janssens, W.; Van Neuten, J.; Xhonneux, R.; De Cree, J.; Verhaegen, H.;
 Reneman, R. S.; Janssen, P. A. J. *J. Cardiovasc. Pharmacol.* 1988, *11*, 552.

- 39. a) Lee, K-S.; Park, J-H.; Lee, S.; Lim, H-J.; Jang, Y.; Park, H-Y. *Biochem. Biophys. Res. Commun.* 2006, *346*, 83. b) Deng, R.; Nie, A.; Jian, F.; Liu, Y.; Tang, H.; Zhang, J.; Zhang, Y.; Shao, L.; Li, F.; Zhou, L.; Wang, X.; Ning, G. *Biochim. Biophys. Acta* 2014, *1840*, 577. c) Wang, X.; Zhou, L.; Shao, L.; Qian, L.; Fu, X.; Li, G.; Luo, T.; Gu, Y.; Li, F.; Li, J.; Zheng, S.; Luo, M. *Life Sci.* 2007, *81*, 160.
- 40. a) Tang, Y.; Oppenheimer, J.; Song, Z.; You, L.; Zhang, X.; Hsung, R. P. *Tetrahedron* 2006, 62, 10785. b) Shi, Y.-L.; Shi, M. Org. Biomol. Chem. 2007, 5, 1499. c) Ferreira, S. B.; de C. da Silva, F.; Pinto, A. C.; Gonzaga, D. T. G.; Ferreira, V. F. J. Heterocycl. Chem. 2009, 46, 1080. d) Masesane, I. B.; Desta, Z. Y. Beilstein J. Org. Chem. 2012, 8, 2166.
- 41. a) Alamsetti, K. S.; Spanka, M.; Schneider, C. Angew. Chem. Int. Ed. 2016, 55, 2392. b) Wang, P-S.; Liu, P.; Zhai, Y-J.; Lin, H-C.; Han, Z-Y.; Gong, L-Z. J. Am. Chem. Soc. 2015, 137, 12732. c) Saha, P.; Biswas, A.; Molleti, N.; Singh, V. K. J. Org. Chem. 2015, 80, 11115. d) Zhou, R.; Wu, Q.; Guo, M.; Huang, W.; He, X.; Yang, L.; Peng, F.; He, G.; Han, B. Chem. Commun. 2015, 51, 13113. e) Shen, H. C. Tetrahedron 2009, 65, 3931. f) Netscher, T. Vitam. Horm. 2007, 76, 155.
- 42. Bolzoni, L.; Casiraghi, G.; Casnati, G.; Sartori, G. Angew. Chem. Int. Ed. Engl. 1978, 17, 684.
- 43. Raju, G. N.; Suresh, P. V.; Nadendla, R. R.; Anusha, K. Der Pharma Chemica 2015, 7, 346.
- 44. Smith, L. I.; Ungnade, H. E.; Hoehn, H. H.; Wawzonek, S. J. Org. Chem. 1939, 4, 311.
- 45. a) Bader, A. R.; Bean, W. C. J. Am. Chem. Soc. 1958, 80, 3073. b) Ahluwalia, V. K.; Arora, K. K. Tetrahedron 1981, 37, 1437. c) Ahluwalia, V. K.; Arora, K. K.; Jolly, R. S. J. Chem. Soc. Perkin Trans. 1 1982, 335.
- 46. Dewhirst, K. C.; Rust, F. F. J. Org. Chem. 1963, 28, 798.
- 47. Matsui, M.; Yamamoto, H. Bull. Chem. Soc. Jpn. 1995, 68, 2657.
- 48. Matsui, M.; Yamamoto, H. Bull. Chem. Soc. Jpn. 1995, 68, 2663.
- 49. Kalena, G. P.; Jain, A.; Banerji, A. Molecules 1997, 2, 100.
- 50. Bigi, F.; Carloni, S.; Maggi, R.; Muchetti, C.; Rastelli, M.; Sartori, G. Synthesis 1998, 301.
- 51. Bienaymé, H.; Ancel, J.-E.; Meilland, P.; Simonato, J.-P. Tetrahedron Lett. 2000, 41, 3339.
- 52. Youn, S. W.; Eom, J. I. J. Org. Chem. 2006, 71, 6705.
- 53. Dang, T. T.; Boeck, F.; Hintermann, L. J. Org. Chem. 2011, 76, 9353.
- 54. Youn, S. W. Synlett 2007, 3050.
- 55. Adrio, L. A.; Hii, K. K. Chem. Commun. 2008, 2325.
- 56. Judd, K. E.; Caggiano, L. Org. Biomol. Chem. 2011, 9, 5201.
- 57. Villani-Gale, A.-J.; Eichman, C. C. Eur. J. Org. Chem. 2016, 2016, 2925.
- a) Mamalis, P.; Mchale, D.; Green, J.; Marcinkiewicz, S. J. Chem. Soc. 1958, 1850. b) Harel,
 D.; Khalid, S. A.; Kaiser, M.; Brun, R.; Wünsch, B.; Schmidt, T. J. J. Ethnopharmacol. 2011,
 137, 620.
- 59. Wehrli, P. A.; Fryer, R. I.; Metlesics, W. J. Org. Chem. 1971, 36, 2910.

- 60. Ismail, F. M. D.; Hilton, M. J.; Štefinović, M. Tetrahedron Lett. 1992, 33, 3795.
- 61. Lee, J. H.; Bang, H. B.; Han, S. Y.; Jun, J.-G. Bull. Korean Chem. Soc. 2006, 27, 2104.
- 62. Malkov, A. V.; Spoor, P.; Vinader, V.; Kočovský, P. J. Org. Chem. 1999, 64, 5308.
- Malkov, A. V.; Davis, S. L.; Baxendale, I. R.; Mitchell, W. L.; Kočovský, P. J. Org. Chem. 1999, 64, 2751.
- 64. Yamamoto, Y.; Itonaga, K. Org. Lett. 2009, 11, 717.
- Ishino, Y.; Mihara, M.; Hayakawa, N.; Miyata, T.; Kaneko, Y.; Miyata, T. Synth. Commun. 2001, 31, 439.
- Vece, V.; Ricci, J.; Poulain-Martini, S.; Nava, P.; Carissan, Y.; Humbel, S.; Duñach, E. *Eur. J. Org. Chem.* 2010, *32*, 6239.
- 67. Varghese, S.; Anand, C.; Dhawale, D.; Mano, A.; Balasubramanian, V. V.; Raj, G. A. G.; Nagarajan, S.; Wahab, M. A.; Vinu, A. *Tetrahedron Lett.* **2012**, *53*, 5656.
- Varghese, S.; Anand, C.; Dhawale, D.; Mane, G. P.; Wahab, M. A.; Mano, A.; Raj, G. A. G.; Nagarajan, S.; Vinu, A. *Chem. Cat. Chem.* 2013, *5*, 899.
- 69. Murthy, Y. L. N.; Suhasini, K.; Jha, A. J. Serb. Chem. Soc. 2012, 77, 859.
- Madabhushi, S.; Jillella, R.; Godala, K. R.; Mallu, K. K. R.; Beeram, C. R.; Chinthala, N. *Tetrahedron Lett.* 2012, 53, 5275.
- 71. Coutant, E.; Young, P. C.; Barker, G.; Lee, A.-L. Beilstein J. Org. Chem. 2013, 9, 1797.
- 72. Miller, J. A.; Wood, H. C. S. J. Chem. Soc. C 1968, 1837.
- 73. Cardillo, G.; Cricchio, R.; Merlini, L. Tetrahedron 1968, 24, 4825.
- 74. Dintzner, M. R.; McClelland, K. M.; Morse, K. M.; Akroush, M. H. Synlett 2004, 11, 2028.
- 75. Camps, F.; Coll, J.; Messeguer, A.; Pericás, M. A.; Ricart, S. Synthesis 1979, 126.
- 76. Jetter, M. M.; Heindel, N. D.; Laskin, J. D. J. Heterocycl. Chem. 1990, 27, 995.
- 77. Dauben, W. G.; Cogen, J. M.; Behar, V. Tetrahedron Lett. 1990, 31, 3241.
- 78. Pogrebnoi, S. I.; Kal'yan, Y. B.; Krimer, M. Z.; Smit, V. A. Bull. Acad. Sci. USSR, Div. Chem. Sci. 1991, 40, 733.
- 79. Bernard, A. M.; Cocco, M. T.; Onnis, V.; Piras, P. P. Synthesis 1998, 256.
- 80. Ollevier, T.; Mwene-Mbeja, T. M. Synthesis 2006, 3963.
- 81. Barluenga, J.; Trincado, M.; Rubio, E.; González, J. M. J. Am. Chem. Soc. 2004, 126, 3416.
- 82. Trost, B. M.; Toste, F. D. J. Am. Chem. Soc. 1998, 120, 9074.
- 83. Aristoff, P. A.; Harrison, A. W.; Huber, A. M. Tetrahedron Lett. 1984, 25, 3955.
- 84. Yus, M.; Foubelo, F.; Ferrández, J. V.; Bachki, A. Tetrahedron 2002, 58, 4907.
- Jetson, R.; Malik, N.; Luniwal, A.; Chari, V.; Ratnam, M.; Erhardt, P. Eur. J. Med. Chem. 2013, 63, 104.
- Tanaka, T.; Miyaguchi, M.; Mochisuki, R. K.; Tanaka, S.; Okamoto, M.; Kitajima, Y.; Miyazaki, T. *Heterocycles* 1987, 25, 463.
- 87. Nilsson, J. L. G.; Sievertsson, H.; Selander, H. Acta Chem. Scand. 1968, 22, 316.

- 88. Wang, Y.; Wu, J.; Xia, P. Synth. Commun. 2006, 36, 2685.
- Macone, A.; Lendaro, E.; Comandini, A.; Rovardi, I.; Matarese, R. M.; Carraturo, A.; Bonamore, A. *Bioorg. Med. Chem.* 2009, 17, 6003.
- 90. Schlüter, J.; Blazejak, M.; Hintermann, L. Chem. Cat. Chem. 2013, 5, 3309.
- 91. Francesco, I. N.; Cacciuttolo, B.; Pucheault, M.; Antoniotti, S. Green Chem. 2015, 17, 837.
- 92. Hurd, C. D.; Hoffman, W. A. J. Org. Chem. 1940, 5, 212.
- 93. Jiménez, M. C.; Márquez, F.; Miranda, M. A.; Tormos, R. J. Org. Chem. 1994, 59, 197.
- 94. a) Nicolaou, K. C.; Pfefferkorn, J. A.; Cao, G.-Q. Angew. Chem. Int. Ed. 2000, 39, 734. b)
 Nicolaou, K. C.; Pfefferkorn, J. A.; Roecker, A. J.; Cao, G.-Q.; Barluenga, S.; Mitchell, H. J. J. Am. Chem. Soc. 2000, 122, 9939. c)
 Nicolaou, K. C.; Pfefferkorn, J. A.; Schuler, F.; Roecker, A. J.; Cao, G.-Q.; Casida, J. E. Chem. Biol. 2000, 7, 979.
- 95. Boltze, K.-H.; Dell, H.-D. Angew. Chem. Int. Ed. 1966, 5, 415.
- 96. Gopalakrishnan, G.; Kasinath, V.; Singh, N. D. P.; Thirumurugan, R.; Raj, S. S. S.; Shanmugam, G. *Molecules* 2000, 5, 880.
- 97. a) Bravo, P.; Ticozzi, C. J. Heterocycl. Chem. 1978, 15, 1051. b) Grundon, M. F.; Okely, H. M. J. Chem. Soc. Perkin Trans. I 1975, 150.
- 98. Sato, K.; Inoue, S.; Miyamoto, O.; Ikeda, H.; Ota, T. Bull. Chem. Soc. Jpn. 1987, 60, 4184.
- Grigg, R.; Kongkathip, N.; Kongkathip, B.; Luangkamin, S.; Dondas, H. A. *Tetrahedron* 2001, 57, 7965.
- 100. Trend, R. M.; Ramtohul, Y. K.; Stoltz, B. M. J. Am. Chem. Soc. 2005, 127, 17778.
- 101. Ward, A. F.; Xu, Y.; Wolfe, J. P. Chem. Commun. 2012, 48, 609.
- 102. Xu, B.; Xue, J.; Zhu, J.; Li, Y. Chem. Lett. 2008, 37, 202.
- 103. Palucki, M.; Wolfe, J. P.; Buchwald, S. L. J. Am. Chem. Soc. 1996, 118, 10333.
- 104. Torraca, K. E.; Kuwabe, S.-I.; Buchwald, S. L. J. Am. Chem. Soc. 2000, 122, 12907.
- 105. a) Shelby, Q.; Kataoka, N.; Mann, G.; Hartwig, J. J. Am. Chem. Soc. 2000, 122, 10718. b)
 Kataoka, N.; Shelby, Q.; Stambuli, J. P.; Hartwig, J. F. J. Org. Chem. 2002, 67, 5553.
- 106. Niu, J.; Guo, P.; Kang, J.; Li, Z.; Xu, J.; Hu, S. J. Org. Chem. 2009, 74, 5075.
- 107. Suchand, B.; Krishna, J.; Ramulu, B. V.; Dibyendu, D.; Reddy, A. G. K.; Mahendar, L.; Satyanarayana, G. *Tetrahedron Lett.* **2012**, *53*, 3861.
- 108. Furuyama, S.; Togo, H. Synlett 2010, 2325.
- Hamamoto, H.; Hata, K.; Nambu, H.; Shiozaki, Y.; Tohma, H.; Kit, Y. *Tetrahedron Lett.* **2004**, 45, 2293.
- 110. Zheng, Y-W.; Ye, P.; Chen, B.; Meng, Q-Y.; Feng, K.; Wang, W.; Wu, L-Z.; Tung, C-H. *Org. Lett.* **2017**, *19*, 2206.
- 111. Chiba, K.; Hirano, T.; Kitano, Y.; Tada, M. Chem. Commun. 1999, 691.
- 112. Yadav, J. S.; Reddy, V. S.; Parisse, C.; Carvalho, P.; Rao, T. P. *Tetrahedron Lett.* **2002**, *43*, 2999.

- 113. Nakamura, S.; Uchiyama, M.; Ohwada, T. J. Am. Chem. Soc. 2003, 125, 5282.
- Radomkit, S.; Sarnpitak, P.; Tummatorn, J.; Batsomboon, P.; Ruchirawat, S.; Ploypradith, P. *Tetrahedron* 2011, 67, 3904.
- 115. Smith, L. I.; Ungnade, H. E.; Prichard, W. W. J. Org. Chem. 1939, 4, 358.
- 116. Fatope, M. O.; Abraham, D. J. J. Med. Chem. 1987, 30, 1973.
- 117. Teng, M.; Duong, T. T.; Johnson, A. T.; Klein, E. S.; Wang, L.; Khalifa, B.; Chandraratna, R. A. S. J. Med. Chem. 1997, 40, 2445.
- 118. Bernier, D.; Brückner, R. Synthesis 2007, 2249.
- 119. a) Bridge, W.; Crocker, A. J.; Cubin, T.; Robertson, A. J. Chem. Soc. 1937, 1530. b) Casas, J.; Gorchs, G.; Sbchez-Baeza, F.; Teixidor, P.; Messeguer, A. J. Agric. Food Chem. 1992, 40, 585. c) Garazd, Y. L.; Garazd, M. M.; Khilya, V. P. Chem. Nat. Compd. 2004, 40, 427.
- 120. Anioł, M.; Łusiak, P.; Wawrzeńczyk, C. Heterocycles 1994, 38, 991.
- 121. Webb, J. L.; Hall, W. L. J. Org. Chem. 1973, 38, 1621.
- 122. a) Verhé, R.; Schamp, N.; De Buyck, L. Synthesis 1975, 392. b) Verhé, R.; Schamp, N.; De Buyck, L.; De Kimpe, N.; Sadones, M. Bull. Soc. Chim. Belg. 1975, 84, 747.
- 123. Bravo, P.; Ticozzi, C.; Maggi, D. J. Chem. Soc., Chem. Commun. 1976, 789.
- 124. De Renzi, A.; Panunzi, A.; Saporito, A.; Vitagliano, A. J. Chem. Soc. Perkin Trans. II 1983, 993.
- 125. Larock, R. C.; Berrios-Peña, N. G.; Fried, C. A.; Yum, E. K.; Tu, C.; Leong, W. J. Org. Chem. 1993, 58, 4509.
- 126. Widenhoefer, R. A.; Zhong, H. A.; Buchwald, S. L. J. Am. Chem. Soc. 1997, 119, 6787.
- 127. Knight, D. W.; Little, P. B. Tetrahedron Lett. 1998, 39, 5105.
- 128. Yadav, J. S.; Subba Reddy, B. V.; Rao, T. P. Tetrahedron Lett. 2000, 41, 7943.
- 129. Hata, K.; Hamamoto, H.; Shiozaki, Y.; Cämmerer, S. B.; Kita, Y. *Tetrahedron* **2007**, *63*, 4052.
- Cichewicz, R. H.; Kenyon, V. A.; Whitman, S.; Morales, N. M.; Arguello, J. F.; Holman, T. R.; Crews, P. J. Am. Chem. Soc. 2004, 126, 14910.
- Jiménez , M. C.; Leal, P.; Miranda, M. A.; Scaiano, J. C.; Tormos, R. *Tetrahedron* 1998, 54, 4337.
- 132. Duddy, S. K.; Hsia, M. T. Chem. Biol. Interact. 1989, 71, 187.
- 133. Wen, B.; Doneanu, C. E.; Gartner, C. A.; Roberts, A. G.; Atkins, W. M.; Nelson, S. D. Biochemistry 2005, 44, 1833.
- 134. Itoigawa, M.; Ito, C.; Tan, H. T.-W.; Okuda, M.; Tokuda, H.; Nishino, H.; Furukawa, H. *Cancer Lett.* **2001**, *174*, 135.
- 135. Solladié, G.; Boeffel, D.; Maignan, J. Tetrahedron 1996, 52, 2065.
- 136. Lamcharfi, E.; Menguy, L.; Zamarlik, H. Synth. Commun. 1993, 23, 3019.
- 137. Lee, Y. R.; Kim, Y. M. Helv. Chim. Acta 2007, 90, 2401.

































































<u>CHAPTER 3</u>

Synthetic studies of flavones using

pyrrolidine and molecular iodine

catalysts and their

anti-diabetic activity



3.1: Introduction

Flavones 1 (flavus = yellow) also known as 2-phenyl-4*H*-chromen-4-one or 2-phenylchromones (Figure 1) are naturally occurring oxygen heterocyclic compounds belonging to a class of compounds called as flavonoids. These are the secondary metabolites secreted by plants for protection from microbial attack, ultra violet rays and attract insects for pollination. In addition to flavones, flavonoids family comprises of several other members such as flavonois, flavanones, flavans, isoflavones, anthocyanidins, etc. These compounds occur in various parts of plants including bark, grains, stems, fruits, flowers, vegetables, roots, tea and wine.¹



Figure 1: General structure of Flavone 1.

3.2: Occurrence

Owing to their broad range of biological activities continuous investigation has led to the isolation of over 4000 chemically unique flavonoids from plants.² These multifarious naturally occurring compounds occur in free state known as aglycones as well as O- and/or C-glycosides exhibiting simple to complex structure diversity. Some of the common members attributing important and diverse biological activities are listed below (Figure 2, Table 1).



Figure 2: Naturally occurring biologically potent flavones 2.

Table 1: Source of isolation of naturally occurring flavones 2.

2	Name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆
a	Chrysin ³	OH	Н	OH	Н	Н	Н
	Passiflora coerulea L., Passiflora incarnata, Pleurotus ostreatus						
b	Apigenin ⁴	OH	Н	OH	Н	Н	OH
	Perilla frutescens, Matricaria						
---	--------------------------------------	-----	-----	-----	-----	-----	-----
	chamomilla						
с	Luteolin ^{4,5}	OH	Н	OH	Н	OH	OH
	Perilla frutescens, Matricaria						
	chamomilla, Scutellaria lateriflora						
	L., Aiphanes aculeata						
d	Nobiletin ⁶	OMe	OMe	OMe	OMe	OMe	OMe
	Citrus sinensis, Citrus aurantium						
e	Tangeritin ⁶	OMe	OMe	OMe	OMe	Н	OMe
	Citrus sinensis, Citrus aurantium						
f	Scutellarein ⁷	OH	OH	OH	Н	Н	OH
	Asplenium belangeri, Scutellaria						
	baicalensis						
g	Jaceosidin ⁸	OH	OMe	OH	Н	OMe	OH
	Artemisia princeps, Artemisia argyi						
h	Eupatilin ^{8a,9}	OH	OMe	OH	Н	OMe	OMe
	Artemisia princeps, Artemisia						
	asiatica						
i	Baicalein ¹⁰	OH	OH	OH	Н	Н	Н
	Scutellaria baicalensis, Scutellaria						
	lateriflora,						
	Oroxylum indicum						
j	Wogonin ¹¹	OH	Н	OH	OMe	Н	Н
	Scutellaria baicalensis						

Structures of some of the selected naturally occurring flavone glycosides **3-14** are listed below along with their source of isolation (Figure 3, Table 2).



Figure 3: Structures of naturally occurring flavone glycosides 3-14.

 Table 2: Source of isolation of naturally occurring flavone glycosides 3-14.

No.	Name
	Source of isolation
3	Apigenin 7- <i>O</i> -glucoside (Apigetrin) ¹²
	Teucrium gnaphalodes
4	Apigenin 6- <i>C</i> -glucoside (Isovitexin) ¹³
	Serjania erecta
5	Apigenin 8- <i>C</i> -glucoside (Vitexin) ¹³
	Serjania erecta
6	Luteolin 7- <i>O</i> -glucoside (Luteoloside) ¹²
	Teucrium gnaphalodes
7	Luteolin 6- <i>C</i> -glucoside (Isoorientin) ¹⁴
	Gentiana olivieri ,Pueraria tuberosa
8	Luteolin 8- <i>C</i> -glucoside (Orientin) ¹⁵

	Cannabis sativa
9	Chrysin 7- O - β -galactopyranuronoside ¹⁶
	Centaurea pseudoscabiosa, Scutellaria schachristanica
10	Baicalein 7- <i>O</i> -glucuronide (Baicalin) ^{10c,17}
	Scutellaria baicalensis, Oroxylum indicum
11	Wogonin 7- O - β -D-glucuronide (Oroxindin) ¹⁸
	Oroxylum indicum, Bacopa monnieri, Holmskioldia sanguinea
12	Apigenin 7-O-cellobioside ¹⁹
	Salvia uliginosa
13	Apigenin 7,4'- <i>O</i> , <i>O</i> -diglucoside ¹⁹
	Salvia uliginosa
14	Apigenin 7-O-cellobioside-4'-O-glucoside ¹⁹
	Salvia uliginosa

Thousands of flavone compounds isolated have been found to exhibit immense biological activities.²⁰ A broad range of activities exhibited by several members of flavone includes antiinflammatory,²¹ anti-viral,²² estrogenic/anti-estrogenic,²³ anticancer,²⁴ antioxidants,²⁵ leishmanicidal,²⁶ anticonvulsants,^{3a,27} antihistamines,²⁸ ovipositor stimulant phytoalexins,²⁹ anti-HIV,³⁰ vasodilators,³¹ antispasmodics,³² antidiabetics,³³ antimutagenic,³⁴ antiallergic,³⁵ DNA cleavage,³⁶ antiaging,³⁷ antidepressant,³⁸ etc. Some flavonoids are known to show monoamine oxidase (MAO) inhibitory activity³⁹ whereas some have a repelling property against insects acting as antifeedant.⁴⁰

3.3: Literature synthetic methods

The numerous biological activities exhibited by several members of flavones have attracted scientists to constantly study flavones and develop variety of new strategies to synthesize them in large quantities. Among these Baker-Venkataraman rearrangement⁴¹ and Allan-Robinson reaction⁴² are well established methods. Several reviews incorporating diverse routes to flavones have appeared recently.⁴³



Scheme 1: Baker-Venkataraman rearrangement.

Baker-Venkataraman rearrangement⁴¹ involves 2-*O*-benzoylacetophenone as substrate which undergoes intramolecular rearrangement to 2-hydroxydibenzoylmethane on heating. It further on treatment with conc. H_2SO_4 or in boiling AcOH and AcONa gives flavones (Scheme 1).



Scheme 2: Allan-Robinson synthesis.

Allan-Robinson synthesis⁴² involves the reaction of *o*-hydroxyaryl ketones with anhydrides in presence of base to form flavones (Scheme 2).

Other routes to obtain flavones have been broadly divided into 5 categories and retrosynthetically depicted as shown in the following schemes. These 5 classes have been categorized on the basis of i) oxidative cyclization of 2'-hydroxychalcones (Scheme 3A), ii) cyclodehydration of 1-(2-hydroxyphenyl)-3-phenyl- propane-1,3- diones (Scheme 3B), iii) dehydrogenation of flavanones (Scheme 3C), iv) metal catalyzed reactions from various substrates (Scheme 4) and v) miscellaneous routes (Scheme 5).



Scheme 3: A) Oxidative cyclization of 2'-hydroxychalcones; B) Cyclodehydration of 1-(2-hydroxyphenyl)-3-phenyl-propane-1,3-diones; C) Dehydrogenation of flavanones.



Scheme 4: Metal catalyzed reactions from various substrates.



Scheme 5: Miscellaneous routes.

3.3.1: Oxidative cyclization of 2'-hydroxychalcones

Most of the methodologies have been developed by using 2'-hydroxychalcones, 1-(2-hydroxyphenyl)-3-phenyl-propane-1,3-diones and flavanones as substrates. Oxidative

cyclization of 2'-hydroxychalcones using various reagents/catalysts as oxidants have been carried out to form large number of flavone derivatives (Table 3-4).⁴⁴⁻⁷⁴



Table 3: Synthesis of flavones by oxidative cyclization of 2'-hydroxychalcones using vari	ous
reagents.	

Sr.	Reagent	Reaction condition	Ref.
No.			
1)	$Ce(SO_4)_2.4H_2O$	DMSO, 110 °C	44
2)	Silica gel supported	solvent-free, 100 °C	45
	$Ce(SO_4)_2 \cdot 4H_2O$		
3)	ICI-DMSO, ultrasound	50 °C, 30 min	46
4)	Na ₂ SeO ₃ -DMSO	heat/MW	47
5)	Na ₂ TeO ₃ -DMSO	130-140 °C	48
6)	a) Selenium bromide resin, cat.	CH ₂ Cl ₂ , rt, 12 h	49
	ZnCl ₂		
	b) 30 % H ₂ O ₂	THF, 0 °C-rt	
7)	SeO ₂	isoamyl alcohol/DMSO, heat	50
8)	SeO ₂ , SiO ₂	DMSO, MW	51
9)	Sodium perborate tetrahydrate	АсОН, 50-60 °С	52
10)	Ph-S-S-Ph	260-290 °C	53
11)	FeCl ₃ .6H ₂ O	MeOH, reflux	54
12)	a) $NH_4Br-(NH_4)_2S_2O_8$	H ₂ O, grinding, rt	55
	b) Ba(OH) ₂	EtOH, grinding, rt	
13)	Nickel peroxide	benzene	56
14)	PIDA	МеОН, КОН	57
15)	[EtNH ₃]NO ₃	MW, 250 °C	58
16)	Li ₂ PdCl ₄	NaOMe, EtOH/MeCN, rt	59

The diverse reagents used for the oxidative cyclization of 2'-hydroxychalcones are listed in Table 3. Recently, Liu *et al.*⁴⁴ reported Ce(SO₄)₂·4H₂O mediated synthesis of flavones from 2'-hydroxychalcones in DMSO solvent at 100 °C. Later, the same group⁴⁵ developed an environmentally friendly approach using silica gel supported Ce(SO₄)₂·4H₂O as an efficient

reagent under solvent-free at 100 °C for the synthesis of flavones. Iodine monochloride in DMSO solvent under ultrasound irradiation was reported by Lahyani and Trabelsi.⁴⁶ The cyclization reaction was enhanced by ultrasound irradiation with reduced reaction time and temperature with high yields of flavones. Lamba and Makrandi⁴⁷ explored sodium selenitedimethylsulfoxide reagent for this oxidative cyclization under thermal as well as microwave condition. Similarly, Kumar and Sharma⁴⁸ synthesized flavones using sodium tellurite on heating. Huang et al.⁴⁹ reported ZnCl₂ catalysed solid phase oxidative cyclization of 2'hydroxychalcones using polystyrene supported selenium bromide resin. The oxidative cleavage of selenium resins led to flavones. Selenium dioxide has also been used for synthesis of flavones.⁵⁰ Gupta *et al.*⁵¹ later modified the reaction condition by employing selenium dioxide over silica in DMSO solvent under microwave irradiation. Ganguly et al.⁵² utilized excess of sodium perborate tetrahydrate in acetic acid to convert 2'-hydroxychalcones to flavones. Diphenyl disulphide is another reagent reported by Hoshino et al.⁵³ for this conversion. Kumar and Perumal⁵⁴ employed FeCl₃.6H₂O in methanol under reflux condition. Jakhar and Makrandi⁵⁵ prepared α,β -dibromo-2'-hydroxychalcones using ammonium bromide and ammonium persulphate by grinding at room temperature which were converted to flavones by cyclodebromination using barium hydroxide. Nickel peroxide⁵⁶ is also known to give flavones from 2'-hydroxychalcones along with aurones. Flavone product was isolated on treatment of substituted 2'-hydroxychalcone with phenyliodine(III) diacetate (PIDA).⁵⁷ Parveen⁵⁸ carried out flavone synthesis in ethyl ammonium nitrate ([EtNH₃]NO₃) under microwave irradiation. Also, on treatment of lithium chloropalladite (Li₂PdCl₄) with sodium salt of 2'-hydroxychalcones in polar solvent such as ethanol or acetonitrile delivered flavones.⁵⁹ However expensive metal used as reagent and flavones accompanied with small amounts of flavanone makes it less efficient.

Sr.	Catalyst	Reaction condition	Ref.
No.			
1)	CuI	[Bmim][NTf ₂], O ₂ (1 atm), 50 °C	60
2)	CuI	DMA, 130 °C	61
3)	NH ₄ I	solvent-free, 120 °C	62
4)	Oxalic acid	EtOH, reflux	63
5)	Silica gel supported InBr ₃ and	solvent-free, 130-140 °C	64
	InCl ₃		

Table 4: Synthesis of flavones by oxidative cyclization of 2'-hydroxychalcones using various catalysts.

6)	I ₂ -SiO ₂	80 °C	65
7)	Na ₂ PdCl ₄ .3H ₂ O (10 mol%)	NaOAc, AcOH, t-BuOH:H ₂ O (1:1), t-	66
		BuOOH, 70 °C	
8)	I ₂	DMSO, reflux	67
9)	I ₂	triethylene glycol, 150 °C	68
10)	I ₂	solvent-free, 110-130 °C	69
11)	I ₂ -Al ₂ O ₃	solvent-free, MW, 400 W	70
12)	I ₂ (20 mol%)	DMSO, MW	71

Besides these reagents several catalysts have also been reported for the synthesis of flavones from chalcones (Table 4). CuI catalyzed ionic liquid [Bmim][NTf₂] (1-butyl-3methylimidazolium *bis*(trifluoromethanesulfonyl)imide) mediated *oxa*-Michael-oxidation was disclosed by Wang and co-workers⁶⁰ to deliver flavones in good yields. Recently, Chen and Liu⁶¹ also employed CuI in *N*,*N*-dimethylacetamide solvent to synthesize flavones. Varala and co-workers⁶² demonstrated the catalytic activity of NH₄I as an alternative source for iodine in this oxidative cyclization under solvent-free condition. Zambare *et al*.⁶³ carried out the cyclization using oxalic acid catalyst giving flavones in good yields.

Ahmed *et al.*⁶⁴ employed silica gel supported InBr₃ and InCl₃ catalyst for the conversion of 2'hydroxychalcones to flavones under solvent-free condition. Madhavarao and co-workers⁶⁵ carried out flavones synthesis using silica gel supported iodine (I₂-SiO₂) as an heterogeneous catalyst at 80 °C. Lorenz *et al.*⁶⁶ developed a catalytic oxidation procedure employing Na₂PdCl₄.3H₂O catalyst and excess of *t*-BuOOH. Iodine is one of the widely used reagent for flavones synthesis. Catalytic amount of iodine in DMSO solvent,⁶⁷ triethylene glycol at 150 °C⁶⁸ and solvent-free condition at 110-130 °C⁶⁹ are some of the conditions used to deliver flavones.

Also microwave irradiation was reported by several groups to yield flavones in short duration of time. Sarda *et al.*⁷⁰ used I₂-Al₂O₃ as an heterogeneous catalyst under solvent-free microwave condition. Our group^{71a} also carried out microwave assisted synthesis of flavones using iodine catalyst in DMSO solvent. Later, Borse *et al.*^{71b} and Belsare and Kazi^{71c} focused on the comparative study of flavones synthesis using iodine catalyst by conventional and microwave methods.



Scheme 6

Bose *et al.*⁷² reported an environmentally benign synthesis of natural flavones in 3 steps from 2'-acetoxychalcones through bromination using *n*-tetrabutylammonium tribromide (TBATB) followed by dehydrobromination and finally cyclization (Scheme 6).





Lokhande *et al.*⁷³ carried out one pot deprotection of 2'-allyloxychalcones followed by oxidative cyclization on treatment with catalytic iodine in DMSO solvent to deliver flavones. Recently they have also reported flavones synthesis from 2'-allyloxy- α - β -dibromochalcones *via* deallylation and cyclization leading to the formation of 3-bromoflavanones followed by dehydrobromination using same oxidative reaction condition⁷⁴ (Scheme 7).

3.3.2: Cyclodehydration of 1-(2-hydroxyphenyl)-3-phenyl-propane-1,3-diones



Table 5: Synthesis of flavones by cyclodehydration of 1-(2-hydroxyphenyl)-3-phenyl-propane-1,3-diones by using various reagent.

Sr.	Reagent	Reaction condition	Ref.
No.			
1)	[EtNH ₃]NO ₃	MW, 22-50 sec	75
2)	[bmim]BF ₄	100 °C, 45-75 min	76
3)	AcOH/H ₂ SO ₄	rt/100 °C	77
4)	Conc. H ₂ SO ₄	CH ₃ CN, rt	78
5)	AcOH/HC1	100 °C	79
6)	AcOH/HI	100 °C, 2 h	80
7)	I ₂	DMSO, heat	81

8)	P ₂ O ₅	grinding, rt, 10-15 min	82
9)	<i>p</i> -TSA	grinding, rt	83
10)	Br ₂ /I ₂	hv, CHCl ₃ /MeOH	84
11)	Montmorillonite K-10	MW, 1-1.5 min	85
12)	NH ₄ OAc	solvent-free, heat/MW	86
13)	Silica-H ₂ SO ₄	grinding, rt, 8-9 h	87
14)	Malic acid	solvent-free, heat/MW	88
15)	CuBr ₂	DMF, 130 °C	89

Another extensively used starting material for flavones synthesis has been 1-(2hydroxyphenyl)-3-phenyl-propane-1,3-diones which can be converted in presence of various reagents/catalysts via cyclodehydration (Table 5-6). Pawar and co-workers⁷⁵ carried out flavones synthesis from this substrate in ionic liquid ethyl ammonium nitrate [EtNH₃]NO₃ under microwave irradiation. Similarly, ionic liquid 1-butyl-3-methylimidazolium tetrafluoroborate [bmim]BF₄ at 100 °C delivered flavones.⁷⁶ Acetic acid in presence of conc. H₂SO₄⁷⁷ was employed to carry out cyclodehydration. Later, conc. H₂SO₄⁷⁸ in acetonitrile solvent has also been used. Acetic acid containing hydrochloric acid⁷⁹ or hydriodic acid⁸⁰ is also known to give flavones. Heating in presence of iodine in DMSO⁸¹ is another way to obtain flavones. Grinding technique was used by Sharma and Makrandi employing phosphorous pentoxide⁸² and by Jakhar and Makrandi using p-toluenesulphonic $acid^{83}$ to synthesize flavone derivatives. Photochemical transformation using Br₂/CHCl₃ or I₂/MeOH⁸⁴ is also known. Solid state cyclodehydration was reported by Varma et al.⁸⁵ using montmorillonite K-10 under microwave irradiation. Recently, ammonium acetate promoted solvent-free synthesis of flavones under thermal and microwave condition.⁸⁶ Also, silicasulphuric acid was utilized as a heterogeneous acidic reusable medium by solvent-free grinding method at room temperature.⁸⁷ Natural organic acid like malic acid⁸⁸ promoted the solvent-free synthesis of flavones under conventional thermal and microwave heating condition. 3-Bromoflavones were synthesized by employing CuBr₂ in DMF.⁸⁹

Table	6:	Synthesis	of	flavones	by	cyclodehydration	of	1-(2-hydroxyphenyl)-3-phenyl-
propan	e-1,	,3-diones by	y us	ing variou	is ca	talysts.		

Sr. No.	Catalyst	Reaction condition	Ref.
1)	Tungstophosphoric/molybdoph	АсОН, 90 °С	90

CHAPTER 3

	osphoric acids (bulk/silica		
	supported)		
2)	Trifluoromethanesulfonic acid	toluene, reflux	91
	(supported on titanium,		
	calcined at 100 °C)		
3)	Trifluoromethanesulfonic acid	toluene, reflux	92
	(supported on carbon)		
4)	$H_6P_2W_{18}O_{62}.24H_2O$	bulk- solvent-free & silica supported-	93
	(bulk/silica supported)	toluene, 110 °C	
5)	H ₆ P ₂ W ₁₈ O ₆₂ .24H ₂ O (silica	MW, solvent-free	94
	supported)		
6)	Preyssler, Keggin	solvent-free, 110 °C or CHCl ₃ , reflux	95
	heteropolyacids & Preyssler		
	(silica supported)		
7)	Keggin heteropolyacids	CHCl ₃ , reflux or acetonitrile reflux or	96
		solvent-free, 110 °C	
8)	P and Si Keggin	MeCN	97
	heteropolyacids		
9)	Amberlyst 15	isopropyl alcohol, reflux	98
10)	NaHSO ₄ -silica	toluene, reflux, 1-4 h	99
11)	KHSO ₄ , silica	high speed ball milling, 5-15 min	100
12)	KHSO ₄	solvent-free, 120 °C	101
13)	FeCl ₃	CH_2Cl_2, rt	102
14)	Silica-PCl ₅	solvent-free	103
15)	$TiO_2/H_3PW_{12}O_{40}$	toluene or solvent-free, 110 °C	104
16)	Ga(OTf) ₃	MeNO ₂ /DCE, 80 °C	105
17)	InCl ₃	toluene, reflux	106
18)	L-Ascorbic acid	solvent-free heat/MW	107
19)	bis-	CH ₂ Cl ₂ , 0 °C to rt, 1.5-3.5 h	108
	(Trichloromethyl)carbonate/		
	DMF		
20)	CuCl ₂	EtOH, MW, 80 °C, 5 min	109
21)	Proline	MeOH, H ₂ O, 55 °C	110
	phenylsulphonylhydrazide or		

	pyrrolidine		
22)	Co(salpr)(OH)	MeOH, 60 °C	111
23)	N-Triflyl phosphoramide	MeOH, 40 °C, 16 h	112

Tungstophosphoric acid (H₃PW₁₂O₄₀.nH₂O) or molybdophosphoric acid (H₃PMo₁₂O₄₀.nH₂O) both bulk or supported on silica were explored as catalysts for flavones synthesis.⁹⁰ Trifluoromethanesulfonic acid supported on titanium after calcined at 100 °C catalyzed cyclodehydration reaction to yield flavones.⁹¹ Trifluoromethanesulfonic acid supported on carbon was also found to catalyze this reaction.⁹² Wells-Dawson heteropolyacid H₆P₂W₁₈O₆₂.24H₂O either as bulk under solvent-free or silica supported in toluene at 110 °C was also employed as catalyst.⁹³ Solvent-free synthesis using silica supported H₆P₂W₁₈O₆₂.24H₂O was also employed under microwave irradiation.⁹⁴

Preyssler structured heteropolyacid catalysts such as $H_{14}[NaP_5W_{30}O_{110}]$, $(H_{14}P_5)$, $H_{14}[NaP_5W_{29}MoO_{110}]$, (H_{14} - P_5Mo) and Keggin structured heteropolyacid catalyst H₃[PW₁₂O₄₀] successfully delivered flavones under solvent-free conditions. Also silica supported Preyssler catalysts $H_{14}[NaP_5W_{30}O_{110}]$, $(H_{14}P_5/SiO_2 50 \%, H_{14}P_5/SiO_2 40 \%)$ $H_{14}P_5/SiO_2$ 30 %) in refluxing chloroform are effective. ⁹⁵ Keggin heteropolyacids such as tungstophosphoric acid (H₃PW₁₂O₄₀.nH₂O), tungstosilicic acid (H₄SiW₁₂O₄₀.nH₂O), molybdophosphoric acid (H₃PMo₁₂O₄₀.nH₂O) and molybdosilicic acid (H₄SiMo₁₂O₄₀.nH₂O) without or with dehydration at 100 and 200 °C for 6 h were utilized to synthesize flavone in homogeneous condition in refluxing acetonitrile as well as in heterogeneous condition in refluxing toluene. Also molybdophosphoric acid calcined at 100 °C successfully delivered substituted flavones in refluxing acetonitrile and under solvent-free conditions.⁹⁶ P and Si Keggin heteropolyacids have also been utilized to obtain flavones.⁹⁷ Amberlyst 15, a nonaqueous cation exchange resin is also known to deliver flavones.⁹⁸ Kucukislamoglu et al.99 developed flavones synthesis using silica gel supported sodium bisulphate (NaHSO4) catalyst in toluene. Later, Su and co-workers¹⁰⁰ explored the catalytic activity of potassium bisulphate (KHSO₄) in presence of silica to synthesize flavones using high speed ball milling technique. Recently, Romanelli and co-workers¹⁰¹ investigated the catalytic activity and recyclability of KHSO₄ to synthesize flavones under solvent-free conditions.

Catalytic FeCl₃ was also effective in the dehydrative cyclization reaction.¹⁰² Also an efficient and reusable silica-PCl₅ as solid acid catalyst was utilized to afford flavones under solvent-free conditions.¹⁰³ Mesoporous titania/tungstophosphoric acid (TiO₂/H₃PW₁₂O₄₀, 10 w/w) composite was prepared and investigated by Pérez *et al.*¹⁰⁴ to synthesize flavones in toluene or solvent-free conditions at 110 °C. Catalytic gallium(III) triflate¹⁰⁵ on heating at 80 °C in

nitromethane or dichloroethane converted 1-(2-hydroxyphenyl)-3-phenyl-propane-1,3-diones to flavones. Similarly, indium(III) chloride¹⁰⁶ in toluene at reflux can be used to carry out intramolecular cyclization. Recently, L-ascorbic acid¹⁰⁷ promoted synthesis of flavones under conventional and microwave heating. Flavones can also be synthesized under Vilsmeier-Haack conditions using *bis*-(trichloromethyl)carbonate and dimethylformamide.¹⁰⁸ Microwave irradiation in presence of 10 mol% CuCl₂¹⁰⁹ is another way to acquire flavones. A facile organocatalytic approach employing proline phenylsulphonylhydrazide or pyrrolidine has been developed recently by Yang and co-workers.¹¹⁰ Co(salpr)(OH)¹¹¹ complex is also known to promote cyclization leading to flavones. Recently, an organocatalytic method for the dehydrative cyclization utilizing *N*-triflyl phosphoramide has been developed.¹¹²



Scheme 8

Ismail and Aziem¹¹³ acylated 2-(*t*-butyldimethylsilyloxy)-4-methoxyacetophenone with acyl chloride using lithium diisopropylamide to form 1-[2'-(t-butyldimethylsilyloxy)-3-substituted propane-1,3-diones. These on treatment with glacial acetic acid containing H₂SO₄ on heating delivered flavones through the cleavage of the silyl protecting group followed by cyclization (Scheme 8).



Scheme 9

Fu and co-workers¹¹⁴ developed an intramolecular Ullmann-type O-arylation method which converted substituted 1-(2-haloaryl)-propane-1,3-diones to flavones using K₂CO₃ base in DMF solvent (Scheme 9).



Scheme 10

Also, Fu and co-workers¹¹⁵ developed a methodology employing K_2CO_3 as a catalyst for flavones synthesis in moderate to good yields. This transition metal free base catalyzed

intramolecular nucleophilic substitution reaction involves selective cleavage of the aromatic C-O bond of 1-(2-alkoxyphenyl)-3-alkylpropane-1,3-diones (Scheme 10).

3.3.3: Dehydrogenation of flavanones



Table 7: Synthesis of flavones from dehydrogenation of flavanone by using various reagents.

Sr.	Reagent	Reaction condition	Ref.
No.			
1)	SeO ₂	xylene, 140-150 °C, 5 h	50a
2)	LTA	90 °C, 2.5 h	116
3)	DDQ	dioxane/ AcOH	117
4)	Ph-S-S-Ph	260-290 °C	53
5)	Nickel peroxide	benzene	56
6)	2,4,6-Triphenylpyrylium	CH ₂ Cl ₂ , hv	118
	tetrafluoroborate		
7)	Tl(NO ₃) ₃	MeOH, rt	119
8)	Tl(OAc) ₃	AcOH or MeOH or MeCN	120
9)	Mn(OAc) ₃ ,	HClO ₄ , AcOH, 100 °C, 1 h	121
10)	NBS, cat. AIBN	MW, 10 min	122
11)	Na ₂ SeO ₃ -DMSO	heat/MW	47
12)	Nano Fe ₃ O ₄ , NiO ₂	toluene, 90 °C, 24 h flow reactor	123
13)	PPh ₃ .HBr	DMSO, 50 °C	124
14)	PhI(OAc) ₂	MeOH	125
15)	PhI(OAc) ₂	0.1 N KOH in MeOH, MW, 4-5 min	126
16)	[Hydroxy(tosyloxy)iodo]benzene	MeOH	127
17)	[Hydroxy(tosyloxy)iodo]benzene	([bbim] ⁺ Br ⁻), 60-70 °C, 2-4 h	128
18)	IBX	DMSO, 90 °C, 24 h	129
19)	CuCl ₂ .2H ₂ O	DMSO, 110 °C	130

Flavanone is another widely used substrate to synthesize flavones *via* dehydrogenation. Several reagents/catalysts have been explored for this conversion (Table 7-8). Selenium dioxide was developed many decades ago to carry out oxidation of flavanones.^{50a} Similarly, lead tetraacetate can also be used.¹¹⁶ DDQ in dioxane or acetic acid was an effective reagent for dehydrogenation reaction.¹¹⁷ Diphenyl disulphide⁵³ is also used for this conversion. Also, the use of nickel peroxide was demonstrated.⁵⁶ Photosensitized dehydrogenation of flavanones to flavones was carried out using 2,4,6-triphenylpyrylium tetrafluoroborate.¹¹⁸ Thallium(III) nitrate¹¹⁹ and thallium(III) acetate¹²⁰ have also been explored for this conversion. Singh *et al.*¹²¹ heated manganese(III) acetate in presence of perchloric acid in acetic acid to synthesize flavones. A highly selective transformation of flavanones to 3bromoflavones or flavones using *N*-bromosuccinimide (NBS) in presence of catalytic amount of 2,2'-azobis(isobutyronitrile) under microwave irradiation was reported by Yang and coworkers.¹²²

Sodium selenite-dimethylsulfoxide⁴⁷ combination is also effective for this oxidation process under thermal as well as microwave condition. Flavanone oxidation was carried out by inductive heating of Fe_3O_4 nanoparticles and nickel peroxide as solid oxidant in fixed bed reactors.¹²³ Recently, the applicability of PPh₃.HBr-DMSO reagent system was unveiled by Das and co-workers¹²⁴ in the flavone synthesis.

Hypervalent iodine reagent such as (diacetoxyiodo)benzene has also been utilized in flavanone oxidation.¹²⁵ Later, it was also studied under microwave irradiation.¹²⁶ Similarly, [hydroxy(tosyloxy)iodo]benzene¹²⁷ in methanol was explored for this reaction and it was also effective in presence 1,3-di-*n*-butylimadazolium bromide ([bbim]⁺Br⁻) ionic liquid.¹²⁸ The ability of 2-iodoxybenzoic acid (IBX) has also been examined for flavanone conversion to flavones.¹²⁹ Lokhande *et al.*¹³⁰ employed copper (II) chloride for oxidative aromatization of flavanones to obtain flavones in high yields.

Sr.	Catalyst	Reaction condition	Ref.
No.			
1)	I ₂	DMSO	67a
2)	I ₂	conc. H ₂ SO ₄ , DMSO, 100 °C	131
3)	Silica gel supported InBr ₃ and InCl ₃	solvent-free, 130-140 °C, 120 min	64
4)	CuI	[Bmim][NTf ₂], O ₂ (1 atm), 50 °C, 18 h	60
5)	Pd(DMSO) ₂ (TFA) ₂	O ₂ (1 atm), AcOH, 100 °C	132
6)	Pd(TFA) ₂ /4,5-diazaflurenone	O ₂ (1 atm), DMSO, 100 °C	133

Table 8: Synthesis of flavones from dehydrogenation of flavanone by using various catalysts.

Several catalysts have also been reported for dehydrogenation of flavanones (Table 8). Catalytic iodine^{67a, 131} in DMSO solvent alone or in presence of a drop of conc. H₂SO₄ has been used. Silica gel supported InBr₃ and InCl₃ has also been employed at 130-140 °C to oxidize flavones under solvent-free conditions.⁶⁴ Wang and co-workers⁶⁰ demonstrated the conversion of parent flavanone to flavone using 10 mol% of CuI in [Bmim][NTf₂] ionic liquid. Diao and Stahl¹³² revealed the application of Pd(DMSO)₂(TFA)₂ as catalyst using oxygen as an oxidant. Later, another catalytic system containing Pd(TFA)₂ and 4,5-diazaflurenone was also developed to carry out flavanone oxidation.¹³³

3.3.4: Metal catalyzed reactions from various substrates



Scheme 11

Gogoi and co-workers¹³⁴ carried out Ru(II)-catalyzed C-H activation and annulation reaction between salicylaldehydes and terminal/internal alkynes to afford variety of flavones. Broad substrate scope, lower catalyst loading and high regioselectivity are some of the advantages of this methodology (Scheme 11).



Scheme 12

Liu *et al.*¹³⁵ synthesized flavones by a Pd-catalyzed regioselective intramoleular nucleophilic substitution of *gem*-dichloroalkene derivatives with salicylaldehydes. It works well to deliver various flavones in presence of triphenylphosphine sulphide ligand, sodium carbonate base and benzyltrietylammonium chloride as additive (Scheme 12).



Scheme 13

Maiti *et al.*¹³⁶ demonstrated dual catalytic role of $FeCl_3$ -Lewis acid and piperidine-an organocatalyst for one pot synthesis of flavones. It involves *o*-hydroxy aromatic aldehydes and phenyl acetylenes as substrates (Scheme 13).



Scheme 14

Mizuno and co-workers¹³⁷ developed a one pot flavone synthesis from 2'hydroxyacetophenones and benzaldehydes using gold nanoparticles supported on a Mg-Al layered double hydroxide (Au/LDH) catalyst in moderate to good yields (Scheme 14).



Scheme 15

Zhu *et al.*¹³⁸ developed a highly efficient and selective flavone synthesis by Pd/C catalyzed ligand-free cyclocarbonylation reaction between 2-iodophenols and terminal acetylenes in CO atmosphere. Various derivatives were synthesized in excellent yields. Also catalyst was reusable (Scheme 15).



Scheme 16

Carbene complex palladium(II) benzimidazolin-2-ylidene [PdBr₂(i Pr₂-bimy)L] with co-ligand L= *N*-phenylimidazole has been used as a catalyst for flavones synthesis by Li and co-workers¹³⁹ from 2-iodophenols and phenyl acetylenes (Scheme 16).



Scheme 17

Miao and Yang¹⁴⁰ reported a regiospecific carbonylative annulation of *o*-iodophenol acetates and acetylenes using catalytic medium containing palladium-thiourea-dppp complex, base and CO at 40 °C (Scheme 17).



Awuah and Capretta¹⁴¹ synthesized flavones through palladium catalyzed microwave-assisted sequential Sonogashira and carbonylative annulation reactions from aryl halide, TMS acetylene and iodophenols in presence of 1,3,5,7-tetramethyl-2,4,8-trioxa-6-phenyl-6-phosphaadamantane (PA-Ph) ligand (Scheme 18). Also they separately synthesized flavones from carbonylative cyclization of 2-iodophenol and terminal alkynes under this reaction condition (Scheme 19).



Scheme 20

Yang and Alper¹⁴² synthesized flavones through ligand-free palladium catalyzed cyclocarbonylation of *o*-iodophenols with terminal acetylenes in presence of CO and phosphonium salt ionic liquid $C_{14}H_{29}(C_6H_{13})_3P^+Br^-$. Different derivatives of flavones were obtained in good to excellent yields (Scheme 20).





Liang *et al.*¹⁴³ carried out palladium catalyzed sequential carbonylative coupling of *o*iodophenols with terminal acetylenes to obtain α,β -unsaturated ketones which underwent an intramolecular cyclization to deliver flavones (Scheme 21). The competing side reaction leading to aurones was not observed.





Figure 4

A simple and efficient Pd-catalyzed cascade carbonylative cyclization of 2-bromophenols and phenylacetylenes for the synthesis of flavones was reported by Liu *et al.*¹⁴⁴ The ligand employed in the reaction medium is benzimidazole-triazole (Figure 4) and dipropylamine as solvent (Scheme 22).



Wu and co-workers¹⁴⁵ developed a carbonylative synthesis of flavones from phenols and internal alkynes using Ir catalyst in presence of $Cu(OAc)_2$ as oxidant and *bis*(2-methoxyphenyl)(phenyl)phosphane L₁ (Figure 5) as ligand (Scheme 23).



Scheme 24

Wu and co-workers¹⁴⁶ developed a palladium catalyzed carbonylative synthesis of flavones in moderate to good yields from easily available 2'-hydroxyacetophenones and (hetero)aryl bromides in presence of 1,4-*bis*(diphenylphosphino)butane (DPPB) ligand, DBU base and DMSO solvent. This approach is better than other carbonylative reactions as they involve expensive substrates such as 2-iodophenols and terminal/internal alkynes (Scheme 24).



Scheme 25

An oxidative Heck reaction was reported by Jafarpour and Khoobi *et al.*¹⁴⁷ It is an atom economical base-free palladium catalyzed reaction allowing regioselective direct arylation of chromenones to give flavones (Scheme 25).



Scheme 26

A palladium catalyzed 1,4-addition of arylboronic acids to chromones has been developed by Hong and co-workers¹⁴⁸ In addition to Pd catalyst, $Fe(OTf)_3$, DDQ and KNO₂ in catalytic amounts were required to obtain flavones in good yields (Scheme 26).





Pd(II) trifluoroacetate catalyst was employed in the flavones synthesis by Hong and coworkers.¹⁴⁹ It involves oxidative cross coupling of chromones and non-activated arenes *via* twofold C-H functionalization in conjunction of AgOAc/CsOPiv in pivalic acid (PivOH) (Scheme 27).



Scheme 28

Kim *et al.*¹⁵⁰ developed a palladium catalyzed oxidative arylation of chromones to give flavones in moderate to good yields. It involves a regioselective 2-arylation of chromone *via* a double C-H activation process (Scheme 28).





Regioselective zincation at C-2 using TMP₂Zn.2MgCl₂.2LiCl (TMP = 2,2,6,6-tetramethylpiperidide) was applied for the synthesis of naturally occuring flavones by Klier *et al.*¹⁵¹ The *bis*-heterocyclic zinc intermediate formed was converted to desired flavone products in presence of Pd catalyst, aryl iodide and *tris*-(2-furyl)phosphine (tfp) (Scheme 29).



Scheme 30

Two flavone derivatives were synthesized along with several other compounds by Muto *et al.*¹⁵² *via* decarbonylative organoboron cross coupling of corresponding phenyl ester with phenyl boronic acids using nickel catalyst (Scheme 30).



Scheme 31

Lee *et al.*¹⁵³ reported a one pot synthesis of flavones from chromanones and arylboronic acid pinacol esters by palladium catalyzed dehydrogenation and oxidative boron-Heck coupling. In addition, the methodology is also useful for synthesis of naturally occurring apigenin and luteolin (Scheme 31).



Scheme 32

Kraus and Gupta¹⁵⁴ devised a synthetic route to flavones from phenol and 3,3-dichloroacrylic acid using DCC and DMAP. It was converted into ketone by Fries rearrangement using AlCl₃

which was further on treatment with dilute base afforded chromones. Finally Suzuki reaction delivered flavones in good yields (Scheme 32).



Scheme 33

Palladium catalyzed sequential dehydrogenation/arylation of chromanones was carried out successfully by Hong and co-workers¹⁵⁵ in presence of Cu and Ag oxidants. The intermediate chromones formed undergoes oxidative cross-coupling with arenes *via* a twofold C-H functionalization leading to flavones (Scheme 33).



Scheme 34

Jiang *et al.*¹⁵⁶ reported a direct intramolecular acylation of esters using catalytic FeCl₃ with 1,1-dichloromethyl methyl ether (Cl₂CHOCH₃) to yield flavones (Scheme 34).

3.3.5: Miscellaneous routes



Scheme 35

A convenient one pot synthesis of 6-bromo-7-hydroxyflavones was developed by Yakovenko *et al.*¹⁵⁷ from hydrolysis of the substituted 4-ethoxy-6-bromo-7-hydroxyflavylium salts. These salts were obtained either from the corresponding chalcones or by condensation of 5-bromoresacetophenone with the corresponding aryl aldehydes using ethyl orthoformate in the

presence of perchloric acid (Scheme 35). This method has also been utilised for the preparation of a library of synthetic 4'-hydroxyflavones exhibiting casein kinase 2 (CK2) inhibition activity.¹⁵⁸



Scheme 36

Seijas *et al.*¹⁵⁹ carried out microwave irradiation of phloroglucinol and β -ketoesters under solvent-free condition to deliver flavones. It proceeds *via* a cycloaddition of α -oxo ketene intermediate which then undergoes thermal Fries rearrangement in absence of any catalyst (Scheme 36).



Scheme 37

Larock and co-workers¹⁶⁰ developed ICl induced cyclization of alkynones to 3-iodoflavones in CH_2Cl_2 solvent at low temperature (Scheme 37).



Scheme 38

An efficient method for flavones synthesis was developed by Chuang *et al.*¹⁶¹ from 6-endo cyclization of *o*-alkynoylphenyl acetates using 18-crown-6 ether in moderate to good yields (Scheme 38).





Similarly, flavones were synthesized in excellent yields *via* 6-endo cyclization of acylated *o*-alkynoylphenols by Yang *et al.*¹⁶² using piperazine catalyst (Scheme 39).



Scheme 40

Doi and co-workers¹⁶³ demonstrated DMAP catalyzed 6-endo cyclization of 1-(2-hydroxyphenyl)-2-propyn-1-ones in good yields (Scheme 40).



Scheme 41

6-Endo cyclization of *o*-alkynoylphenol compounds was demonstrated by Doi and coworkers^{164a} to desired flavone products under acidic condition using trifluoromethanesulfonic acid (TfOH). Also demethylation was observed in 3-methoxy substrates to provide 5-hydroxy flavones derivatives. However, later Taylor and Bolshan^{164b} carried out similar cyclization of 3-methoxy-2-alkynoylphenol substrates in excess of trifluoromethanesulfonic acid to provide 5-methoxy flavone derivatives without demethylation (Scheme 41).



Also Zhang *et al.*¹⁶⁵ developed LiO*t*Bu mediated synthesis of flavones from substituted 2-(1-hydroxy-3-phenylprop-2yn-1-yl)phenol *via* 6-*endo-dig* cyclization (Scheme 42).



Scheme 43

Resin capture method employing piperazinyl Merrifield resin was developed by Brueggemeier and co-workers¹⁶⁶ for the flavones synthesis. It involves reaction of alkynyl ketone with piperazine thethered to a solid support to form support bound enaminones which undergoes cyclization to deliver final product (Scheme 43).



Scheme 44

Brueggemeier and co-workers¹⁶⁷ carried out the cyclization of alkynones on refluxing with 10 equiv of diethylamine to yield flavones *via* enaminoketone intermediate. Dimethylamine and *N*-benzylethylamine also gave similar results (Scheme 44).



Scheme 45

Agarwal and Soni¹⁶⁸ converted 2'-hydroxy chalcone dibromide to flavones by heating in triethanolamine or aqueous triethylamine for 10-15 min (Scheme 45).



Scheme 46

Oxidative conversion of chromenes to flavones in excellent yields was demonstrated by Banerjee *et al.*¹⁶⁹ using *tert*-butylhydroperoxide and catalytic copper bromide (Scheme 46).



Scheme 47

Nagata and co-workers¹⁷⁰ reported KMnO₄ oxidation of chromenes to flavones in acetone solvent at room temperature (Scheme 47).



Scheme 48

Ghodile¹⁷¹ synthesized flavones from substituted 1-(2-hydroxy-5-chlorophenyl)ethyldiphenylamines with heptanedioylchloride to form intermediate which was later dehydrogenated using Pd/C (Scheme 48).



Scheme 49

Varma and Kumar¹⁷² used clay supported nitrite salts "clayfen or clayan" for microwave thermolysis of thioketones under solvent-free condition to form flavones (Scheme 49).



Scheme 50

A method developed by Kumar and Bodas¹⁷³ utilised acylphosphoranes prepared by treating silyl ester of O-acyl(aroyl)salicylic acids with (trimethylsilyl)methylenetriphenylphosphorane to furnish flavones in good to excellent yields *via* intramolecular Wittig reaction (Scheme 50).



Scheme 51

Similarly, intramolecular photochemical Wittig reaction in water was developed by Das and Ghosh¹⁷⁴ to synthesize flavones from suitable starting materials (Scheme 51).

3.4: Results and Discussion

Literature studies showed various routes for the synthesis of flavones. Recently flavanone synthesis is reported using aniline and catalytic amount of iodine from 2'-hydroxyacetophenone and aryl aldehydes.¹⁷⁵ Also it is well known that 2'-hydroxychalcone/flavanone get cyclised to flavone using catalytic iodine in DMSO solvent.^{67,71,131} In view of this we speculated that it should be possible to devise a new synthetic route to flavones directly from aryl aldehyde and 2'-hydroxyacetophenone in one pot (Scheme 52). However the use of iodine catalyst from simple substrates in one pot was so far not reported.



Scheme 52: Retrosynthetic analysis of flavones 1.

We envisioned that a combination of secondary amine and iodine as oxidant would be a perfect combination for obtaining flavone **1** in one pot. The secondary amine would catalyze the aldol reaction of 2'-hydroxyacetophenone **16** and aromatic aldehydes **17** and also further the intramolecular Michael reaction of the resulting 2'-hydroxychalcone **15** and finally iodine

would bring about the oxidation of *in situ* formed flavanone **1'**. However sequential catalysis has its own limitation. For example, in the present case, firstly iodine and secondary amine should no way interfere with their roles of catalyst in aldol reaction and oxidation i.e. iodine should not poison secondary amine or vice versa. Secondly, regeneration of secondary amine and iodine *via* oxidation of HI by DMSO should also be not affected. Thirdly, α -iodination followed by Kornblum oxidation should not take place. Lastly, the formation of frequently encountered side product aurone, the flavone isomer was to be prevented (Scheme 53).



Scheme 53: Various possible products in the synthesis of flavone 1.

We commenced our work by carrying out a reaction between 2'-hydroxyacetophenone **16a** and 3,4-dimethoxybenzaldehyde **17a** as model substrates in presence of secondary amine and catalytic amount of iodine in DMSO solvent under reflux condition (Scheme 54).



Scheme 54: Synthesis of flavone 1.

At the outset, various secondary amines such as pyrrolidine, L-proline, piperidine, *N*-methylaniline and morpholine were screened individually (Table 9). To our delight, a new spot was observed when the reaction was carried out using pyrrolidine in DMSO solvent. Its isolation and characterization showed the formation of the desired flavone **1a** in 75 % yield (entry 1). The structure of **1a** was confirmed by the following spectral data.

2-(3,4-Dimethoxyphenyl)-4H-chromen-4-one (1a)



colorless solid; m.p. 155-157 °C; lit.^{164a} 156-157 °C.

IR (KBr): $\tilde{v} = 3061, 2841, 1643$ (C=O), 1514, 1465, 1145 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.99 (s, 3H), 4.01 (s, 3H), 6.85 (s, 1H), 7.01 (d, *J* = 8.4 Hz, 1H), 7.42 (s, 1H), 7.45 (t, *J* = 8.0 Hz, 1H), 7.60 (d, *J* = 8.4 Hz, 2H), 7.73 (t, *J* = 8.0 Hz, 1H), 8.25 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 56.2 (2 X OCH₃), 106.0 (CH), 108.9 (CH), 111.2 (CH), 118.1 (CH), 120.4 (CH), 123.4 (Cq), 123.9 (Cq), 125.5 (CH), 125.7 (CH), 134.0 (CH), 149.4 (Cq), 152.4 (Cq), 156.2 (Cq), 164.2 (Cq), 178.4 (Cq).

Other amines such as L-proline and piperidine were also effective to deliver the desired product but with diminished yields (entries 2-3). However, *N*-methylaniline showed two minor spots (not characterized) along with the unreacted substrates whereas morpholine was unreactive under given reaction condition and failed to give the desired product (entries 4-5).

Sr. No.	Secondary amine (0.5 equiv)	Time (h)	Yield (%) ^a
1)	Pyrrolidine	2	75
2)	L-proline	3	36
3)	Piperidine	3	22
4)	<i>N</i> -methylaniline	2	00 b
5)	Morpholine	2	00°

Table 9: Screening of secondary amines in the flavone formation.

^a Isolated yields of **1a**.

^b Two minor spots were formed.

^c No reaction.

With the successful product formation, standardization of various parameters such as pyrrolidine concentration, temperature (neat/DMSO), solvents and iodine concentration were examined in succession. Firstly, the standardization of pyrrolidine was investigated by performing the reaction in absence of iodine. The reaction furnished a new spot which was

characterized as 2-(3,4-dimethoxyphenyl)chroman-4-one **1a'** (Scheme 55) using the following spectral data.

2-(3,4-Dimethoxyphenyl)chroman-4-one (1a')



colorless solid; m.p. 120-122 °C; lit.¹⁷⁶ 123-124 °C.

IR (KBr): $\tilde{v} = 3010, 2837, 1687$ (C=O), 1598, 1026 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 2.88 (dd, *J* = 16.8, 2.4 Hz, 1H), 3.13 (dd, *J* = 16.8, 13.6 Hz, 1H), 3.91 (s, 3H), 3.93 (s, 3H), 5.43 (dd, *J* = 13.2, 2.0 Hz, 1H), 6.91 (d, *J* = 8.0 Hz, 1H), 7.01-7.08 (m, 4H), 7.51 (t, *J* = 8.4 Hz, 1H), 7.93 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 44.6 (CH₂), 55.9 (OCH₃), 56.0 (OCH₃), 79.6 (CH), 109.4 (CH), 111.1 (CH), 118.2 (CH), 118.8 (CH), 120.9 (Cq), 121.6 (CH), 127.1 (CH), 131.2 (Cq), 136.2 (CH), 149.3 (Cq), 149.4 (Cq), 161.6 (Cq), 192.2 (Cq).

The exclusive formation of flavanone **1a'** without flavone formation suggested the mandatory role of iodine as oxidant and discounting the possibilities of oxidation taking place due to dissolved oxygen or DMSO. Also, when flavanone **1a'** (prepared separately) was subjected to heating in DMSO in absence of iodine no flavone formation was observed on TLC even after prolonged heating, thus indicating the role of iodine as an exclusive oxidant.

The amount of pyrrolidine was varied from 0.1-1.5 equiv (Table 10). 0.1 and 0.2 equiv of pyrrolidine were not very effective and resulted in incomplete conversion even after refluxing for prolonged reaction time (entries 1-2). On increasing the concentration to 0.3 equiv the reaction completed in 45 min (entry 3). Similarly, the reaction completed in 15 min when 0.5 and 1 equiv of pyrrolidine were screened (entries 4-5). Further increase to 1.5 equiv reduced the time considerably to 5 min (entry 6). Based on these observations 0.5 equiv of pyrolidine was found to be the optimum concentration and was selected for further studies.



Scheme 55: Synthesis of flavanone 1a'.

Sr. No.	Pyrrolidine (equiv)	Time (min)	Result
1)	0.1	120	Incomplete reaction
2)	0.2	120	Incomplete reaction
3)	0.3	45	Reaction completed
4)	0.5	15	Reaction completed
5)	1.0	15	Reaction completed
6)	1.5	5	Reaction completed

Table 10: Standardization of pyrrolidine in absence of iodine.

Table 11: Temperature study under neat and in DMSO solvent.

		OMe		OMe OMe
	OH	OMe pyrrolidine (0.5 equiv)		OMe
	— + н	iodine (0.1 equiv)		or
10	6a 0	17a	0	1a 1a' 0
Sr.	Solvent	Temperature	Time	Result
No.		(° C)	(min)	
1)	Neat	60	24	Formation of 1a'
				No formation of 1a
2)	Neat	100	24	Formation of 1a'
				No formation of 1a
3)	Neat	150	24	No reaction
4)	DMSO	room temperature	48	No reaction
5)	DMSO	60	24	Formation of 1a'
				No formation of 1a
6)	DMSO	100	24	Formation of 1a ' and little 1a
7)	DMSO	150	6	Formation of 1a (80 %)
8)	DMSO	reflux	2	Formation of 1a (60 %)

Temperature study was carried out under neat condition as well as in DMSO solvent (Table 11). Neat or solvent-less reaction was carried out at different temperatures such as 60, 100 and 150 °C. At 60 °C, the formation of flavanone **1a**' was observed but the desired product **1a** was not seen even after heating for 24 h (entry 1). This may be attributed to the high temperature required for the conversion of flavanone **1a**' to flavone **1a**. Similar results were obtained at 100 °C (entry 2). Further increasing the temperature to 150 °C did not show any

formation of **1a'** or **1a** which may be because of the loss of pyrrolidine from the reaction mixture prior to commencement of reaction owing to high temperature (entry 3).

Next, the temperature studies were performed in DMSO solvent. At room temperature, there was no reaction even after stirring for 48 h (entry 4). On heating at 60 °C, formation of flavanone **1a'** was seen but no flavone **1a** was formed even after heating for 24 h (entry 5). Similarly, at 100 °C, showed formation of flavanone along with the detection of trace amount of flavone **1a** (not isolated) (entry 6). This may be attributed to the high temperature needed for the formation of flavone **1a** from flavanone **1a'**. Further heating the reaction mixture to 150 °C exclusively offered flavone **1a** in 80 % yield in 6 h (entry 7) whereas on refluxing the reaction, the yield got reduced to 60 % (entry 8). Hence, 150 °C was found to be the optimum temperature.

[OH OH H H H H O O 17a OMe pyrrolidine (0.5 equiv) iodine (0.1 equiv) solvent, reflu		OMe
Sr. No.	Solvent (reflux)	Time (h)	Yield (%)
1)	Ethanol	24	00
2)	Methanol	24	00
3)	Toluene	24	00
4)	Xylene	24	00
5)	Tetrahydrofuran	24	00
6)	DMSO	2	75

Table 12: Solvent screening.

Solvent study was performed by carrying out the reaction in various solvents (Table 12). Ethanol, methanol, toluene, xylene and tetrahydrofuran were screened but none of these solvents showed any product formation even after refluxing for 24 h (entries 1-5). The reaction was effective only in DMSO solvent (entry 6).

Table 13: Optimization of iodine concentration at 150 °C.



Sr. No.	Iodine (equiv)	Time (h)	Yield (%)
1)	0.01	24	30
2)	0.05	10	88
3)	0.1	6	80
4)	0.15	7	76
5)	0.2	6	38
6)	1.0	24	ND

ND: Not Determined.

With these observations in hand we persuaded to detect the ideal iodine concentration. It was optimized by varying its concentration from 0.01-1 equiv (Table 13). When 0.01 equiv was employed 30 % of product **1a** was obtained after 24 h (entry 1). A slight increase to 0.05 equiv delivered maximum yield of 88 % (entry 2). Similarly on increasing the iodine concentration to 0.1, 0.15 and 0.2 equiv resulted in 80, 76 and 38 % yields respectively (entries 3-5). 1 equiv of iodine showed trace amount of product on TLC even after 24 h, hence yield of the product was not determined (entry 6). Thus, 5 mo% of iodine was considered as standard iodine concentration.



Scheme 56: Optimum reaction condition used for flavones synthesis.

Sr.	2'-Hydroxy-	Substituted	Time	Product	Yield
No.	acetophenone	aromatic	(h)	(1)	(%)
	(16 a)	aldehydes (17)			
1)	OH	OMe OHC OMe	10	OMe OMe OMe	88
	16a	17a		0	
				1 a	

Table 14: Screening of aromatic aldehydes.

2)	OH O 16a	OHC OMe 17b	24	OMe O 1b	82
3)	OH O 16a	OHC OMe OHC OMe	7	OMe OMe OMe OMe	78
4)	он О 16а	онс 17d	9	o Id	85
5)	OH O 16a	онс F 17e	9	Ie	82
6)	он о 16а	OHC CI	12	Cl Cl If	84
7)	С ОН О 16а	OHC Br 17g	13	Br O 1g	80
8)	I6a	OHC Br 17h	6	Br Br 1h	70
9)	С ОН О 16а	онс NO ₂ 17і	12		62

10)	OH O 16a	онс 17ј	24	lij	74
11)	OH O 16a	OHC 17k	9	lk	80
12)	OH O 16a	OHC S 171	6		75
13)	OH O 16a	CHO S 17m	8	o Im	72

On obtaining the ideal reaction condition (Scheme 56) in hand, we subjected 2'hydroxyacetophenone 16a with various aromatic aldehydes 17a-m to check the feasibility of our one pot method (Table 14). Electron rich aromatic aldehydes 17a-c furnished desired flavones **1a-c** in good yields (entries 1-3). Benzaldehyde **17d** also smoothly reacted to form required product 1d in good yield (entry 4). Aromatic aldehydes with p-halogen atoms (fluoro, chloro, bromo) were well tolerated to provide flavones 1e-1g in good yields which are good scaffolds for further functionalization (entries 5-7). *m*-Bromobenzaldehyde 17h as well as *m*-nitrobenzaldehyde 17i with electron withdrawing groups at *m*-position delivered flavones 1h and 1i but with slightly diminished yields (entries 8-9). Thus our methodology could be applied to both electron rich as well as electron deficient aromatic aldehydes which are well tolerated under the reaction condition as the yields were unchanged on changing the substituent. Furthermore, 3,4-methylenedioxy benzaldehyde 17j smoothly furnished the desired flavone 1j in satisfactory yield (entry 10). Reports have shown that the biological activity of flavones is enhanced when 5 or 6 membered heterocyclic group is attached at its C-2 position.^{160,177} Motivated from this we subjected different heterocyclic aromatic aldehydes to the reaction condition to achieve the desired products 1k-1m in good yields (entries 11-13).

Sr.	Substituted 2'-	Substituted	Time	Product	Yield
No.	hydroxy-	aromatic	(h)	(1)	(%)
	acetophenone	aldehydes (17)			
	(16b-e)				
1)	MeO OH	OHC	8	MeO O OMe	81
	16b	17b		ہ 1n	
2)	MeO OH	OHC	10	MeO	88
	16b			10	
3)		OHC 17d	12		78
	Ive			1p	
4)	Ph_O_OH O 16d	OHC 17d	12	Ph_O_O O	60
				1q	
5)		OHC	8		70
	100			1r	

Table	15:	Screening	of 2'-hydro	oxvaceto	ohenones
Lanc	10.	Dereening	or 2 - fryund	JAyaceto	sitementes.

After scanning aromatic aldehydes, substituted 2'-hydroxyacetophenones **16b-e** were put forth for determining the substrate scope (Table 15). 4-Methoxy-2'-hydroxyacetophenone **16b** was reacted with benzaldehydes **17b** and **17d** to provide flavones **1n-o** in good yields (entries 14-15). Similarly, 4-ethoxy-2'-hydroxyacetophenone **16c** and 4-benzyloxy-2'-hydroxyacetophenone **17d** reacted under standardized condition to furnish respective flavones **1p** and **1q** in reasonable yields (entries 16-17). One of the reports had shown deprotection of 2'-allyloxychalcone leading to flavone in I₂-DMSO.⁷³ Interestingly, we got the desired flavone **1r** from 4-allyloxy-2'-hydroxyacetophenone **16e** without the cleavage of the allyloxy group (entry 18). It could be due to the difference in the position of the allyloxy group as 2'-

position is comparatively more reactive one. The reaction protocol was also successfully scaled up to 5 g of starting aryl aldehyde **17a** to get consistent yield of desired flavone **1a**. Although the literature survey shows a one pot method¹⁵⁷ with perchloric acid and ethyl chloroformate (Scheme 35) but our approach is an alternative route avoiding the use of explosive perchloric acid.

Spectral data of flavones

2-(4-Methoxyphenyl)-4H-chromen-4-one (1b)



yield (0.152 g, 82 %); colorless solid; m.p. 154-156 °C; lit.⁶⁵ 154-156 °C.

IR (**KBr**): $\tilde{v} = 3051$, 1641 (C=O), 1465, 1377, 1026 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.88 (s, 3H), 6.77 (s, 1H), 7.02 (d, *J* = 8.8 Hz, 2H), 7.41 (t, *J* = 7.6 Hz, 1H), 7.55 (d, *J* = 8.4 Hz, 1H), 7.68 (t, *J* = 8.4 Hz, 1H), 7.88 (d, *J* = 8.8 Hz, 2H), 8.22 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.5 (OCH₃), 106.1 (CH), 114.5 (2 X CH), 117.9 (CH), 123.9 (Cq), 123.9 (Cq), 125.1 (CH), 125.7 (CH), 128.0 (2 X CH), 133.6 (CH), 156.2 (Cq), 162.4 (Cq), 163.5 (Cq), 178.5 (Cq).

2-(3,4,5-Trimethoxyphenyl)-4*H*-chromen-4-one (1c)



yield (0.179 g, 78 %); colorless solid; m.p. 175-177 °C; lit.^{164a} 176-178 °C.

IR (**KBr**): $\tilde{v} = 3076$, 1641 (C=O), 1467, 1367, 1128 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 3.94 (s, 3H), 3.97 (s, 6H), 6.79 (s, 1H), 7.14 (s, 2H), 7.43 (t, J = 7.6 Hz, 1H), 7.59 (d, J = 8.4 Hz, 1H), 7.71 (t, J = 8.0 Hz, 1H), 8.23 (d, J = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 56.3 (2 X OCH₃), 61.1 (OCH₃), 103.7 (2 X CH), 107.4 (CH), 118.1 (CH), 123.9 (Cq), 125.3 (CH), 125.7 (CH), 126.9 (Cq), 133.8 (CH), 141.2 (Cq), 153.6 (2 X Cq), 156.2 (Cq), 163.3 (Cq), 178.4 (Cq).
2-Phenyl-4H-chromen-4-one (1d)



yield (0.139 g, 85 %); colorless solid; m.p. 94-96 °C; lit.^{164a} 95-97 °C.

IR (KBr): $\tilde{v} = 3070, 1643$ (C=O), 1465, 1375, 1130 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 6.88 (s, 1 H), 7.45 (t, *J* = 7.6 Hz, 1 H), 7.53-7.61 (m, 4 H), 7.73 (t, *J* = 8.8 Hz, 1 H), 7.94-7.96 (m, 2 H), 8.25 (dd, *J* = 8.0, 1.2 Hz, 1 H).

¹³C NMR (CDCl₃, 100 MHz): δ 107.6 (CH), 118.1 (CH), 123.9 (Cq), 125.3 (CH), 125.7 (CH), 126.4 (2 X CH), 129.1 (2 X CH), 131.7 (CH), 131.7 (Cq), 133.9 (CH), 156.3 (Cq), 163.5 (Cq), 178.6 (Cq).

2-(4-Fluorophenyl)-4H-chromen-4-one (1e)



yield (0.145 g, 82 %); colorless solid; m.p. 144-146 °C; lit.¹⁷⁸ 145-148 °C.

IR (KBr): $\tilde{v} = 3076$, 1662 (C=O), 1508, 1234, 835 cm⁻¹.

¹**H NMR** (**CDCl**₃, **400 MHz**): δ 6.78 (s, 1H), 7.22 (t, *J* = 8.4 Hz, 2H), 7.43 (t, *J* = 7.6 Hz, 1H), 7.56 (d, *J* = 8.4 Hz, 1H), 7.71 (t, *J* = 7.6 Hz, 1H), 7.93 (t, *J* = 7.6 Hz, 2H), 8.23 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 107.3 (CH), 116.2 (CH), 116.4 (CH), 118.0 (CH), 123.8 (Cq), 125.3 (CH), 125.7 (CH), 127.8 (Cq), 127.9 (Cq), 128.4 (CH), 128.5 (CH), 133.9 (CH), 156.1 (Cq), 162.3 (Cq), 163.5 (Cq), 165.9 (Cq), 178.3 (Cq).

2-(4-Chlorophenyl)-4H-chromen-4-one (1f)



yield (0.159 g, 84 %); colorless solid; m.p. 186-188 °C; lit.¹⁷⁹ 187-188 °C.

IR (**KBr**): $\tilde{v} = 3072$, 1639 (C=O), 1467, 1375, 1093, 754 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 6.79 (s, 1H), 7.40 (t, *J* = 7.6 Hz, 1H), 7.47 (d, *J* = 8.0 Hz, 2H), 7.53 (d, *J* = 8.4 Hz, 1H), 7.69 (t, *J* = 8.0 Hz, 1H), 7.83 (d, *J* = 8.4 Hz, 2H), 8.20 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 107.5 (CH), 118.0 (CH), 123.8 (Cq), 125.4 (CH), 125.7 (CH), 127.5 (2 X CH), 129.4 (2 X CH), 130.1 (Cq), 133.9 (CH), 137.9 (Cq), 156.1 (Cq), 162.2 (Cq), 178.4 (Cq).

2-(4-Bromophenyl)-4H-chromen-4-one (1g)



yield (0.176 g, 80 %); colorless solid; m.p. 174-177 °C; lit.¹⁸⁰ 177 °C.

IR (KBr): $\tilde{v} = 3086$, 1666 (C=O), 1465, 1259, 1130 cm⁻¹.

¹**H NMR** (**CDCl**₃, **400 MHz**): δ 6.79 (s, 1H), 7.37 (t, *J* = 7.2 Hz, 1H), 7.50 (d, *J* = 8.4 Hz, 1H), 7.59 (d, *J* = 8.4 Hz, 2H), 7.65 (dt, *J* = 8.8, 1.6 Hz, 1H), 7.73 (d, *J* = 8.4 Hz, 2H), 8.16 (dd, *J* = 8.0, 1.6 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 107.7 (CH), 118.1 (CH), 123.9 (Cq), 125.4 (CH), 125.9 (CH), 126.4 (Cq), 127.8 (2 X CH), 130.7 (Cq), 132.4 (2 X CH), 133.9 (CH), 156.2 (Cq), 162.4 (Cq), 178.3 (Cq).

2-(3-Bromophenyl)-4H-chromen-4-one (1h)



yield (0.154 g, 70 %); colorless solid; m.p. 113-115 °C; lit.¹⁸⁰ 115 °C.

IR (KBr): $\tilde{v} = 3064$, 1641 (C=O), 1467, 1261, 1128 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 6.77 (s, 1H), 7.30-7.38 (m, 2H), 7.51 (d, J = 8.4 Hz, 1H), 7.59 (d, J = 8.0 Hz, 1H), 7.65 (dt, J = 8.4, 1.6 Hz, 1H), 7.76 (d, J = 8.0 Hz, 1H), 8.00 (s, 1H), 8.15 (dd, J = 8.0, 1.2 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 108.1 (CH), 118.2 (CH), 123.3 (Cq), 123.8 (Cq), 124.9 (CH), 125.6 (CH), 125.8 (CH), 129.3 (CH), 130.6 (CH), 133.7 (Cq), 134.2 (CH), 134.6 (CH), 156.2 (Cq), 161.9 (Cq), 178.4 (Cq).

2-(3-Nitrophenyl)-4H-chromen-4-one (1i)



yield (0.122 g, 62 %); colorless solid; m.p. 194-197 °C; lit.¹⁴⁸ 195-198 °C.

IR (KBr): $\tilde{v} = 3086$, 1643 (C=O), 1527, 1350, 1138 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 6.87 (s, 1H), 7.40 (t, J = 8.0 Hz, 1H), 7.57 (d, J = 8.0 Hz, 1H), 7.67-7.72 (m, 2H), 8.17 (dd, J = 7.6, 1.6 Hz, 2H), 8.33 (dd, J = 8.0, 1.6 Hz, 1H), 8.75 (t, J = 1.6 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 108.8 (CH), 118.2 (CH), 121.3 (CH), 123.8 (Cq), 125.8 (CH), 125.9 (CH), 126.0 (CH), 130.4 (CH), 131.8 (CH), 133.6 (Cq), 134.4 (CH), 148.8 (Cq), 156.2 (Cq), 160.6 (Cq), 178.2 (Cq).

2-(1,3-Benzodioxol-5-yl)-4H-chromen-4-one (1j)



yield (0.145 g, 74 %); colorless solid; m.p. 196-198 °C; lit.^{49a} 198-200 °C.

IR (**KBr**): $\tilde{v} = 3078$, 1643 (C=O), 1446, 1346, 1026 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 6.09 (s, 2H), 6.72 (s, 1H), 6.95 (d, *J* = 8.0 Hz, 1H), 7.38 (s, 1H), 7.42 (t, *J* = 7.6 Hz, 1H), 7.50-7.56 (m, 2H), 7.69 (t, *J* = 8.4 Hz, 1H), 8.22 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 101.9 (CH₂), 106.3 (CH), 106.6 (CH), 108.8 (CH), 117.9 (CH), 121.5 (CH), 123.9 (Cq), 125.2 (CH), 125.7 (CH, Cq), 133.7 (CH), 148.5 (Cq), 150.7 (Cq), 156.1 (Cq), 163.1 (Cq), 178.4 (Cq).

2-(Furan-3-yl)-4H-chromen-4-one (1k)



yield (0.125 g, 80 %); colorless solid; m.p. 118-119 °C; lit.¹⁸¹ 119 °C.

IR (KBr): $\tilde{v} = 3118$, 1631 (C=O), 1463, 1357, 756 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 6.54 (s, 1H), 6.76 (s, 1H), 7.42 (d, *J* = 5.6 Hz, 1H), 7.50 (s, 1H), 7.56 (s, 1H), 7.69 (d, *J* = 5.6 Hz, 1H), 8.09 (s, 1H), 8.22 (s, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 107.2 (CH), 107.6 (CH), 117.9 (CH), 120.3 (Cq), 123.9 (Cq), 125.2 (CH), 125.7 (CH), 133.8 (CH), 143.1 (CH), 144.7 (CH), 156.0 (Cq), 158.8 (Cq), 178.2 (Cq).

2-(Thiophen-2-yl)-4H-chromen-4-one (11)



yield (0.126 g, 75 %); colorless solid; m.p. 90-92 °C; lit.¹⁸² 91-94 °C.

IR (**KBr**): $\tilde{v} = 3072$, 1631 (C=O), 1460, 1261, 1126 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 6.71 (s, 1H), 7.11 (t, *J* = 4.0 Hz, 1H), 7.35 (t, *J* = 7.6 Hz, 1H), 7.46 (d, *J* = 8.4 Hz, 1H), 7.52 (d, *J* = 5.2 Hz, 1H), 7.62 (dt, *J* = 8.4, 2.0 Hz, 1H), 7.67 (d, *J* = 3.6 Hz, 1H), 8.13 (dd, *J* = 8.0, 1.6 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 104.9 (CH), 116.9 (CH), 122.7 (Cq), 124.4 (CH), 124.7 (CH), 127.6 (CH), 127.8 (CH), 129.6 (CH), 132.9 (CH), 133.9 (Cq), 154.9 (Cq), 158.4 (Cq), 176.9 (Cq).

2-(Benzo[b]thiophen-3-yl)-4H-chromen-4-one (1m)



yield (0.147 g, 72 %); colorless solid; m.p. 142-144 °C.¹⁴⁶

IR (**KBr**): $\tilde{v} = 3082$, 1629 (C=O), 1465, 1220, 1118 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 6.77 (s, 1H), 7.35-7.47 (m, 3H), 7.51 (d, J = 8.4 Hz, 1H), 7.63-7.67 (m, 1H), 7.87 (d, J = 8.0 Hz, 1H), 8.04 (s, 1H), 8.18-8.22 (m, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 109.4 (CH), 118.0 (CH), 123.2 (CH), 123.4 (CH), 124.0 (Cq), 125.4 (CH), 125.5 (CH), 125.8 (CH), 128.9 (Cq), 130.5 (CH), 133.9 (CH), 135.5 (Cq), 140.7 (Cq), 156.3 (Cq), 160.4 (Cq), 178.4 (Cq).

7-Methoxy-2-(4-methoxyphenyl)-4H-chromen-4-one (1n)



yield (0.168 g, 81 %); colorless solid; m.p. 140-142 °C; lit.¹⁸⁰ 143 °C.

IR (KBr): $\tilde{v} = 2983$, 1653 (C=O), 1438, 1265, 1184 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.81 (s, 3H), 3.86 (s, 3H), 6.76 (s, 1H), 6.89-6.95 (m, 4H), 7.80 (d, *J* = 8.8 Hz, 2H), 8.05 (d, *J* = 8.4 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.5 (OCH₃), 55.9 (OCH₃), 100.4 (CH), 105.5 (CH), 114.5 (2 X CH), 114.6 (CH), 117.2 (Cq), 123.8 (Cq), 126.9 (CH), 128.1 (2 X CH), 158.0 (Cq), 162.5 (Cq), 163.7 (Cq), 164.4 (Cq), 177.8 (Cq).

<u>7-Methoxy-2-phenyl-4*H*-chromen-4-one (10)</u>



yield (0.163 g, 88 %); colorless solid; m.p. 104-106 °C; lit.^{164a} 105-106 °C.

IR (KBr): $\tilde{v} = 3059, 1654$ (C=O), 1438, 1274, 1165 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.96 (s, 3H), 6.81 (s, 1H), 7.00-7.04 (m, 2H), 7.54-7.56 (m, 3H), 7.93-7.96 (m, 2H), 8.15-8.18 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.8 (OCH₃), 100.3 (CH), 107.3 (CH), 114.5 (CH), 117.6 (Cq), 126.1 (2 X CH), 126.9 (CH), 128.9 (2 X CH), 131.5 (CH), 131.6 (Cq), 157.9 (Cq), 163.0 (Cq), 164.2 (Cq), 177.9 (Cq).

7-Ethoxy-2-phenyl-4H-chromen-4-one (1p)



yield (0.153 g, 78 %); colorless solid; m.p. 136-138 °C; lit.¹⁸³ 138-139 °C. **IR (KBr):** $\tilde{v} = 2983$, 1631 (C=O), 1494, 1246, 1180 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 1.42 (t, J = 6.8 Hz, 3H), 4.09 (q, J = 6.8 Hz, 2H), 6.70 (s, 1H), 6.88-6.92 (m, 2H), 7.43-7.47 (m, 3H), 7.82-7.85 (m, 2H), 8.06 (d, J = 8.8 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 14.6 (CH₃), 64.3 (CH₂), 100.8 (CH), 107.5 (CH), 114.8 (CH), 117.7 (Cq), 126.2 (2 X CH), 126.9 (CH), 129.0 (2 X CH), 131.4 (CH), 131.9 (Cq), 158.0 (Cq), 162.9 (Cq), 163.6 (Cq), 177.9 (Cq).

7-(Benzyloxy)-2-phenyl-4H-chromen-4-one (1q)



yield (0.144 g, 60 %); colorless solid; m.p. 173-175 °C; lit.¹⁸⁴ 174-175 °C. **IR (KBr):** $\tilde{v} = 3066, 1635$ (C=O), 1450, 1253, 1180 cm⁻¹. ¹**H NMR (CDCl₃, 400 MHz):** δ 5.11 (s, 2H), 6.81 (d, *J* = 6.8 Hz, 1H), 6.99 (s, 1H), 7.01 (s, 1H), 7.29-7.45 (m, 8H), 7.83 (d, *J* = 7.2 Hz, 2H), 8.07 (d, *J* = 8.4 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 70.6 (CH₂), 101.5 (CH), 107.3 (CH), 115.2 (CH), 117.7 (Cq), 126.3 (2 X CH), 127.2 (CH), 127.6 (2 X CH), 128.5 (CH), 128.8 (2 X CH), 129.1 (2 X CH), 131.7 (CH), 135.7 (Cq), 158.0 (Cq), 163.5 (Cq), 177.9 (Cq).

7-Allyloxy-2-phenyl-4H-chromen-4-one (1r)



yield (0.143 g, 70 %); colorless solid; m.p. 94-96 °C; lit.¹⁸⁵ 95-96 °C.

IR (KBr): $\tilde{v} = 3061$, 1629 (C=O), 1450, 1261, 1166 cm⁻¹.

¹**H NMR** (**CDCl**₃, **400 MHz**): δ 4.59-4.61 (m, 2H), 5.28-5.32 (m, 1H), 5.38-5.44 (m, 1H), 5.97-6.07 (m, 1H), 6.72 (s, 1H), 6.91-6.96 (m, 2H), 7.45-7.48 (m, 3H), 7.83-7.86 (m, 2H), 8.07 (d, *J* = 8.8 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 68.3 (CH₂), 100.3 (CH), 106.4 (CH), 113.9 (CH), 116.8 (Cq), 117.5 (CH₂), 125.2 (2 X CH), 126.1 (CH), 127.9 (2 X CH), 130.4 (CH), 130.7 (Cq), 131.0 (CH), 156.9 (Cq), 162.1 (2 X Cq), 176.9 (Cq).

The probable mechanism was proposed for the synthesis of flavones (Scheme 57). Aromatic aldehyde **17** forms iminium ion **18** by reacting with secondary amine. The substrate **16** can enolise due to which its enolic form can act as a nucleophile attacking **18** to form intermediate **19**. Regeneration of pyrrolidine results in chalcone **20** which undergoes Michael addition to form flavanone **1'** in presence of pyrrolidine. Further reaction with iodine leads to the formation of iodo intermediate **21**. Alternatively **21** could also be obtained directly from **20** in presence of iodine. The loss of HI results in the completely oxidized required flavone **1**. The steps involved in the regeneration of pyrrolidine and iodine catalysts plays an important role for the successful formation of flavones.



Scheme 57: Probable mechanism for the formation of flavone 1 *via* chalcone 20 and flavanone 1'.

Regarding the proposed mechanism, hydrogen release by dissociation of HI is a well known reaction. This is supported by our observation of formation of 1-(2-hydroxyphenyl)-3-(pyridin-2-yl)propan-1-one **22** along with the corresponding flavanone **1s'** and flavone **1s** when 2'-hydroxyacetophenone **16a** and 2-pyridinecarboxaldehyde **17n** were subjected to this protocol (Scheme 58). 1-(2-Hydroxyphenyl)-3-(pyridin-2-yl)propan-1-one **22** was obtained due to the reduction of the intermediate chalcone by the liberated hydrogen gas. However, we do admit that so far in literature to our knowledge whenever people have used iodine as an oxidant there is no clear cut mention of decomposition of HI to I₂ and H₂ though it is a well established reaction and formation of HI is invoked to explain iodine catalyzed reactions. May be it is considered as an obvious pathway whenever iodine is used as an oxidant for aromatization reaction.



Scheme 58: Reaction of 2'-hydroxyacetophenone and 2-pyridinecarboxaldehyde led to formation of the reduced product 1-(2-hydroxyphenyl)-3-(pyridin-2-yl)propan-1-one 22 along with the corresponding flavanone 1s' and flavones 1s.

Spectral data of 22, 1s' and 1s

1-(2-Hydroxyphenyl)-3-(pyridin-2-yl)propan-1-one (22)



yield (0.033 g, 10 %); pale yellow liquid.¹⁸⁶

IR (neat): $\tilde{v} = 3053, 2927, 1643$ (C=O), 1487, 752 cm⁻¹.

¹**H NMR** (**CDCl**₃, **400 MHz**): δ 3.17 (t, J = 7.2 Hz, 2H), 3.48 (t, J = 7.2 Hz, 2H), 6.79-6.83 (m, 1H), 6.89 (d, J = 8.4 Hz, 1H), 7.06 (dd, J = 7.2, 5.6 Hz, 1H), 7.19 (d, J = 7.8 Hz, 1H), 7.36-7.40 (m, 1H), 7.52-7.56 (m, 1H), 7.77 (dd, J = 8.0, 1.6 Hz, 1H), 8.45 (d, J = 4.8 Hz, 1H). ¹³**C NMR** (**CDCl**₃, **100 MHz**): δ 30.7 (CH₂), 36.3 (CH₂), 117.4 (CH), 117.9 (CH), 118.4 (Cq), 120.4 (CH), 122.4 (CH), 129.0 (CH), 135.4 (CH), 135.6 (CH), 148.1 (CH), 159.1 (Cq), 161.2 (Cq), 204.4 (Cq).

2-(Pyridin-2-yl)chroman-4-one (1s')



yield (0.031 g, 9 %); colorless solid; m.p. 64-68 °C; lit.¹⁸⁷ 68 °C.

IR (KBr): $\tilde{v} = 3064$, 2926, 1693 (C=O), 1606, 1462, 1305, 763 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.06-3.16 (m, 2H), 5.60 (dd, *J* = 10.8, 5.2 Hz, 1H), 6.98-7.03 (m, 2H), 7.26-7.29 (m, 1H), 7.44-7.48 (m, 1H), 7.59 (d, *J* = 8.0 Hz, 1H), 7.75-7.79 (m, 1H), 7.87 (dd, *J* = 8.0, 2.0 Hz, 1H), 8.58 (d, *J* = 4.8 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 42.8 (CH₂), 79.1 (CH), 118.1 (CH), 121.2 (Cq), 121.4 (CH), 121.9 (CH), 123.8 (CH), 127.1 (CH),136.2 (CH), 138.3 (CH), 148.4 (CH), 157.1 (Cq), 160.8 (Cq), 191.2 (Cq).

2-(Pyridin-2-yl)-4H-chromen-4-one (1s)



yield (0.01 g, 3 %); colorless solid; m.p. 120-122 °C.¹³⁹

IR (KBr): $\tilde{v} = 3007, 2927, 1643$ (C=O), 1465, 1381, 756 cm⁻¹.

¹**H NMR** (**CDCl**₃, **400 MHz**): δ 7.37 (s, 1H), 7.38-7.41 (m, 1H), 7.54 (dd, *J* = 7.2, 5.2 Hz, 1H), 7.67-7.69 (m, 2H), 8.00 (t, *J* = 7.6, 1H), 8.12 (d, *J* = 7.6 Hz, 1H), 8.18 (dd, *J* = 8.0, 1.2 Hz, 1H), 8.81 (d, *J* = 4.0 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 108.3 (CH), 117.3 (CH), 120.4 (CH), 123.3 (Cq), 124.5 (CH), 124.8 (CH), 124.9 (CH), 133.1 (CH), 137.2 (CH), 147.7 (Cq), 148.3 (CH), 155.1 (Cq), 159.4 (Cq), 177.4 (Cq).

3.5: Anti-diabetic activity

Diabetes mellitus (DM) is a metabolic and heterogeneous disorder caused by inherited and/or acquired insulin secretion deficiency and/or by lowered responsiveness of the organs to the secreted insulin, affecting approximately 5 % of total global population.¹⁸⁸ Among the two major types of diabetes, Type I diabetes mellitus also known as insulin dependent diabetes mellitus (IDDM) commonly occurs in children and accounts to only 5-10 % of total diabetic patients. However, Type II diabetes mellitus also known as non-insulin dependent diabetes mellitus (NIDDM) is highly related to diet or lifestyle and is prevalent in adults, called as adult-onset diabetes. The high blood-glucose level arises due to high consumption of carbohydrate enriched diet leading to hyperglycemia in affected individuals. In a study done by Shaw *et al.*¹⁸⁹ 6.4 % of adults of world (aged 20–79 years) were found to be affected in 2010 and it is estimated to increase to 7.7 % by 2030. Recently, Guariguata *et al.*¹⁹⁰ conducted a literature search depicting diabetes prevalence globally for 2013 and estimated the greatest increase by 55 % up to 2035.

Herbal drugs have been a major source of medicines over the centuries for the prevention and cure of various diseases including diabetes mellitus. There are more than 200 species of plants that exhibit hypoglycaemic properties and its use along with drugs including insulin helps to lower the drug dosage and/or decrease the frequency of drug administration with reduced side effects.¹⁹¹ But the detection of anti-diabetic compound and purification from plant crude extracts is a tedious work allowing the isolation of anti-diabetic compounds only in trace amounts from herbs due to which the number of drugs is limited. Number of

natural compounds belonging to major classes of chemical compounds like alkaloid, carbohydrate, terpenoids, flavonoids and phenolic compounds extracted from various plant medicated sources are reported to be potential inhibitors for the two enzymes *viz.* α -glucosidase and α -amylase responsible for Type 2 DM. In the process of screening, anti-diabetic compounds are all natural compounds such as pycnogenol, galegine (isolated from plants) and acarbose, miglitol, and voglibose (isolated from microbes) which were found to demonstrate inhibitory activity of these enzymes.¹⁹² Zhao *et al.*¹⁹³ investigated the anti-diabetic activity of flavone, the main pharmacological ingredient isolated from *Ipomoea batatas* leaf. It was found effective against non-insulin dependent diabetes mellitus (NIDDM) in rats.

Natural compound can be modified into more potential derivatives on the basis of structure activity relationship (SAR) studies. Various dose dependent inhibition assays have been shown that synthetic derivatives show more potent α -glucosidase inhibitory activity than their parent compound/s.¹⁹⁴ For example, salacinol derivatives inhibited rat small intestinal α -glucosidase more strongly than the naturally occurring salacinol.¹⁹⁵ Few natural as well as synthetic flavones have been studied for anti-diabetic activity.³³ This encouraged us to evaluate the flavones **1a-r** synthesized in our laboratory for their anti-diabetic activity.

The synthesized 18 flavones **1a-r** were submitted for *in vitro* inhibition of α -glucosidase activity in the Marine Biotechnology Department of Goa University. These derivatives were firstly dissolved in methanol solvent (1 mg/mL). It was observed that only 5 derivatives **1c**, **1d**, **1i**, **1j** and **1r** were completely soluble whereas the remaining derivatives were partially soluble forming aggregates in the solution. Hence these partially soluble derivatives were excluded from further studies. The selected compounds **1c**, **1d**, **1i**, **1j** and **1r** were then evaluated for their *in vitro* inhibition of α -glucosidase activity.

The selected samples **1c**, **1d**, **1i** and **1r** showed α -glucosidase inhibition of 74, 53, 98 and 93 % respectively at 300 µg/mL, however, 2-(1,3-benzodioxol-5-yl)-4*H*-chromen-4-one **1j** showed highest inhibitory activity of 99 % at 300 µg/mL. Further, the dose dependent studies of these 5 samples showed proper graph pattern only for **1j**, hence it was selected for further studies.

α -Glucosidase inhibition assay:

In vitro studies using **1j** demonstrated remarkable inhibition of α -glucosidase suggesting the presence of potential enzyme inhibiting activity of a synthetic compound. Significant α -glucosidase inhibitions were depicted at all the doses of **1j** in comparison to their respective

controls (t-test) (Figure 6). Dose dependent inhibition of α -glucosidase by **1j** ranged from 8.4 \pm 0.37 % at 1 µg/mL to 99.3 \pm 0.26 % at 7.6 µg/mL. Further ANOVA of the percentage inhibition of α -glucosidase observed in the control and different doses of **1j** showed significant difference at P < 0.0001.



Figure 6: Percentage of inhibition of α -glucosidase in presence of various doses of **1j**. Data are mean \pm SD (*P < 0.05, **P < 0.01, ***P < 0.001 denotes statistically significant difference from the test control as determined by student's t-test significance).

Enzyme Kinetic studies

Enzyme kinetic study results were analyzed using Michaelis–Menten plot & Lineweaver– Burk plot analysis. The type of α -glucosidase inhibition shown by **1j** was non-competitive (Figure 7). In non-competitive type of reactions, Michaelli's-Menten constant (Km) remains same whereas maximum velocity (Vmax) of the enzymatic reaction decreases. In this type of reaction, inhibitor reduces the activity of the enzyme irrespective of substrate binding. The Km and Vmax of α -glucosidase with **1j** inhibitor were depicted as 71.42 μ M⁻¹ and 0.02 μ M/min and without inhibitor (control) as 71.42 μ M⁻¹ and 0.04 μ M/min respectively. Whereas the acarbose, a standard anti-diabetic drug is reported to show a competitive type inhibition of α -glucosidase.¹⁹⁶ However, for the management of type-2 diabetes, use of drugs which shows non-competitive type inhibition of α -glucosidase enzyme should be given preference over the drugs that depict competitive type inhibition (e.g. acarbose). Competitive type inhibition depends on the substrate concentration and α -glucosidase inhibition potential of these drugs can be overcome by increasing the concentration of substrate. So, if a type-2 diabetes patient is given acarbose tablets and is having excess food with more carbohydrates,

CHAPTER 3

then the acarbose effect on enzyme would be overcome by higher concentration of carbohydrates whereas if a patient is taking a drug which shows non-competitive type of inhibition and simultaneously having higher carbohydrate containing diet, the carbohydrates would have no effect on the enzyme inhibition potential of the drug.



Figure 7: Lineweaver–Burk plots of α -glucosidase inhibition at different concentrations of substrate and 2-(benzo[*d*][1,3]dioxol-5-yl)-4*H*-chromen-4-one **1***j*.

Statistical analysis

Statistical analysis for α -glucosidase assay was carried out using graph pad prism-5 software. All samples were evaluated in triplicates and standard deviation was calculated. Sample data were analyzed with student's t-test and one-way ANOVA with tukey's test that was performed using graph pad prism version 5.00 for windows, graph pad software, San Diego California USA (www.graphpad.com).

α - glucosidase inhibition assay

The α -glucosidase inhibition was determined spectrophotometrically using *p*-nitrophenyl- α -D-glucopyranoside (*p*NPG) as substrate.¹⁹⁷ α -Glucosidase, 7.5 μ L (0.5 U/mL) was mixed with various concentrations of **1j**. After incubation of this mixture at 37 °C for 30 min, 100 μ L of *p*NPG (3 mM) was added. Reaction mixture was then again incubated for 10 min at 37 °C. To stop the reaction, 750 μ L of Na₂CO₃ (0.1 M) was added and absorbance was determined at 405 nm in triplicates. Standard anti-diabetic drug acarbose (PHR1253, Fluka)

was used as a positive control for α -glucosidase inhibition assay. The percentage inhibition of the enzyme activity was calculated by using the following formula.

Inhibition (%) = Absorbance of test control - Absorbance of sample/ Absorbance of control $\times 100$.

Kinetic analysis of α -glucosidase inhibition

For kinetic studies, α -glucosidase enzyme 7.5 µL (0.5 U/mL) mixed with **1j** (12.5 µg/mL) and incubated for 15 min. Further, *p*NPG was added at different concentrations (0.0109-0.612 mM) to individual reactions and the absorbance was recorded at 405 nm with the time interval of 3 sec up to 180 sec. The Km and Vmax values were determined from the Michaelis-Menten equation and mode of inhibition by **1j** are represented graphically using Lineweaver–Burk plot.

Molecular docking studies of 1j

Allosteric and competitive binding mode can be possible with more than one pocket present in protein. Acarbose competes with substrate for active sites of α -glucosidase enzyme. Acarbose binding to active site of α -glucosidase was determined using Autodock 4.2 tool. Total 10 best docking models were obtained, out of them the best fit docked model was chosen to reveal the molecular interaction between acarbose and α -glucosidase with the minimum binding energy of (Δ G) of -6.04 and minimum inhibition constant (Ki) of 37.38 μ M. The structural model of the complex between acarbose and α -glucosidase and its 3D representation is depicted in figures 8A and 8B respectively. The 2D representation of the interaction between acarbose and α -glucosidase enzyme was analyzed by LIG-PLOT (Figure 8C). Acarbose depicted hydrogen bonding with His 98, His 207 amino acids of active sites of α -glucosidase. Other amino acids shows the hydrophobic and Pi-Pi interaction with the acarbose (Figure 8C).

In contrary to the acarbose, flavone **1j** depicted allosteric interaction with α -glucosidase (Figure 9A). Binding pocket on α -glucosidase for **1j** was different from that of the acarbose binding site (Figure 9B), so these results support the results obtained in enzyme kinetic study that enzyme inhibition behaviour of **1j** is non-competitive. Out of total 10 docked model obtained, the best fit model depicted the allosteric interaction of **1j** with α -glucosidase with minimum binding energy of -0.6.39 and minimum inhibition constant (Ki) of 20.76 µM. It was observed that amino acid Lys373 was involved in hydrogen bonding with **1j** inhibitor (Figure 9C). The other surrounding amino acids depicted hydrophobic and Pi-Pi interaction with **1j** inhibitor.



Figure 8: (A) Structural model of the complex between acarbose and α-glucosidase; (B) 3DRepresentation of the interaction between acarbose and α-glucosidase in the predicted binding site; (C) 2D Representation of the interactions between α-glucosidase and acarbose.



Figure 9: (A) Structural model of the complex between **1j** and α -glucosidase; (B) 3D Representation of the interaction between **1j** and α -glucosidase in the predicted binding site;

(C) 2D Representation of the interactions between α -glucosidase and 1j.

The mode of α -glucosidase inhibition shown by **1j** is a non-competitive type and is different from acarbose which shows competitive type of inhibition. Since, non-competitive inhibition of α -glucosidase exhibited by any drug is independent on substrate (carbohydrate) concentration, during the clinical trials, drugs having non-competitive type of α -glucosidase inhibition are more preferable over competitive type inhibiting drugs. Hence, at this stage, **1j** has shown a significant anti-diabetic potential which is more advantageous than acarbose, so in future, along with the other lead compounds, **1j** can also be a potent future lead compound for the management of type-2 diabetes.

Molecular docking

General remarks: PDB structure of α -glucosidase was downloaded from Pubchem (PDB ID: 3A4A). Acarbose a standard molecule for docking studies was downloaded from https://pubchem.ncbi.nlm.nih.gov/. The 3D structure of the **1j** was drawn and validated using Marvin sketch (https://www.chemaxon.com /products/marvin/marvinsketch). Windows based automated docking tool Autodock 4.2 was used for docking study. Discovery studio (http://accelrys.com/products/collaborative-science/biovia-discovery-studio/) and LIGPLOT v.5.4.3 (http://www.ebi.ac.uk/thornton-srv/software/LIGPLOT/) were used to analyze the molecular interactions between docked molecules.

Using the Autodock 4.2 tools, all water molecules of α -glucosidase were removed and essential hydrogen atoms and Gasteiger charges were assigned. Grid parameter file (GPF) was prepared with the grid spacing of 1 and ligand dimension $88 \times 87 \times 126$ for acarbose and 0.819 and ligand dimension $98 \times 88 \times 104$ for **1j**. Docking parameter file (DPF) was prepared using Lamarckian Genetic Algorithm (LGA) with parameters set to 10 runs, whereas energy evaluation was set to 2,500,000 and 27,000 generation. Autogrid4 and autodock4 were run to calculate the lowest energy conformation between ligand and target. Different energy terms including intermolecular energy (vdm + hbond + desolv energy + electrostatic energy), internal energy, torsional energy and binding energy can be obtained from the output DLG (Docking Log file) format which was further analysed using as PyMOL and LIG-PLOT.

3.6: Conclusion

Synthesis of flavones is established from 2'-hydroxyacetophenones and aromatic aldehydes in one pot using pyrrolidine and iodine catalysts in DMSO solvent at 150 °C in 60-88 %. The methodology involves domino aldol-Michael-oxidation reaction sequence catalyzed by pyrrolidine and iodine as base and oxidant respectively.

Also, this method avoids the step of isolation of chalcone or flavanone intermediates and then subjecting them to further oxidation as in case of most of the reported methods for flavones. Thus providing a straightforward route to flavones.

Several advantages of this methodology including inexpensive catalysts, broad substrate scope, lack of metal catalysts and products in high yields with no side reactions makes it a better synthetic approach to flavones.

The reaction protocol was also successfully scaled up to 5 g of starting aryl aldehyde **17a** to get consistent yield of the desired flavone **1a**.

The selected synthesized flavones were screened for their *in vitro* anti-diabetic activity with acarbose as standard, among which flavone **1j** exhibited highest activity.

Significant α -glucosidase inhibitions were performed at different doses of **1j** showing very high % inhibition of 99.3 ± 0.26 % at very low concentration of 7.6 µg/mL.

The inhibition kinetics analyzed by Lineweaver-Burk and Michaelis–Menten plot analysis indicated non-competitive type of inhibition of α -glucosidase enzyme by **1j** which is different from the standard acarbose which shows competitive type of inhibition.

3.7: Experimental



3.7.1: A procedure for the synthesis of 2-(3,4-dimethoxyphenyl)chroman-4-one 1a': 2'-Hydroxyacetophenone 16a (0.1 g, 0.7 mmol) and 3,4-dimethoxybenzaldehyde 17a (0.12 g, 0.7 mmol) were mixed together along with pyrrolidine (0.026 g, 0.35 mmol) in DMSO solvent (10 mL). The resulting mixture was then refluxed for 15 minutes. After completion of reaction (monitored by TLC) the reaction mass was allowed to cool and diluted with ethyl acetate (20 mL). The resulting solution was then washed with water (5-6 times, 10 mL each). Combined water layers were extracted with ethyl acetate (3-4 times, 10 mL each) and the two ethyl acetate extracts were combined which was washed with dilute HCl solution. It was followed by water washing, drying over anhydrous sodium sulphate and concentrating under reduced pressure to furnish the crude product. The residue obtained was purified by column chromatography using benzene as an eluent to afford flavanone 1a'.



3.7.2: A general procedure for the synthesis of flavones 1a-r: Substituted 2'hydroxyacetophenone 16 (0.7 mmol) and substituted aromatic/heteroaromatic aldehyde 17 (0.7 mmol) were mixed together along with pyrrolidine (0.35 mmol) and iodine (0.035 mmol) in DMSO solvent (10 mL). The resulting mixture was then heated at 150 °C for the given time. After completion of reaction (monitored by TLC) the reaction mass was allowed to cool and diluted with ethyl acetate (20 mL). The resulting solution was then washed with water (5-

6 times, 10 mL each). Combined water layers were extracted with ethyl acetate (3-4 times, 10 mL each) and the two ethyl acetate extracts were combined which was washed with saturated sodium thiosulphate solution. It was followed by drying over anhydrous sodium sulphate and concentrating under reduced pressure to furnish the crude product. The residue obtained was purified by column chromatography using petroleum ether-ethyl acetate as an eluent to afford flavones **1a-r**.



3.7.3: A procedure for the synthesis of flavone 1a: 2'-Hydroxyacetophenone 16a (0.1 g, 0.7 mmol) and 3,4-dimethoxybenzaldehyde **17a** (0.12 g, 0.7 mmol) were mixed together along with pyrrolidine (0.025 g, 0.35 mmol) and iodine (0.0045 g, 0.035 mmol) in DMSO solvent (10 mL). The resulting mixture was then heated at 150 °C for 10 h. After completion of reaction (monitored by TLC) the reaction mass was allowed to cool and diluted with ethyl acetate (20 mL). The resulting solution was then washed with water (5-6 times, 10 mL each). Combined water layers were extracted with ethyl acetate (3-4 times, 10 mL each) and the two ethyl acetate extracts were combined which was washed with saturated sodium thiosulphate solution. It was followed by drying over anhydrous sodium sulphate and concentrating under reduced pressure to furnish the crude product. The residue obtained was purified by column chromatography using petroleum ether-ethyl acetate (6:4) as an eluent to afford flavone **1a** (0.182 g, 88 %).



3.7.4: A procedure for the synthesis of 1-(2-hydroxyphenyl)-3-(pyridin-2-yl)propan-1one 22, 2-(pyridin-2-yl)chroman-4-one 1s' and 2-(pyridin-2-yl)-4*H*-chromen-4-one 1s: 2'-Hydroxyacetophenone 16a (0.2 g, 1.5 mmol) and 2-pyridinecarboxaldehyde 17n (0.16 g, 1.5 mmol) were mixed together along with pyrrolidine (0.052 g, 0.35 mmol) and iodine (0.019 g, 0.035 mmol) in DMSO solvent (10 mL). The resulting mixture was then heated at 150 °C for 2 h. After completion of reaction (monitored by TLC) the reaction mass was allowed to cool and diluted with ethyl acetate (20 mL). The resulting solution was then washed with water (5-6 times, 10 mL each). Combined water layers were extracted with ethyl acetate (3-4 times, 10 mL each) and the two ethyl acetate extracts were combined which was washed with saturated sodium thiosulphate solution. It was then washed with 2N NaOH solution (2 times, 10 mL each) and then with water. The ethyl acetate layer was dried over anhydrous sodium sulphate and concentrated under reduced pressure to furnish the crude product. The residue obtained was purified by column chromatography to afford 2-(pyridin-2-yl)chroman-4-one **1s'** and 2-(pyridin-2-yl)-4*H*-chromen-4-one **1s** using petroleum ether-ethyl acetate (8:2) and (7.5:2.5) as an eluent respectively.

The NaOH layer was neutralized by adding conc. HCl solution and then extracted with ethyl acetate (3-4 times, 10 mL each). It was then dried over anhydrous sodium sulphate and concentrated under reduced pressure to furnish the crude product. The residue obtained was purified by column chromatography using petroleum ether-ethyl acetate (1:1) as an eluent to afford 1-(2-hydroxyphenyl)-3-(pyridin-2-yl)propan-1-one **22**.

3.8 References

- a) Middleton, E. Jr. Adv Exp Med Biol. 1998, 439, 175. b) Aherne, S. A.; O'Brien, N. M. Nutrition 2002, 18, 75. c) Havsteen B. Biochem. Pharmacol. 1983, 32, 1141.
- The Flavonoids, advances in research since 1986; Harborne, J. B. Ed.; Chapman and Hall: London, 1993.
- a) Medina, J. H.; Paladini, A. C.; Wolfman, C.; Levi de Stein, M.; Calvo, D.; Diaz, L. E.; Peña, C. *Biochem. Pharmacol.* **1990**, *40*, 2227. b) Seetharaman, P.; Gnanasekar, S.; Chandrasekaran, R.; Chandrakasan, G.; Kadarkarai, M.; Sivaperumal, S. *Ann. Microbiol.* **2017**, *67*, 321. c) Anandhi, R.; Annadurai, T.; Anitha, T. S.; Muralidharan, A. R.; Najmunnisha, K.; Nachiappan, V.; Thomas, P. A.; Geraldine, P. *J. Physiol. Biochem.* **2013**, *69*, 313.
- a) Ishikura, N. Agric. Biol. Chem. 1981, 45, 1855. b) McKay, D. L.; Blumberg, J. B. Phytother. Res. 2006, 20, 519.
- a) Kuroda, M.; Iwabuchi, K.; Mimaki, Y. *Nat. Prod. Commun.* 2012, *7*, 471. b) Lee, D.; Cuendet, M.; Vigo, J. S.; Graham, J. G.; Cabieses, F.; Fong, H. H. S.; Pezzuto, J. M.; Kinghorn, A. D. *Org. Lett.* 2001, *3*, 2169.
- 6. Li, S.; Yu, H.; Ho, C-T. Biomed. Chromatogr. 2006, 20, 133.
- a) Umikalsom, Y.; Harborne, J. B. *Pertanika* 1991, *14*, 297. b) Sun, C.; Wang, H.; Wang, Y.;
 Xiao, S. *Molecules* 2016, *21*, 1067.

- a) Min, S. W.; Kim, N. J.; Baek, N. I.; Kim, D. H. *J. Ethnopharmacol.* 2009, *125*, 497. b) Lee,
 H. G.; Yu, K. A.; Oh, W. K.; Baeg, T. W.; Oh, H. C.; Ahn, J. S.; Jang, W. C.; Kim, J. W.;
 Lim, J. S.; Choe, Y. K.; Yoon, D. Y. *J. Ethnopharmacol.* 2005, *98*, 339.
- Kim, D-H.; Na, H-K.; Oh, T. Y.; Kim, W-B.; Surh, Y-J. Biochem. Pharmacol. 2004, 68, 1081.
- a) Lin, Y. L.; Lin, R. J.; Shen, K. P.; Dai, Z. K.; Chen, I. J.; Wu, J. R.; Wu, B. N. J. *Ethnopharmacol.* 2011, *138*, 373. b) Eiden, M.; Leidel, F.; Strohmeier, B.; Fast, C.; Groschup, M. H. *Front. Psychiatry* 2012, *3*, 9. c) Chen, L. J.; Games, D. E.; Jones, J. J. *Chromatogr. A.* 2003, 988, 95.
- Hui, K. M.; Huen, M. S. Y.; Wang, H. Y.; Zheng, H.; Sigel, E.; Baur, R.; Ren, H.; Li, Z. W.; Wong, J. T-F.; Xue, H. *Biochem. Pharmacol.* 2002, 64, 1415.
- 12. Barberan, F. A. T.; Gil, M. I.; Tom, F.; Ferreres, F. J. Nat. Prod. 1985, 48, 859.
- Guimarães, C. C.; Oliveira, D. D.; Valdevite, M.; Saltoratto, A. L. F.; Pereira, S. I. V.; França, S. De C.; Pereira, A. M. S.; Pereira, P. S. *Food Chem. Toxicol.* 2015, *86*, 88.
- a) Sezik, E.; Aslan, M.; Yesilada, E.; Ito, S. *Life Sci.* 2005, *76*, 1223. b) Anilkumar, K.;
 Reddy, G. V.; Azad, R.; Yarla, N. S.; Dharmapuri, G.; Srivastava, A.; Kamal, M. A.; Pallu, R. *Oxid. Med. Cell. Longev.* 2017, *2017*, 1.
- 15. Segelman, A. B.; Segelman, F. P.; Star, A. E.; Wagner, H.; Seligmann, O. *Phytochemistry* **1978**, *17*, 824.
- a) Flamini, G.; Pardini, M.; Morelli, I.; Ertugrul, K.; Dural, H.; Bagci, Y.; Kargioglu, M. Phytochemistry 2002, 61, 433. b) Eshbakova, K. A.; Toshmatov, Z. O.; Yili, A.; Aisa, H. A.; Abdullaev, N. D. Chem. Nat. Comp. 2013, 49, 103.
- Peng-fei, L.; Fu-gen, H.; Bin-bin, D.; Tain-sheng, D.; Xiang-lin, H.; Ming-qin, Z. J. Food Sci. Technol. 2013, 50, 615.
- a) Ramachandran Nair, A. G.; Joshi, B. S. Proc. Indian Acad. Sci. 1979, 88, 323. b)
 Chaudhuri, P. K.; Srivastava, R.; Kumar, S.; Kumar, S. Phytother. Res. 2004, 18, 114.
- 19. Veitch, N. C.; Grayer, R. J.; Irwin, J. L.; Takeda, K. Phytochemistry 1998, 48, 389.
- a) Cushnie, T. P.T.; Lamb, A. J. Int. J. Antimicrob. Agents 2005, 26, 343. b) Middleton, Jr. E.; Kandaswami, C.; Theoharides, T. C. Pharmacol. Rev. 2000, 52, 673. c) Martens, S.; Mithöfer, A. Phytochemistry 2005, 66, 2399.
- Comalada, M.; Ballester, I.; Bailón, E.; Sierra, S.; Xaus, J.; Gálvez, J.; Sánchez de Medina, F.; Zarzuelo, A. *Biochem. Pharmacol.* 2006, 72, 1010.
- 22. Kun, Q.; Zheng-Ru, K.; Jie, Z.; Xiao-Wei, C.; Zong-Yi, W.; Cheng-Xi, G.; Hong-Xia, S.; Ai-Jian, Q. *Virus Res.* **2018**, *248*, 63.
- Resende, F. A.; de Oliveira, A. P. S.; de Camargo, M. S.; Vilegas, W.; Varanda, E. A. *PLoS* ONE 2013, 8, e74881.

- 24. a) Shukla, S.; Gupta, S. *Pharm. Res.* 2010, 27, 962. b) Lin, Y.; Shi, R.; Wang, X.; Shen, H-M. *Curr. Cancer Drug Targets* 2008, 8, 634. c) Khoo, B. Y.; Chua, S. L.; Balaram, P. *Int. J. Mol. Sci.* 2010, *11*, 2188. d) Gupta, S.; Afag, F.; Mukhtar, H. *Biochem. Biophys. Res. Commun.* 2001, 287, 914.
- 25. Pietta, P-G. J. Nat. Prod. 2000, 63, 1035.
- 26. a) Mittra, B.; Saha, A.; Chowdhury, A. R.; Pal, C.; Mandal, S.; Mukhopadhyay, S.; Bandyopadhyay, S.; Majumder, H. K. *Mol. Med.* 2000, *6*, 527. b) Beer, M. F.; Frank, F. M.; Elso, O. G.; Bivona, A. E.; Cerny, N.; Giberti, G.; Malchiodi, E. L.; Martino, V. S.; Alonso, M. R.; Sülsen, V. P.; Cazorla, S. I. *Pharm. Biol.* 2016, *54*, 2188. c) Salem, M. M.; Capers, J.; Rito, S.; Werbovetz, K. A. *Phytother. Res.* 2011, *25*, 1246.
- 27. a) Diniz, T. C.; Silva, J. C.; de Lima-Saraiva, S. R. G.; de Almeida Ribeiro, F. P. R.; Pacheco, A. G. M.; de Freitas, R. M.; Quintans-Júnior, L. J.; de Souza Siqueira Quintans, J.; Mendes, R. L.; da Silva Almeida, J. R. G. *Oxid. Med. Cell Longev.* 2015, 2015, 171756. b) Park, H. G.; Yoon, S. Y.; Choi, J. Y.; Lee, G. S.; Choi, J. H.; Shin, C. Y.; Son, K. H.; Lee, Y. S.; Kim, W. K.; Ryu, J. H.; Ko, K. H.; Cheong, J. H. *Eur. J. Pharmacol.* 2007, 574, 112.
- 28. Kuwabara, H.; Mouri, K.; Otsuka, H.; Kasai, R.; Yamasaki, K. J. Nat. Prod. 2003, 66, 1273.
- 29. Harborne, J. B. Nat. Prod. Rep. 1999, 16, 509.
- a) Li, B. Q.; Fu, T.; Yan, Y. D.; Baylor, N. W.; Ruscetti, F. W.; Kung, H. F. *Cell Mol. Biol. Res.* **1993**, *39*, 119. b) Hu, C-Q.; Chen, K.; Shi, Q.; Kilkuskie, R. E.; Cheng, Y-C.; Lee, K-H. *J. Nat. Prod.* **1994**, *57*, 42. c) Critchfield, J. W.; Butera, S. T.; Folks, T. M. *AIDS Res. Hum. Retroviruses* **1996**, *12*, 39. d) Li, B. Q.; Fu, T.; Dongyan, Y.; Mikovits, J. A.; Ruscetti, F. W.; Wang, J. M. *Biochem. Biophys. Res. Commun.* **2000**, *276*, 534.
- 31. a) Ajay, M.; Gilani, A. H.; Mustafa, M. R. *Life Sci.* 2003, 74, 603. b) Ferreira, L. L. D. M.; Gomes, M. V.; Paes, B. M.; do Carmo, P. L.; Konno, T. U. P.; de Assis Esteves, F.; Lopes, N. P.; Tomaz, J. C.; Correa, I.; Leal, R.; Guimarães, D. O.; Muzitano, M. F.; Raimundo, J. M. *Planta Med.* 2017, 83, 63.
- a) Ragone, M. I.; Sella, M.; Conforti, P.; Volonté, M. G.; Consolini, A. E. *J Ethnopharmacol.* **2007**, *113*, 258. b) Prabhakar, M. C.; Bano, H.; Kumar, I.; Shamsi, M. A.; Khan, M. S. Y. *Planta Med.* **1981**, *43*, 396.
- 33. a) Torres-Piedra, M.; Ortiz-Andrade, R.; Villalobos-Molina, R.; Singh, N.; Medina-Franco, J. L.; Webster, S. P.; Binnie, M.; Navarrete-Vázquez, G.; Estrada-Soto, S. *Eur. J. Med. Chem.*2010, 45, 2606. b) Kato, A.; Nasu, N.; Takebayashi, K.; Adachi, I.; Minami, Y.; Sanae, F.; Asano, N.; Watson, A. A.; Nash, R. J. *J. Agric. Food Chem.* 2008, 56, 4469.
- 34. a) Kanazawa, K.; Yamashita, T.; Ashida, H.; Danno, G. Biosci. Biotechnol. Biochem. 1998, 62, 970. b) Miyazawa, M.; Hisama, M. Biosci. Biotechnol. Biochem. 2003, 67, 2091. c) Gulluce, M.; Orhan, F.; Yanmis, D.; Arasoglu, T.; Guvenalp, Z.; Demirezer, L. O. Toxicol. Ind. Health 2015, 31, 831.

- 35. a) Cheong, H.; Ryu, S-Y.; Oak, M-H.; Cheon, S-H.; Yoo, G-S.; Kim, K-M. Arch. Pharm. Res. 1998, 21, 478. b) Hirano, T.; Higa, S.; Arimitsu, J.; Naka, T.; Ogata, A.; Shima, Y.; Fujimoto, M.; Yamadori, T.; Ohkawara, T.; Kuwabara, Y.; Kawai, M.; Matsuda, H.; Yoshikawa, M.; Maezaki, N.; Tanaka, T.; Kawase, I.; Tanaka, T. Biochem. Biophys. Res. Commun. 2006, 340, 1. c) Tanaka, T.; Higa, S.; Hirano, T.; Kotani, M.; Matsumoto, M.; Fujita, A.; Kawase. I. Curr. Med. Chem.-Anti-Inflammatory Anti-Allergy Agents 2003, 2, 57. d) Yano, S.; Tachibana, H.; Yamada, K. J. Agric. Food Chem. 2005, 53, 1812. e) Hirano, T.; Higa, S.; Arimitsu, J.; Naka, T.; Shima, Y.; Ohshima, S.; Fujimoto, M.; Yamadori, T.; Kawase, I.; Tanaka, T. Int. Arch. Allergy Immunol. 2004, 134, 135.
- 36. a) Austin, C. A.; Patel, S.; Ono, K.; Nakane, H.; Fisher, L. M. *Biochem. J.* 1992, 282, 883. b)
 Ahmad, M. S.; Fazal, F.; Rahman, A.; Hadi, S. M.; Parish, J. H. *Carcinogenesis* 1992, 13, 605.
- Anand, K. V.; Jaabir, M. S. M.; Thomas, P. A.; Geraldine, P. Geriatr. Gerontol. Int. 2012, 12, 741.
- 38. a) Nakazawa, T.; Yasuda, T.; Ueda, J.; Ohsawa, K. *Biol. Pharm. Bull.* 2003, 26, 474. b) Yi, L.
 T.; Li, J. M.; Li, Y. C.; Pan, Y.; Xu, Q.; Kong, L. D. *Life Sci.* 2008, 82, 741.
- Han, X. H.; Hong, S. S.; Hwang, J. S.; Lee, M. K.; Hwang, B. Y.; Ro, J. S. Arch. Pharm. Res. 2007, 30, 13.
- 40. a) Morimoto, M.; Tanimoto, K.; Nakano, S.; Ozaki, T.; Nakano, A.; Komai, K. J. Agric. Food Chem. 2003, 51, 389. b) Duchowicz, P. R.; Goodarzi, M.; Ocsachoque, M. A.; Romanelli, G. P.; Ortiz, E. del V.; Autino, J. C.; Bennardi, D. O.; Ruiz, D. M.; Castro, E. A. Sci. Total Environ. 2009, 408, 277.
- 41. a) Baker, W. J. Chem. Soc. 1933, 1381. b) Mahal, H. S.; Venkataraman, K. J. Chem. Soc. 1934, 1767. c) Mahal, H. S.; Venkataraman, K. Curr. Sci. 1933, 4, 214. d) Wheeler, T. S. Org. Synth. 1952, 32, 72. e) Wheeler, T. S. Org. Synth. 1963, 4, 478. For recent examples: f) Riva, C.; De Toma, C.; Donadd, L.; Boi, C.; Pennini, R.; Motta, G.; Leonardi, A. Synthesis 1997, 1997, 195. g) Bois, F.; Beney, C.; Mariotte, A. M.; Boumendjel, A. Synlett 1999, 1999, 1480. h) Ganguly, A. K.; Kaur, S.; Mahata, P. K.; Biswas, D.; Pramanik, B. N.; Chan, T. M. Tetrahedron Lett. 2005, 46, 4119. i) Ganguly, A. K.; Mahata, P. K.; Biswas, D. Tetrahedron Lett. 2015, 46, 4119. i) Chee, C. F.; Buckle, M. J. C.; Rahman, N. A. Tetrahedron Lett. 2011, 52, 3120.
- 42. a) Allan, J.; Robinson, R. J. Chem. Soc. 1924, 125, 2192. b) Fukui, K.; Matsumoto, T.; Nakamura, S.; Nakayam, M. Bull. Chem. Soc. Jpn. 1968, 41, 1413. c) Fukui, K.; Nakayama, M.; Horie, T. Bull. Chem. Soc. Jpn. 1969, 42, 1649. d) Fukui, K.; Matsumoto, T.; Nakayama, M.; Horie, T. Experientia 1969, 25, 349. e) Kaneta, M.; Hikichi, H.; Endo, S.; Sugiyama, N. Bull. Chem. Soc. Jpn. 1978, 51, 1784.

- 43. a) Verma, A. K.; Pratap. R. *Tetrahedron* 2012, *68*, 8523. b) Wang, Z.; Yang, L.; Yang, X.; Zhang, X. *Synth. Commun.* 2013, *43*, 3093. c) Kshatriya, R. B.; Shaikh, Y. I.; Nazeruddin, G. M. *Orient. J. Chem.* 2013, *29*, 1475. d) Das, M.; Manna, K.; Banik, U.; Ghosh, P. S.; Sarkar, P. *Int. J. Pharm. Sci. Res.* 2014, *5*, 3840. e) Tawfik, H. A.; Ewies, E. F.; El-Hamouly, W. S. *Int. J. Res. Pharm. Chem.* 2014, *4*, 1046. f) Masesane, I. B. *Int. J. Chem. Stud.* 2015, *3*, 53. g) Bhatt, D.; Soni, R.; Sharma, G. K.; Dashora, A. *Indo Am. J. Pharm. Res.* 2016, *6*, 4345.
- 44. Liu, R.; Wang, X.; Cheng, F.; Li, F.; Xu, K.; Tan, G. Chin. J. Org. Chem. 2016, 36, 2677.
- 45. Liu, R.; Zhang, Y.; Xu, K.; Tan, G. Synth. Commun. 2017, 47, 1.
- 46. Lahyani, A.; Trabelsi, M. Ultrason. Sonochem. 2016, 31, 626.
- 47. Lamba, M.; Makrandi, J. K. J. Chem. Res. 2008, 2008, 225.
- 48. Kumar, S.; Sharma, D. Orient. J. Chem. 2011, 27, 761.
- 49. a) Huang, X.; Tang, E.; Xu, W-M.; Cao, J. J. Comb. Chem. 2005, 7, 802. b) Cao, J.; Tang,
 E.; Huang, X.; WU, L. L.; Huang, X. Chin. Chem. Lett. 2006, 17, 857.
- 50. a) Mahal, H. S.; Rai, H. S.; Venkataraman, K. J. Chem. Soc. 1935, 866. b) Makrandi, J. K.; Seema Chem. Ind. 1989, 18, 607.
- 51. Gupta, M.; Paul, S.; Gupta, R.; Loupy, A. Org. Prep. Proced. Int. 2000, 32, 280.
- 52. Ganguly, N. C.; Chandra, S.; Barik, S. K. Synth. Commun. 2013, 43, 1351.
- 53. Hoshino, Y.; Oohinata, T.; Takeno, N. Bull. Chem. Soc. Jpn. 1986, 59, 2351.
- 54. Kumar, K. H.; Perumal, P. T. Tetrahedron 2007, 63, 9531.
- 55. Jakhar, K.; Makrandi, J. K. Indian J. Chem. 2013, 52B, 141.
- 56. Mallik, U. K.; Saha, M. M.; Mallik, A. K. Indian J. Chem. 1989, 28B, 970.
- 57. Gulácsi, K.; Litkei, G.; Antus, S.; Gunda, T. E. Tetrahedron 1998, 54, 13867.
- 58. Parveen, A. Int. J. Curr. Microbiol. App. Sci. 2013, 2, 296.
- 59. Kasahara, A.; Izumi, T.; Ooshima, M. Bull. Chem. Soc. Jpn. 1974, 47, 2526.
- 60. Du, Z.; Ng, H.; Zhang, K.; Zeng, H.; Wang, J. Org. Biomol. Chem. 2011, 9, 6930.
- 61. Chen, J-F.; Liu, Z-Q. Chem. Res. Toxicol. 2015, 28, 451.
- 62. Kulkarni, P. S.; Kondhare, D. D.; Varala, R.; Zubaidha, P. K. J. Serb. Chem. Soc. 2013, 78, 909.
- 63. Zambare, A. S.; Sangshetti, J. N.; Kokare, N. D.; Shinde, D. B. Chin. Chem. Lett. 2009, 20, 171.
- 64. Ahmed, N.; Ali, H.; Van Lier, J. E. Tetrahedron Lett. 2005, 46, 253.
- 65. Babu, K. R.; Kumar, K. V.; Vijaya, M.; Madhavarao, V. Int. J. Pharm. Technol. 2012, 4, 3943.
- 66. Lorenz, M.; Kabir, M. S.; Cook, J. M. Tetrahedron Lett. 2010, 51, 1095.
- a) Doshi, A. G.; Soni, P. A.; Ghiya, B. J. *Indian J. Chem.* **1986**, *25B*, 759. b) Agrawal, N. N.;
 Soni, P. A. *Indian J. Chem.* **2005**, *44B*, 2601.
- 68. Miyake, H.; Takizawa, E.; Sasaki, M. Bull. Chem. Soc. Jpn. 2003, 76, 835.

- 69. Sashidhara, K. V.; Kumar, M.; Kumar, A. Tetrahedron Lett. 2012, 53, 2355.
- 70. Sarda, S. R.; Jadhav, W. N.; Pawar, R. P. Int. J. ChemTech Res. 2009, 1, 539.
- 71. a) Menezes, M. J.; Manjrekar, S.; Pai, V.; Patre, R. E.; Tilve, S. G. Indian J. Chem. 2009, 48B, 1311. b) Borse, S. L.; Patel, M. R.; Borse, L. B. Int. J. Pharm. Res. Dev. 2011, 3, 147. c) Belsare, D. P.; Kazi, A. IOSR J. Pharm. 2013, 3, 23.
- 72. Bose, G.; Mondal, E.; Khan, A. T.; Bordoloi, M. J. Tetrahedron Lett. 2001, 42, 8907.
- 73. Lokhande, P. D.; Sakate, S. S.; Taksande, K. N.; Navghare, B. *Tetrahedron Lett.* **2005**, *46*, 1573.
- 74. Nawghare, B. R.; Gaikwad, S. V.; Raheem, A.; Lokhande, P. D. J. Chil. Chem. Soc. 2014, 59, 2284.
- 75. Sarda, S. R.; Pathan, M. Y.; Paike, V. V.; Pachmase, P. R.; Jadhav, W. N.; Pawar, R. P. *Arkivoc* **2006**, (*xvi*), 43.
- 76. Bhosale, R. S.; Sarda, S. R.; Giram, R. P.; Raut, D. S.; Parwe, S. P.; Ardhapure, S. S.; Pawar, R. P. J. Iran. Chem. Soc. 2009, 6, 519.
- 77. a) Nagarathnam, D.; Cushman, M. J. Org. Chem. 1991, 56, 4884. b) Banerji, A.; Goomer, N. C. Synthesis 1980, 1980, 874.
- 78. Lee, J. I.; Son, H. S.; Park, H. Bull. Korean Chem. Soc. 2004, 25, 1945.
- 79. Hirao, I.; Yamaguchi, M.; Hamada, M. Synthesis 1984, 1984, 1076.
- 80. Lee, J. I.; Son, H. S.; Jung, M. G. Bull. Korean Chem. Soc. 2005, 26, 1461.
- 81. Makrandi, J. K.; Kumari, V. Chem. Ind. 1988, 19, 630.
- 82. Sharma, D.; Makrandi, J. K. Green Chem. Lett. Rev. 2009, 2, 157.
- 83. Jakhar, K.; Makrandi, J. K. Indian J. Chem. 2012, 51B, 770.
- 84. Garg, S.; Ishar, M. P. S.; Sarin, R.; Gandhi, R. P. Indian J. Chem. 1994, 33B, 1123.
- 85. Varma, R. S.; Saini, R. K.; Kumar, D. J. Chem. Research (S) 1998, 348.
- 86. Thorat, N. M.; Dengale, R. A.; Thopate, S. R.; Rohokale, S. V. Lett. Org. Chem. 2015, 12, 574.
- Ramana, M. M. V.; Nimkar, Amey P.; Betkar, Rahul R.; Ranade, Prasanna B. Int. J. Pharm. Sci. Rev. Res. 2014, 25, 202.
- 88. Thorat, Nitin M.; Kote, Santosh R.; Thopate, Shankar R. Lett. Org. Chem. 2014, 11, 601.
- 89. Miyake, H.; Nishino, S.; Nishimura, A.; Sasaki, M. Chem. Lett. 2007, 36, 522.
- Vazquez, P.; Pizzio, L.; Romanelli, G.; Autino, J.; Caceres, C.; Blanco, M. Applied Catal. A: Gen. 2002, 235, 233.
- Bennardi, D. O.; Romanelli, G. P.; Autino, J. C.; Pizzio, L. R. Applied Catal. A: Gen. 2007, 324, 62.
- 92. Bennardi, D. O.; Romanelli, G. P.; Autino, J. C.; Pizzio, L. R. Catal. Commun. 2009, 10, 576.
- 93. Bennardi, D. O.; Romanelli, G. P.; Jios, J. L.; Autino, J. C.; Baronetti, G. T.; Thomas, H. J. *Arkivoc* **2008**, (*xi*), 123.

- 94. Bennardi, D. O.; Ruiz, D. M.; Romanelli, G. P.; Baronetti, G. T.; Thomas, H. J.; Autino, J. C. *Lett. Org. Chem.* 2008, 5, 607.
- 95. a) Gharib, A.; Jahangir, M.; Roshani, M.; Scheeren, J. W. *Bulg. Chem. Commun.* 2010, 42, 210. b) Romanelli, G. P.; Virla, E. G.; Duchowicz, P. R.; Gaddi, A. L.; Ruiz, D. M.; Bennardi, D. O.; Ortiz, E. D. V.; Autino, J. C. *J. Agric. Food Chem.* 2010, 58, 6290.
- 96. Bennardi, D.; Romanelli, G.; Autino, J.; Pizzio, L.; Vázquez, P.; Cáceres, C.; Blanco, M. *Reac. Kinet. Mech. Cat.* 2010, 100, 165.
- 97. Bennardi, D. O.; Romanelli, G. P.; Jios, J. L.; Vazquez, P. G.; Caceres, C. V.; Autino, J. C. *Heterocycl. Commun.* 2007, 13, 77.
- 98. Hoshino, Y.; Takeno, N. Bull. Chem. Soc. Jpn. 1987, 60, 1919.
- 99. Kucukislamoglu, M.; Nebioglu, M.; Zengin, M.; Arslan, M.; Yayli, N. J. Chem. Res. 2005, 2005, 556.
- 100. Zhu, X.; Li, Z.; Shu, Q.; Zhou, C.; Su, W. Synth. Commun. 2009, 39, 4199.
- 101. Pérez, M.; Ruiz, D.; Autino, J.; Sathicq, A.; Romanelli, G. C. R. Chimie 2016, 19, 551.
- 102. Zubaidha, P. K.; Hashmi, A. M.; Bhosale, R. S. Heterocycl. Commun. 2005, 11, 97.
- 103. Vimal, Manorama; Pathak, Uma; Mathur, Sweta; Pandey, Lokesh Kumar; Suryanarayana, M. V. S. *Heterocycl. Commun.* 2010, *16*, 151.
- 104. Pérez, M. E.; Ruiz, D. M.; Autino, J. C.; Blanco, M. N.; Pizzio, L. R.; Romanelli, G. P. J. *Porous Mater* **2013**, *20*, 1433.
- 105. Jin, C.; He, F.; Wu, H.; Chen, J.; Su, W. J. Chem. Res. 2009, 2009, 27.
- 106. Lee, Y. R.; Kang, K. Y. Lett. Org. Chem. 2007, 4, 440.
- 107. Dengale, R. A.; Thorat, N. M.; Thopate, S. R. Lett. Org. Chem. 2016, 13, 734.
- 108. Su, W. K.; Zhu, X. Y.; Li, Z. H. Org. Prep. Proced. Int. 2009, 41, 69.
- 109. Kabalka, G. W.; Mereddy, A. R. Tetrahedron Lett. 2005, 46, 6315.
- 110. Wen, S-S.; Wang, J.; Luo, Y-M.; Yang, H. Tetrahedron 2014, 70, 9314.
- 111. a) Nishinaga, A.; Maruyama, K.; Ando, H.; Sato, R.; Mashino, T.; Akira, I.; Tsutomu, N. *Tetrahedron Lett.* 1990, *31*, 3171. b) Nishinaga, A.; Ando, H.; Maruyama, K.; Mashino, T. *Synthesis* 1992, 1992, 839.
- 112. Stanek, F.; Stodulski, M. Tetrahedron Lett. 2016, 57, 3841.
- 113. Ismail, K. A.; Aziem, T. Abd. El Eur. J. Med. Chem. 2001, 36, 243.
- 114. Zhao, J.; Zhao, Y.; Fu, H. Angew. Chem. Int. Ed. 2011, 50, 3769.
- 115. Zhao, J.; Zhao, Y.; Fu, H. Org. Lett. 2012, 14, 2710.
- 116. Cavill G. W. K.; Dean F. M.; McGookin A.; Marshall B. M.; Robertson A. J. Chem. Soc. 1954, 4573.
- 117. a) Shanker, C. G.; Mallaiah, B. V.; Srimannarayana, G. Synthesis 1983, 1983, 310. b)
 Hoshino, Y.; Takeno, N. Bull. Chem. Soc. Jpn. 1987, 60, 4468.
- 118. Climent, M. J.; Garcia, H.; Iborra, S.; Miranda, M. A.; Primo, J. Heterocycles 1989, 29, 115.

- 119. Varma, R. S.; Varma, M. Synth. Commun. 1982, 12, 927.
- 120. a) Singh, O. V.; Kapoor, R. P. *Tetrahedron Lett.* 1990, *31*, 1459. b) Khanna, M. S.; Singh, O. V.; Garg, C. P.; Kapoor, R. P. *J. Chem. Soc. Perkin Trans.* 1 1992, 2565.
- 121. Singh, O. V.; Muthukrishnan, M.; Raj, G. Synth. Commun. 2005, 35, 2723.
- 122. Zhou, Z.; Zhao, P.; Huang, W.; Yang, G. Adv. Synth. Catal. 2006, 348, 63.
- 123. Wegner, J.; Ceylan, S.; Friese, C.; Kirschning, A. Eur. J. Org. Chem. 2010, 23, 4372.
- 124. Mal, K.; Kaur, A.; Haque, F.; Das, I. J. Org. Chem. 2015, 80, 6400.
- 125. Prakash, O.; Tanwar, M. P. J. Chem. Res. (S) 1995, 213.
- 126. Pande, G. B.; Shirodkar, S. G. Rasayan J. Chem. 2013, 6, 303.
- 127. Prakash, O.; Pahuja, S.; Moriarty, R. M. Synth. Commun. 1990, 20, 1417.
- 128. Muthukrishnan, M.; Patil, P. S.; More, S. V.; Joshi, R. A. Mendeleev Commun. 2005, 15, 100.
- 129. Barontini, M.; Bernini, R.; Crisante, F.; Fabrizi, G. Tetrahedron 2010, 66, 6047.
- Lokhande, P. D.; Dalvi, B. A.; Humne, V. T.; Nawghare, B. R.; Kareem, A. Indian J. Chem.
 2014, 53B,1091.
- 131. Bovicelli, P.; D'Angelo, V.; Collalto, D.; Verzina, A.; D'Antona, N.; Lambusta, D. J. Pharm. *Pharmacol.* **2007**, *59*, 1697.
- 132. Diao, T.; Stahl, S. S. J. Am. Chem. Soc. 2011, 133, 14566.
- 133. Diao, T.; Wadzinski, T. J.; Stahl, S. S. Chem. Sci. 2012, 3, 887.
- 134. Baruah, S.; Kaishap, P. P.; Gogoi, S. Chem. Commun. 2016, 52, 13004.
- 135. Liu, J.; Song, W.; Yue, Y.; Liu, R.; Yi, H.; Zhuo, K.; Lei, A. Chem. Commun. 2015, 51, 17576.
- 136. Maiti, G.; Karmakar, R.; Bhattacharya, R. N.; Kayal, U. Tetrahedron Lett. 2011, 52, 5610.
- 137. Yatabe, T.; Jin, X.; Yamaguchi, K.; Mizuno, N. Angew. Chem. Int. Ed. 2015, 54, 13302.
- 138. Zhu, Fengxiang; Li, Yahui; Wang, Zechao; Wu, Xiao-Feng Catal. Sci. Technol. 2016, 6, 2905.
- 139. Xue, L.; Shi, L.; Han, Y.; Xia, C.; Huynh, H. V.; Li, F. Dalton Trans. 2011, 40, 7632.
- 140. Miao, H.; Yang, Z. Org. Lett. 2000, 2, 1765.
- 141. Awuah, E.; Capretta, A. Org. Lett. 2009, 11, 3210.
- 142. Yang, Q.; Alper, H. J. Org. Chem. 2010, 75, 948.
- 143. Liang, B.; Huang, M.; You, Z.; Xiong, Z.; Lu, K.; Fathi, R.; Chen, J.; Yang, Z. J. Org. Chem. 2005, 70, 6097.
- 144. Liu, J.; Liu, M.; Yue, Y.; Zhang, N.; Zhang, Y.; Zhuo, K. Tetrahedron Lett. 2013, 54, 1802.
- 145. Zhu, F.; Wang, Z.; Li, Y.; Wu, X-F. Chem. Eur. J. 2017, 23, 3276.
- 146. Wu, X-F.; Neumann, H.; Beller, M. Chem. Eur. J. 2012, 18, 12595.
- 147. Khoobi, M.; Alipour, M.; Zarei, S.; Jafarpour, F.; Shafiee, A. Chem. Commun. 2012, 48, 2985.
- 148. Kim, D.; Ham, K.; Hong, S. Org. Biomol. Chem. 2012, 10, 7305.

- 149. Min, M.; Choe, H.; Hong, S. Asian J. Org. Chem. 2012, 1, 47.
- 150. Kim, K. H.; Lee, H. S.; Kim, S. H.; Kim, J. N. Tetrahedron Lett. 2012, 53, 2761.
- 151. Klier, L.; Bresser, T.; Nigst, T. A.; Karaghiosoff, K.; Knochel, P. J. Am. Chem. Soc. 2012, 134, 13584.
- 152. Muto, K.; Yamaguchi, J.; Musaev, D. G.; Itami, K. Nature Commun. 2015, 6, 7508.
- 153. Lee, J.; Yu, J.; Son, S. H.; Heo, J.; Kim, T.; An, J-Y.; Inn, K-S.; Kim, N-J. Org. Biomol. *Chem.* **2016**, *14*, 777.
- 154. Kraus, G. A.; Gupta, V. Org. Lett. 2010, 12, 5278.
- 155. Moon, Y.; Kwon, D.; Hong, S. Angew. Chem. Int. Ed. 2012, 51, 11333.
- 156. Jiang, N.; Li, S-Y.; Xie, S-S.; Yao, H.; Sun, H.; Wang, X-B.; Kong, L-Y. RSC Adv. 2014, 4, 63632.
- 157. Yakovenko, V. I.; Oganesyan, É. T.; Dorofeenko, G. N. Chem. Heterocycl. Compd. 1981, 17, 115.
- 158. Golub, A. G.; Bdzhola, V. G.; Ostrynska, O. V.; Kyshenia, I. V.; Sapelkin, V. M.; Prykhod'ko, A. O.; Kukharenko, O. P.; Yarmoluk, S. M. *Bioorg. Med. Chem.* **2013**, *21*, 6681.
- 159. Seijas, J. A.; Va'zquez-Tato, M. P.; Carballido-Reboredo, R. J. Org. Chem. 2005, 70, 2855.
- 160. Zhou, C.; Dubrovsky, A. V.; Larock, R. C. J. Org. Chem. 2006, 71, 1626.
- 161. Chuang, D-W.; El-Shazly, M.; Balaji D. B.; Chung, Y-M.; Chang, F-R.; Wu, Y-C. Eur. J. Org. Chem. 2012, 2012, 4533.
- 162. Yang, D.; Wang, Z.; Wang, X.; Sun, H.; Xie, Z.; Fan, J.; Zhang, G.; Zhang, W.; Gao, Z. J. Mol. Catal. A: Chem. 2017, 426, 24.
- 163. Yoshida, M.; Fujino, Y.; Saito, K.; Doi, T. Tetrahedron 2011, 67, 9993.
- 164. a) Yoshida, M.; Fujino, Y.; Doi, T. Org. Lett. 2011, 13, 4526. b) Taylor, C.; Bolshan, Y. Tetrahedron Lett. 2015, 56, 4392.
- 165. Zhang, S.; Wan, C.; Wang, Q.; Zhang, B.; Gao, L.; Zha, Z.; Wang, Z. Eur. J. Org. Chem. 2013, 2013, 2080.
- 166. Bhat, A. S.; Whetstone, J. L.; Brueggemeier, R. W. J. Comb. Chem. 2000, 2, 597.
- 167. Bhat, A. S.; Whetstone, J. L.; Brueggemeier, R. W. Tetrahedron Lett. 1999, 40, 2469.
- 168. Agarwal, N. N.; Soni, P. A. Indian J. Heterocycl. Chem. 2005, 14, 259.
- 169. Banerjee, D.; Kayal, U.; Maiti, G. Tetrahedron Lett. 2016, 57, 1667.
- 170. Ashihara, Y.; Nagata, Y.; Kurosawa, K. Bull. Chem. Soc. Jpn. 1977, 50, 3298.
- 171. Ghodile, N. G. Int. J. Pharm. Bio. Sci. 2013, 4, 916.
- 172. Varma, R. S.; Kumar, D. Synth. Commun. 1999, 29, 1333.
- 173. Kumar, P.; Bodas, M. S. Org. Lett. 2000, 2, 3821.
- 174. Das, J.; Ghosh, S. Tetrahedron Lett. 2011, 52, 7189.
- 175. Kavala, V.; Lin, C.; Kuo, C-W.; Fang, H.; Yao, C-F. Tetrahedron 2012, 68, 1321.
- 176. Lee, J. I.; Jung, M. G.; Jung, H. J. Bull. Korean Chem. Soc. 2007, 28, 859.

- 177. a) Kálai, T.; Kulcsar, G.; Osz, E.; Jeko, J.; Sumegi, B., Hidega, K. Arkivoc 2004, (vii), 266. b)
 Khilya, V. P.; Ishchenko, V. V. Chem. Heterocycl. Compd. 2002, 38, 883.
- 178. Bapna, M.; Nema, R. K. Asian J. Chem. 2009, 21, 1244.
- 179. Theja, D. N.; Choudary, T. P.; Reddy, M. I.; Avsss, G.; Reddy, K. U. Int. J. Pharm. Pharm. Sci. 2011, 3, 51.
- 180. Kulkarni, P. S.; Kondhare, D. D.; Varala, R.; Zubaidha, P. K.; J. Serb. Chem. Soc. 2012, 77, 1.
- 181. Costa, A. M. B. S. R. C. S.; Dean, F. M.; Jones, M. A.; Smith, D. A. J. Chem. Soc., Perkin Trans. 1 1986, 1707.
- 182. Bapna, M.; Nema, R. K. Asian J. Chem. 2008, 20, 6022.
- 183. Ares, J. J.; Outt, P. E.; Randall, J. L.; Murray, P. D.; Weisshaar, P. S.; O'Brien, L. M.; Ems, B. L.; Kakodkar, S. V.; Kelm, G. R.; Kershaw, W. C.; Werchowski, K. M.; Parkinson, A. J. Med. Chem. 1995, 38, 4937.
- 184. Manta, I.; Berger, T.; Silaghi, E. Rev. Chim. (Bucharest) 1959, 10, 69.
- 185. Rangaswami, S.; Seshadri, T. R.; Proc. Ind. Acad. Sci. (A) 1939, 9, 1.
- 186. Daniher, A.; Wang, Y. U.S. Pat. Appl. 20100272656, 2010.
- 187. Corvaisier, A.; Tirouflet, J. Compt. Rend. 1960, 251, 1641.
- 188. Matsui, T.; Tanaka, T.; Tamura, S.; Toshima, A.; Tamaya, K.; Miyata, Y.; Tanaka, K.; Matsumoto, K. J. Agric. Food Chem. 2007, 55, 99.
- 189. Shaw, J. E.; Sicree, R.A.; Zimmet, P. Z. Diabetes Res. Clin. Pract. 2010, 87, 4.
- 190. Guariguata, L.; Whiting, D. R.; Hambleton, I.; Beagley, J.; Linnenkamp, U.; Shaw, J. E. *Diabetes Res. Clin. Pract.* 2014, 103, 137.
- 191. Jia, W.; Gao, W.; Tang, L. Phytother. Res. 2003, 17, 1127.
- 192. Ríos, J. L.; Francini, F.; Schinella, G. R. Planta Med. 2015, 81, 975.
- 193. Zhao, R.; Li, Q.; Long, L.; Li, J.; Yang, R.; Gao, D. Int. J. Food Sci. Technol. 2006, 42, 80.
- 194. Mohan, S.; Eskandari, R.; Pinto, B. M. Acc. Chem. Res. 2014, 47, 211.
- 195. Tanabe, G.; Otani, T.; Cong, W.; Minematsu, T.; Ninomiya, K.; Yoshikawa, M.; Muraoka, O. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 3159.
- 196. Roy, A.; Geetha, R. V. Int. J. Pharm. Bio. Sci. 2013, 4, 49.
- 197. Kim, J-S.; Hyun, T. K.; Kim, M-J. Food Chem. 2011, 124, 1647.
























































































CHAPTER 4

Synthetic studies of coumestans

using Cu(OAc)₂



4.1: Introduction

6H-Benzofuro[3,2-c][1]-benzopyran-6-ones **1**, commonly known as coumestans, are polycyclic ring systems having a coumarin ring and a benzofuran ring fused together sharing a common C=C bond. A number of oxygenated compounds with this ring system are isolated from several natural sources, however parent coumestan **1a** is not known to occur in nature.



Figure 1: Structure of coumestan 1 with general numbering.

The general numbering of **1** is shown in figure 1. However it has been numbered in other 3 ways in the literature as shown below (Figure 2). Various names such as coumarino[3':4'- 3:2]coumarone, benzofurano[3',2':3,4]coumarin, coumarino-benzofuran, 6-*oxo*-pterocarp-6aene, pterocarpone, 6H-[1]benzofuro[3,2-*c*]chromen-6-ones, benzofuro- α -benzopyrone, and coumestones are given to this class of compounds from which the coumestones was proposed by Ollis in 1966.¹ The name pterocarpone signifies that it is one of the member of pterocarponoids group analogous to flavonoids.



Figure 2: Different numberings of 1a.

4.2: Occurrence

Coumestans² are found in diverse parts of plants such as leaves, roots and seeds.³ The different plant sources belongs to *Papilionaceae*, *Leguminosae*, and *Compositae* families, most of them being isolated from *Leguminosae* plants. Coumestans are the final oxidation products of pterocarpans and pterocarpenes. It consists of several natural members among which wedelolactone **2a** was the first natural compound isolated by Govindachari *et al.* in 1956⁴ from the leaves of *Wedelia calendulacea* (*Compositae*). Later it was also isolated from *Leclipta species*.⁵ It is interesting to know that most of the natural coumestans isolated from diverse plant sources consists of a resorcinol unit in both the benzene rings i.e. presence of oxygen functionality at C₃ and C₉ positions **2** (Figure 3, Table 1). Also few of the natural members **3** do have oxygen functionality at C₄ and/or C₈ positions (Figure 4, Table 2) whereas other compounds **4** are shown in figure 5 (Table 3).



Figure 3: Naturally occurring coumestans 2.

Table 1: Source of isolation of natural	ly occurring of	coumestans 2.
---	-----------------	---------------

No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅
	Source of isolation					
2a	Wedelolactone ^{4,5}	OH	Н	OMe	OH	OH
	Wedelia calendulacea, Eclipta alba,					
	Eclipta prostrata, Wedelia chinensis					
2b	Norwedelolactone ^{5b-c,5e}	OH	Н	OH	OH	OH
	Eclipta alba, Eclipta prostrate,					
	Wedelia chinensis					
2c	Coumestrol ^{6,8a-c,14a,23a,26a-b,31a,33a-c,34,49}	Н	Н	OH	Н	OH
	Medicago sativa, Trifolium repens,					
	Trifolium fragiferum, leguminous					
	plants, Trifolium pratense, Trifolium					
	subterraneum, annual medics, Chinese					
	milk vetch, Soyabean, legume shoots &					
	sprouts, processed food, Derris species,					
	Melilotus messanensis, Phaseolus					
	aureus, Soja hispida, soyabean roots,					
	Solanum iyratum, Pueraria mirifica,					
	Pueraria lobata, Dolichos biflorus,					
	Phaseolus lunatus, Melilotus indica					
2d	8-Methoxycoumestrol ^{7,8c}	Н	Н	OH	OMe	OH
	Medicago sativa, Arachis hypogaea,					
	Tephrosia purpurea					
2e	9-Methoxycoumestrol ⁸	Н	Н	OH	Н	OMe
	Dalbergia odorifera, Cicer species,					
	Melilotus messanensis, Medicago					
	sativa, Dalbergia oliveri, Dalbergia					
	stevensonii, Trifolium pratense,					
	Centrolobium species, Spatholobi					

	caulis					
2f	Coumestrol dimethyl ether ⁹	Н	Н	OMe	H OMe	
	Dalbergia decipularis					
2g	Medicagol ^{8c-i,10}	Н	Н	OH	-O-CH ₂ -O-	
	Cicer species, Medicago sativa,					
	Sophora chrysophylla, Cyclopia					
	intermedia, Maackia amurensis,					
	Galega officinalis, Flemingia					
	macrophylla, Sophora japonica,					
	Dalbergia oliveri, Dalbergia					
	stevensonii, Sophora tomentosa,					
	Euchresta japonica, Trifolium pratense					
2h	Lucernol ^{8c,11}	Н	OH	OH	Н	OH
	Medicago sativa					
2i	Flemichapparin C ^{10b-c,12}	Н	Н	OMe	-O-CH ₂ -O-	
	Flemingia chappar, Tephrosia					
	hamiltonii, Eysenhardtia polystachya,					
	Cyclopia intermedia, Galega					
	officinalis, Derris scandens					
2j	2-Hydroxyflemichapparin C ¹³	Н	OH	OMe	-O-CH ₂ -O-	
	Swartzia leiocalycina					
2k	Aureol ^{10d,14}	OH	Н	OH	Н	OH
	Phaseolus aureus, Hedysarum					
	multijugum, Flemingia macrophylla					
21	6-hydroxy-5,7-dimethoxy-11,12-	OMe	OH	OMe	-O-C	H ₂ -O-
	methylenedioxycoumestone ^{13a}					
	Swartzia leiocalycina					
2m	7-hydroxy,11,12-	Н	Н	OH	OMe	OMe
	dimethoxycoumestan ^{8b,15}					
	Alfalfa, Melilotus messanensis					
2n	Flemicoumestan A ¹⁶	OH	Н	OH	OMe OH	
	Flemingia philippinensis					
20	Hedysarimcoumestan A ^{14b-c}	OH	Н	OMe	Н	OMe
	Hedysarum multijugum					

2p	Hedysarimcoumestan B ^{14b-c}	OH	Н	OH	Н	OMe
	Hedysarum multijugum					
2q	Hedysarimcoumestan E ^{14b-c}	OH	Н	OH	OMe	OMe
	Hedysarum multijugum					
2r	1,3,9-Trimethoxycoumestan ^{14b}	OMe	Н	OMe	Н	OMe
	Hedysarum multijugum					
2s	2-methoxy-3,9-dihydroxy coumestan ^{12a}	Н	OMe	OH	Н	OH
	Tephrosia hamiltonii					
2t	Isotrifoliol ¹⁷	OMe	Н	OH	Н	OH
	Glycyrrhiza uralensis					



Figure 4: Naturally occurring coumestans 3.

No.	Name	R ₁	R ₂	R ₃	R ₄
	Source of isolation				
3a	Sophoracoumestan B ^{10b,18}	OH	OMe	-0-0	H ₂ -O-
	Sophora franchetiana, Cyclopia intermedia				
3b	Sativol ^{8c,11}	OMe	OH	Н	OH
	Medicago sativa				
3 c	Pongacoumestan ^{7a,19}	OH	OMe	Н	OH
	Pongamia pinnata, Arachis hypogaea				
3d	4-Hydroxycoumestrol ²⁰	OH	OH	Н	OH
	Erythrina sigmoidea				
3 e	3-Hydroxy-4,9-dimethoxycoumestan ²¹	OH	OMe	Н	OMe
	Ononis vaginalis				
3f	3,9-dihydroxy-4, 8-dimethoxycoumestan ^{7a}	OH	OMe	OMe	OH
	Arachis hypogaea				
3g	3-Hydroxy-8-methoxycoumestan ^{8j}	OH	Н	OMe	Н
	Medicago species				



Figure 5: Naturally occurring coumestans 4.

Table 3: Source of isolation of naturally occurring coumesta	ins 4 .
--	----------------

No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆
	Source of isolation						
4 a	Hedysarimcoumestan F ^{14b-c}	OH	Н	OMe	OH	Н	OMe
	Hedysarum multijugum						
4b	Wairol ²²	Н	Н	OH	OMe	Н	OMe
	Medicago sativa						
4c	Tephrosol ²³	Н	OMe	OH	Н	-O-C	H ₂ -O-
	Tephrosia villosa						
4d	Repensol ^{8a}	Н	Н	OH	OH	Н	OH
	Trifolium repens						
4 e	Trifoliol ^{8a,c,24}	Н	Н	OH	OH	Н	OMe
	Trifolium repens, Medicago						
	sativa						
4f	Melimessanol A ^{8b}	Н	OH	OMe	Н	Н	Н
	Melilotus messanensis						
4g	Mutisifurocoumarin ²⁵	CH ₃	Н	Н	Н	OH	OH
	Mutisia orbignyana, Mutisia						
	acuminata						

Some of the prenylated coursestans **5** and **6** are shown in figures 6 and 7 respectively have been isolated from various sources (Table 4-5).



Figure 6: Naturally occurring prenylated coumestans 5.

CHAPTER 4

No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇
	Source of isolation							
5a	Psoralidin ²⁶	Н	Prenyl	OH	Н	Н	Н	Н
	Psoralea corylifolia,							
	Dolichos biflorus,							
	Phaseolus lunatus							
5b	Glycyrol ^{17b}	OMe	Prenyl	OH	Н	Н	Н	Н
	Glycyrrhiza uralensis							
5c	3- <i>O</i> -Methylglycyrol ^{27b-}	OMe	Prenyl	OMe	Н	Н	Н	Н
	с							
	Glycyrrhiza species							
5d	Phaseol ^{14a}	Н	Н	OH	Prenyl	Н	Н	Н
	Phaseolus aureus							
5e	Isosojagol ²⁸	Н	Н	OH	Н	Н	Н	Prenyl
	Phaseolus coccineus,							
	Erythrina abyssinica							
5f	Sigmoidin K ^{28b,29}	Н	Prenyl	OH	Н	Н	Н	Prenyl
	Erythrina sigmoidea,							
	Erythrina abyssinica							
5g	Puerarostan ³⁰	Н	Н	OH	OMe	Н	Prenyl	Н
	Pueraria tuberosa							
5h	Mirificoumestan ³¹	Н	Н	OH	Н	Prenyl	OMe	Н
	Pueraria mirifica,							
	Pueraria hirsuta							

Table 4: Source of isolation of naturally occurring coumestans 5.



Figure 7: Naturally occurring prenylated coumestans 6.

No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇
	Source of isolation							
6a	Erythribyssin N ^{28b}	Η	Η	OH	Н	Н	Н	Prenyl
	Erythrina abyssinica							
6b	Hedysarimcoumestan D ^{14b-c}	OH	Η	OH	Η	Η	Н	Prenyl
	Hedysarum multijugum							
6c	Hedysarimcoumestan G ^{14b-c}	OH	Н	OH	Н	OH	Н	Prenyl
	Hedysarum multijugum							
6d	Hedysarimcoumestan H ^{14b-c}	OH	Η	OH	Н	OH	Prenyl	Н
	Hedysarum multijugum							
6e	Methylhedysarimcoumestan	OH	Н	OMe	Н	OH	Prenyl	Н
	$\mathrm{H}^{\mathrm{14c}}$							
	Hedysarum multijugum							

Table 5: Source of isolation of naturally occurring coumestans 6.

Other natural compounds **7-32** (Figure 8, Table 6) bearing pyran ring, hydroxyalkyl substituent, epoxide, furan ring, chromene ring, isoprenyl, geranyl and sulphate substituents are also known. Also some complex coumestans **33-43** (Figure 9, Table 7) with glucoside rings have also been isolated.



Figure 8: Structurally diverse naturally occurring coumestans 7-32.

Table 6: Source of isolation of naturally occurring coumestans 7-32.

No.	Name
	Source of isolation
7	Iso-glycyrol ^{27a-e}
	Glycyrrhiza species

8	Isopsoralidin ³²
	Psoralea corylifolia
9	Sojagol ^{6z,33}
	Soja hispida, Soyabean, leaves (Glycine max) Phaseolus aureus,
	Cylicodiscus gabunensis
10	9-Hydroxy-2',2'-dimethylpyrano[5',6':2,3]-coumestan ³⁴
	Solanum iyratum
11	Corylidin ³⁵
	Psoralea corylifolia
12	Bavacoumestan A ³⁶
	Psoralea corylifolia
13	Bavacoumestan B ³⁶
	Psoralea corylifolia
14	Glycyrurol ^{17b}
	Glycyrrhiza uralensis
15	Lespedezacoumestan ³⁷
	Lespedeza virgata
16	Mirificoumestan hydrate ^{31a}
	Pueraria mirifica
17	Mirificoumestan glycol ^{31a}
	Pueraria mirifica
18	Psoralidin 2',3'-oxide diacetate ³⁸
	Psoralea corylifolia
19	Erosnin ³⁹
	Pachyrrhizus erosus
20	Plicadin ⁴⁰
	Psoralea plicata
21	Sophoracoumestan A ^{10a,36,41}
	Sophora franchetiana, Psoralea corylifolia, Sophora chrysophylla
22	Tuberostan ⁴²
	Pueraria tuberose
23	Hirtellanines B ⁴³
	Ficus hirta, Campylotropis hirtella
24	Gancaonin F ^{27d}

	Glycyrrhiza species	
25	Tephcalostan ⁴⁴	
	Tephrosia calophylla	
26	Tephcalostan B ⁴⁵	
	Tephrosia calophylla	
27	Tephcalostan C ⁴⁵	
	Tephrosia calophylla	
28	Tephcalostan D ⁴⁵	
	Tephrosia calophylla	
29	2-(α , α -Dimethylallyl)coumestrol ⁴⁶	
	Pueraria lobata	
30	Puerarol ⁴⁷	
	Pueraria radix	
31	Solalyratin A ³⁴	
	Solanum iyratum	
32	Demethylwedelolactone 3-sulfate ^{5e,48}	
	Eclipta prostrate	



Figure 9: Naturally occurring coumestans with glucoside rings 33-43.

No.	Name		
	Source of isolation		
33	Coumestrin ⁴⁹		
	soybean roots		
34	Eriocephaloside ⁵⁰		
	Lasiosiphon eriocephalus		
35	Hedysarimcoumestan C ^{14b}		
	Hedysarum multijugum		
36	Coumestoside A ^{33d}		
	Cylicodiscus gabunensis		
37	Coumestoside B ^{33d}		
	Cylicodiscus gabunensis		

38	Coumestoside C ⁵¹				
	Cylicodiscus gabunensis				
39	Coumestoside D ⁵¹				
	Cylicodiscus gabunensis				
40	Demethylwedelolactone glucoside ^{5b,52}				
	Eclipta alba				
41	3-Hydroxy-9-methoxy-2-[2'(E)-3'-methyl-4'- O - β -D-				
	galactopyranosylbutenyl]-8-isoprenylcoumestan ⁵³				
	Picralima nitida				
42	3-Hydroxy-9-methoxy-2-[2'(<i>E</i>)-3'-methyl-4'- <i>O</i> -β-D-				
	glucopyranosylbutenyl]-8-[2''(E)-3''-methyl-4''-oxobutenyl]coumestan ⁵³				
	Picralima nitida				
43	3-Hydroxy-9-methoxy-4-[2'(<i>E</i>)-3'-methyl-4'- <i>O</i> -β-D-				
	glucopyranosylbutenyl]-8-[2''(<i>E</i>)-3''-methyl-4''-oxobutenyl]coumestan ⁵³				
	Picralima nitida				

Most of these naturally occurring coumestans are known for exhibiting diverse pharmacological activities.⁵⁴ Many herbs/plants rich in coumestans have been used as traditional medicines mostly in China and India to treat varied diseases. ^{5c,14c,55} Wedelolactone, the first natural member of coursetan inhibits many enzymes such as IKK kinase,⁵⁶ 5lipoxygenase,⁵⁷ Na⁺, K⁺ -ATPase,⁵⁸ hepatitis virus C RNA-polymerase⁵⁹ and trypsin.⁶⁰ Also its other activities includes antibacterial,⁶¹ antimicrobial,⁶² antihepatotoxic,⁶³ antimyotoxic,⁶⁴ antitumour,⁶⁵ hepatoprotective,⁶⁶ anticancer,⁶⁷ and electrochemical⁶⁸ activities. It also enhances interferon Y signalling⁶⁹ and is a potent glucosidase inhibitor and anti-glycemic agent.⁷⁰ Recently its metabolism in rats has been studied.⁷¹ Coumestrol is another immensely studied member showing diverse biological activities. It has higher binding affinity for $ER\beta$ than other phytoestrogens.⁷² It is also known to inhibit bone resorption and to stimulate bone mineralization⁷³ including some other activities.⁷⁴ Some coumestans inhibit protein-tyrosine phosphatase 1B⁷⁵ and some are used in the treatment of liver diseases.⁷⁶ Other activities shown by coumestans includes anticancer,⁷⁷ antitumour,⁷⁸ and anti-inflammatory⁷⁹ activities. Also studies have shown their widespread use as phytoalexins,⁸⁰ estrogenic,⁸¹ antibacterial⁸² and antidepressant.⁸³ These tremendous biological activities prompted us to develop an efficient method for coumestan synthesis.

Several synthetic coumestan analogues have also been prepared and studied for biological activities. Some of them such as **44-49** exhibit peculiar biological activities and their structures are depicted below (Figures 10-11, Table 8-9).



Figure 10: Synthetic coursetans 44.

Table 8:	Synthetic co	umestans 44.
----------	--------------	--------------

No.	Name	R ₁	R ₁	\mathbf{R}_2	R ₃	R ₄
44a	3-Methoxy-2,8,9-trihydroxy	Н	OH	OMe	OH	OH
	coumestan ⁸⁴					
44b	2-Methoxy-3,8,9-trihydroxy	Н	OMe	OH	OH	OH
	coumestan ^{59,85}					
44c	6 <i>H</i> -Benzofuro[3,2- <i>c</i>]chromen-6-one ⁸⁶	Н	Н	Н	Н	Н
44d	9-Methoxy-6H-benzofuro[3,2-	Н	Н	Н	Н	OMe
	c]chromen-6-one ⁸⁶					
44e	8,9-Dimethoxy-6H-benzofuro[3,2-	Н	Н	Н	OMe	OMe
	c]chromen-6-one ⁸⁶					
44f	9-Chloro-6H-benzofuro[3,2-	Н	Н	Н	Н	Cl
	c]chromen-6-one ⁸⁶					
44g	9-Bromo-6 <i>H</i> -benzofuro[3,2-	Н	Н	Н	Н	Br
	<i>c</i>]chromen-6-one ⁸⁶					
44h	9-Hydroxy-6H-benzofuro[3,2-	Н	Н	Н	Н	OH
	c]chromen-6-one ⁸⁶					
44i	8,9-Dihydroxy-6H-benzofuro[3,2-	Н	Н	Н	OH	OH
	<i>c</i>]chromen-6-one ^{86,87}					
44j	3-Ethoxy-1,8,9-trihydroxy-6H-	OH	Н	OEt	OH	OH
	benzofuro[3,2-c]chromen-6-one ⁸⁸					
44k	3-Decyloxy-1,8,9-trihydroxy-6 <i>H</i> -	OH	(CH ₂) ₉	OEt	OH	OH
	benzofuro[3,2-c]chromen-6-one ⁸⁸		CH ₃			



Figure 11: Complex synthetic coursetans 45-49.

Table 9:	Complex	synthetic	coumestans	45-49 .
----------	---------	-----------	------------	----------------

No.	Name		
45	2-ferrocenyl-furo[3,2-c]chromen-4-one ⁸⁹		
46	Naphtho coumestans ⁹⁰		
47	3-Hydroxy-9-methoxy-2-[2'(<i>E</i>)-4'-hydroxy-3'-methylbutenyl]-8-		
	isoprenylcoumestan ⁵³		
48	3-Hydroxy-9-methoxy-2-[2'(<i>E</i>)-4'-hydroxy-3'-methylbutenyl]-8-[2''(<i>E</i>)-		
	3''-methyl-4''-oxobutenyl]coumestan ⁵³		
49	3-Hydroxy-9-methoxy-4-[2'(<i>E</i>)-4'-hydroxy-3'-methylbutenyl]-8-[2''(<i>E</i>)-		
	3''-methyl-4''-oxobutenyl]coumestan ⁵³		

4.3: Literature synthetic methods

Owing to immense biological importance of coumestan compounds, several methods for the synthesis of this tetracyclic ring system have been reported.⁹¹ Based on retrosynthetic analysis, the synthetic methods can be broadly divided into 4 categories as depicted below. These 4 approaches have been classified on the basis of i) formation of furan ring on preformed coumarin ring (Scheme 1, Route A), ii) formation of coumarin ring on preformed furan ring (Scheme 2, Route B), iii) simultaneous formation of C-C and C-O bonds to form furan ring (Scheme 3, Route C), iv) from flavonoids (Scheme 4, Route D) and v) miscellaneous routes (Scheme 5, Route E).



Scheme 1: Formation of furan ring on preformed coumarin ring (Route A).



Scheme 2: Formation of coumarin ring on preformed furan ring (Route B).



Scheme 3: Simultaneous formation of C-C and C-O bonds to form furan ring (Route C).



Scheme 4: From flavanoids (Route D).



Scheme 5: Miscellaneous routes (Route E).

All these approaches involving diverse substrates have been discussed ahead. Some of the methods have also been applied for the synthesis of natural coursetans.

4.3.1: Formation of furan ring on preformed coumarin ring (Route A)

4.3.1.1: From demethylative or dehydrative cyclization

Syntheses of coumestans were reported many decades ago from 3-(2-methoxyphenyl)-4hydroxycoumarin intermediate (Scheme 6). This was prepared by various groups using different methods. Bowyer *et al.*⁹² synthesized it *via* cyclization of corresponding deoxybenzoin with ethyl carbonate and sodium dust in 1957. This intermediate was then converted to coumestan using 48 % HBr in boiling acetic acid.

Synthesis of parent ring system **1** present in wedelolactone **2a** and of tri-Omethylwedelolactone was reported by Govindachari *et al.*⁹³ in the same year. The coumarin intermediate was prepared from intramolecular Claisen condensation of methyl-O,O'methoxyphenylacetoxybenzoate in presence of sodium. It was further demethylated on prolonged treatment of pyridine hydrochloride or by heating at 280 °C.



Scheme 6

Deschampo-Vallet and Mentzer⁹⁴ reported coumestan synthesis from the coumarin intermediate by treatment with pyridine hydrochloride. The intermediate was obtained on thermal condensation of phenol with *o*-methoxyphenyl diethyl malonate.

Govindachari *et al.*⁹⁵ synthesized the coumarin intermediate by condensation of corresponding deoxybenzoin with diethyl carbonate and pulverized sodium. Demethylative cyclization of the intermediate with pyridine hydrochloride resulted in coumestan.

Similarly Emerson and Bickoff⁹⁶ synthesized the coumarin intermediate by condensation of deoxybenzoin with methyl chloroformate in presence of potassium carbonate. Subjecting it to aniline hydrochloride delivered coumestan. Aniline hydrochloride was also utilized by Nasipuri and Pyne⁹⁷ for coumestan synthesis. The intermediate coumarin was obtained by treating deoxybenzoin with ethyl chloroformate method. Finally demethylative ring closure with HI at 170 °C yielded coumestan.⁹⁸

Uma Rani and Darbarwar⁹⁹ synthesized various oxygenated coumestans and their acetates by dehydrative cyclization of 4-hydroxy-3-arylcoumarins using methanolic hydrogen chloride (Scheme 7).





4.3.1.2: From disconnection a

Kappe *et al.*¹⁰⁰ demonstrated the preparation of coumestan in 1978. 4-Hydroxycoumarin was reacted with (diacetoxy)iodoarene to from iodonium ylide which rearranged to 4-aryloxy-3-iodocoumarin on heating. Reductive deiodination occurred when it was heated with Zn-HOAc to give 4-aryloxycoumarin which was converted to coumestan on photocyclization in presence of iodine. Also 4-aryloxy-3-iodocoumarin provided coumestan on photocyclization. However, low substrate scope and poor yield limited the use of this approach. Later this method was modified wherein 4-aryloxy-3-iodocoumarin was cyclised using Heck condition¹⁰¹ (Scheme 8).



Scheme 8

Also Hong *et al.*¹⁰² developed an efficient one pot cyclization process by reacting iodoarenes and various arylols *via* oxidative palladium catalysis (Scheme 9). The methodology was extended towards the synthesis of parent coumestan *via* sequential oxidation of iodophenol to hypervalent iodine (III) species, 3-iodination of 4-hydroxycoumarin to form 3-iodo-4-phenoxycoumarin through iodonium ylide intermediate and Pd catalyzed C-H functionalization and cyclization.



Scheme 9

Recently, McGlacken and co-workers¹⁰³ synthesized coumestan by an intramolecular coupling using palladium catalyst. Accordingly 4-phenoxycoumarin was chlorinated at 3 position using NCS in trifluoroacetic acid which on intramolecular coupling between C-Cl and phenoxy C-H bond yielded coumestan. This strategy was successfully applied to the synthesis of natural coumestan flemichapparin C (Scheme 10).



Scheme 10

A palladium catalyzed intramolecular cross dehydrogenative coupling (CDC) was designed by Cheng *et al.*¹⁰⁴ for the synthesis of coumestans. The methodology was also applied for the synthesis of Coumestrol and Flemichapparin C (Scheme 11).



Scheme 11

McGlacken and co-workers¹⁰⁵ reported a palladium catalyzed double C-H activation of coumarin moieties to form coumestans. Flemichapparin C was synthesized using this methodology (Scheme 12).



Scheme 12

Singh and Singh¹⁰⁶ prepared alkylated product from 4-hydroxycoumarin and 2bromocyclohexanone which was cyclised and dehydrogenated by polyphosphoric acid and DDQ respectively (Scheme 13).



Scheme	13
--------	----

Burns *et al.*¹⁰⁷ developed a palladium catalyzed methodology for the C-H functionalization of 2-pyrones which was extended for the synthesis of parent coumestan (Scheme 14).



Kapdi and co-workers¹⁰⁸ achieved an intramolecular C-H bond functionalization of 4-(2bromophenoxy)coumarins with [Pd(PPh₃)₂(saccharinate)₂] as the palladium catalyst source (Scheme 15). The substrates with electron releasing/withdrawing substituent gave good yield of the products.



Scheme 15

A variety of coumesatns were synthesized by Kapdi *et al.*¹⁰⁹ by using phospha-palladacycle *via* intramolecular C-H bond functionalization of 4-(2-bromophenoxy)coumarins. The direct one pot conversion of 4-chlorocoumarin to coumestan was also achieved under microwave irradiation (Scheme 16).



Scheme 16

Coumestan has been synthesized from 3-[2-cyclohexenyl]-4-hydroxy-1-benzopyran-2(H)-one by Majumdar *et al.*¹¹⁰ *via* oxymercuration. It was treated with mercuric acetate in methanol followed by dehydrogenative demercuration with Pd/C in refluxing diphenyl ether. Alternatively it was prepared in 2 steps from 4-(2-cyclohexenyloxy)-1-benzopyran-2(H)-one by refluxing in diphenyl ether first followed by addition of Pd/C. Also direct synthesis of coumestan was obtained when 4-(2-cyclohexenyloxy)-1-benzopyran-2(H)-one was refluxed in presence of Pd/C in diphenyl ether (Scheme 17).



Scheme 17

4.3.1.3: From disconnection b

Synthesis of coumestans was carried out by the oxidative cyclization of 4-hydroxy-3-phenyl-2*H*-chromen-2-one as a starting material¹¹¹⁻¹¹² (Scheme 18). Kappe and Schmidt had employed Pd/C in refluxing diphenyl ether at 258 °C.^{111a} Later coumestrol was synthesized
using this method.^{111b} Recently, Tang *et al*.¹¹² accomplished FeCl₃ mediated synthesis from 4-hydroxy-3-phenyl-2*H*-chromen-2-one.



Scheme 18

4.3.1.4: From disconnection c

Kurosawa and Nogami¹¹³ synthesized coumestans by the oxidative cyclization of 3-(2-hydroxyphenyl)coumarins using lead tetraacetate (LTA). These 3-(2-hydroxyphenyl)coumarins intermediate were obtained by the Perkin reaction of salicylaldehydes and 2'-hydroxyphenylacetic acids. Along with coumestans, 3-(1-acetoxy-4-methoxy-2-oxo-3,5-cyclohexadienyl)coumarins were also formed (Scheme 19).

Wadia and co-workers¹¹⁴ also reported the synthesis of coumestan by using LTA. The method involves the use of salicylaldehyde and 2-phenylthioacetamide as starting materials in presence of POCl₃ to give 3-(2-benzyloxyphenyl)coumarin which underwent debenzylation in acidic medium forming 3-(2-hydroxyphenyl)coumarin. Oxidative cyclization using LTA converted this coumarin to corresponding parent coumestan (Scheme 19).



Mali and Tilve¹¹⁵ carried out Wittig reaction between phosphorane and ethyl-2-(2methoxyphenyl)-2-oxoacetate to form ester which was converted to 3-(2hydroxyphenyl)coumarin by heating with pyridine hydrochloride. DDQ was then employed for the oxidative cyclization to afford coumestan (Scheme 20). Similar strategy was used by Pandit and Gadre¹¹⁶ however 3-(2-hydroxyphenyl)coumarin was developed by an alternative route involving demethylation followed by cyclization of cinnamic acid with pyridine hydrochloride (Scheme 20).



Scheme 1	20
----------	----

Gong *et al.*¹¹⁷ prepared 3-(2-hydroxyphenyl)coumarin intermediate by using Perkin reaction from salicylaldehydes and *o*-hydroxyphenylacetic acids in the presence of sodium acetate, acetic anhydride and acetic acid. The oxidative cyclization of this intermediate was easily achieved by using stoichiometric PdCl₂ in DMF at 150 °C (Scheme 21).



Scheme 21

Chang *et al.*¹¹⁸ described the total synthesis of hedysarimcoumestan B, demethylwedelolactone and wedelolactone by using I_2 in anhyd. pyridine for oxidative cyclization as one of the key steps (Scheme 22).



Vinyl C-H lithiation of bis-*ortho*-methoxy *cis*-stilbene was carried out by O'Shea and coworkers¹¹⁹ followed by CO_2 quench to provide access to the targeted cinnamic acid. This acid

underwent demethylation with BBr₃ followed by treatment with base and oxidative cyclization with DDQ to form coumestan (Scheme 23).





Recently Sheng *et al.*¹²⁰ reported the total synthesis of naturally occurring coumestrol and aureol. Nucleophilic addition of *m*-bromophenol to glyoxylic acid resulted in the formation of 2-bromo-4-hydroxymandelic acid which further on reduction with SnCl₂/HCl afforded 2-bromo-4-hydroxyphenylacetic acid. This on Perkin condensation with *o*-hydroxybenzaldehydes gave acylated 2'bromo-3-arylcoumarins which was deacetylated in hot NaOH followed by acidification. Finally a consecutive Cu-catalyzed hydroxylation and aerobic oxidative cyclization carried out with Cu(OAc)₂/1,10-phen, KOH in DMSO, MW at 120 °C afforded coumestrol and aureol (Scheme 24).



Scheme 24

4.3.2: Formation of coumarin ring on preformed furan ring (Route B)

4.3.2.1: From disconnection a

James *et al.*¹²¹ carried out Suzuki Miyaura cross coupling reaction of aryl-*O*-carbamoyl *ortho*boronic acids with benzofuran iodide delivered to form coupled product. This was converted to coumestans on treatment with LDA *via* directed ortho lithiation followed by acetic acid reflux (Scheme 25).



Scheme 25

4.3.2.2: From disconnection b

Hiroya *et al.*¹²² reported the synthesis of parent coumestan and coumestrol. 2-Iodophenol was converted to diaryl acetylene under an acetylene atmosphere by using palladium catalyzed cross coupling reaction. Next deacetylation was carried out followed by the intramolecular carbonylative cyclization reaction under carbon monoxide atmosphere leading to parent coumestan. For the synthesis of coumestrol appropriately protected compound was converted to corresponding benzofuran tosyl ester. When this compound was treated with BBr₃ double demethylation occurred to obtain corresponding dihydroxy compound. This on alkaline hydrolysis followed by lactonization in presence of acid catalyst rendered coumestrol (Scheme 26).



Scheme 26

Ethyl 2-methoxybenzoylacetates on Michael addition with 1,4-benzoquinone produced benzofuran-carboxylates in presence of $ZnCl_2$, was reported by McPherson and Ponder.¹²³ Further treatment with anhyd. pyridine-HCl at 190-195 °C delivered coumestans (Scheme 27).



Jurd¹²⁴ developed a method starting from appropriate flavylium chloride salt which was oxidized using hydrogen peroxide in aqueous methanol (Scheme 28). This resulted in the formation of carbomethoxybenzofuran intermediate which rapidly lactonised on acidification to deliver coumestrol.^{124a-c} Later Jurd¹²⁵ synthesized 7-methoxycoumestrol and 12-methoxycoumestrol using this strategy. Spencer *et al.*¹²⁶ synthesized three position isomers of coumestrol from appropriately substituted flavylium salts, their dimethyl ethers and diacetates and many monomethyl ethers and their acetates.



Scheme 28

Maeda *et al.*¹²⁷ carried out studies towards the preparation of lignans by oxidative coupling reaction. In addition to this, a coumestan derivative was synthesized from the ester starting in 3 steps involving hydrolysis and lactonization forming coumarin ring in the last step (Scheme 29).



Scheme 29

Coumestrol and its analogues formation from iron based cross dehydrogenative coupling (CDC) approach was disclosed by Pappo and co-workers¹²⁸ This two step method employed FeCl₃, 2,2'-bipyridine and di-*tert*-butyl peroxide (DTDB) as the oxidant for cross coupling of β -ketoesters and phenols to form benzofuran derivatives. Authors later modified the conditions wherein catalytic FeCl₃ in aerobic medium was preferred over the former condition. Benzofuran derivatives were then converted to coumestrol analogues by performing deprotection and lactonization steps in one pot (Scheme 30).



Scheme 30

4.3.2.3: From disconnection a, b

Larock and Harrison¹²⁹ has developed a method using organomercurial compound prepared by acetoxymercuration of 1,2-*bis*-(*o*-methoxyphenyl)acetylene. It involves carbonylation, subsequent demethylation and cyclization of organomercurial to provide coumestan ring system in 90 % yield (Scheme 31).



Scheme 31

Larock and co-workers¹³⁰ synthesized parent coumestan, coumestrol and plicadin from corresponding starting materials by iodocyclization, Pd-catalyzed intramolecular lactonization and deprotection (Scheme 32).





4.3.2.4: From disconnection c

Thasana *et al.*¹³¹ synthesized coumestan from C-O coupling reaction of 2-(2-bromophenyl)-6methoxybenzofuran-3-carboxylic acid catalyzed by copper (I) salts using microwave irradiation. Authors developed two methods of which Cu(I) thiophene-2-carboxylate (CuTc) mediated lactonization (method A) gave better results than CuI catalyzed lactonization (method B) (Scheme 33).





4.3.2.5: From disconnection b, FGI

Kraus and Zhang¹³² synthesized parent coumestan and coumestrol involving photochemical reaction as the key step. Benzofuran dione reacted with MOM protected benzyl bromide in presence of sodium methoxide leading to methyl-2-(2-methoxymethoxybenzyloxy)phenyl glyoxylate which on photochemical irradiation cyclises to give methyl-(2-hydroxy-4-methoxyphenyl) glyoxylate. Treatment with HCl generated required coumestan *via* deprotection, dehydration and lactonization. Similarly for coumestrol synthesis, an appropriate starting keto ester was coupled with the benzyl alcohol substrate forming keto ester intermediate which on irradiation followed by acid treatment resulted in coumestrol dimethyl ether. Finally it was converted to coumestrol using BBr₃ (Scheme 34).



4.3.2.6: From disconnection b, e

Liu *et al.*¹³³ reported an efficient synthesis of coumestans by a novel [3 + 3] annulation strategy (Scheme 35). Authors carried out the palladium catalyzed and copper(I) thiophene-2-carboxylate (CuTc) mediated C-S activated cross coupling of 2-(methylthio)benzofuran-3-carboxylates with 2-hydroxyphenylboronic acids and sequential intramolecular transesterification process under Liebeskind-Srogl conditions.





4.3.2.7: From disconnection b, f

Donnelly *et al.*¹³⁴ carried out the oxidation of 2,4,2',4'-tetrahydroxychalcone using alkaline hydrogen peroxide to form 2',4',6-trihydroxy-2-phenylbenzofuran-3-carboxylic acid which lactonized to coumestrol (Scheme 36).



Scheme 36

4.3.3: Simultaneous formation of C-C and C-O bonds to form furan ring (Route C)

Wanzlick *et al.*¹³⁵ synthesized coumestan by dehydrogenative coupling of catechol with 4hydroxycoumarins in presence of potassium ferricyanide, sodium acetate and aqueous acetone (Scheme 37). Potassium iodate has also been used in place of potassium ferricyanide for coumestan preparation. This method is suitable for the preparation of 8,9-oxygenated coumestans in high yields, however other coumestans without 8,9-oxygenation pattern cannot be synthesized. This procedure has been adopted in the synthesis of many natural and nonnatural coumestan analogs.^{10h,136}



Scheme 37

Many syntheses have employed 4-hydroxycoumarin along with catechol as the starting materials (Scheme 38).¹³⁷ These are mostly electrochemical synthesis wherein catechol and its derivatives produces *o*-quinones which act as a Michael acceptor and a variety of coumarin derivatives act as nucleophile. Also a report showed mushroom tyrosinase catalysed the synthesis of coumestans from same substrates.¹³⁸



Leutbecher *et al.*¹³⁹ developed a Laccase-catalyzed domino method by which coumestans were synthesized from 4-hydroxycoumarins and catechols using molecular oxygen as an oxidant. Similarly, recently Wellington and co-workers¹⁴⁰ synthesized several coumestan derivatives *via* one pot laccase-catalyzed methodology and were evaluated for anticancer activity (Scheme 39).



Scheme 39

Gong *et al.*¹⁴¹ later synthesized wedelolactone derivatives by developing a methodology involving an intermolecular cycloaddition reaction from catechol and 4-hydroxycoumarins using ammonium persulphate as oxidant (Scheme 40).





Shah and Trivedi¹⁴² obtained 3,4-dihydropyranocoumestans and 3,4-dihydropyranocoumestans by oxidative coupling of pyranobenzopyrans with catechol in the presence of HIO_4 (Scheme 41).



Scheme 41

Crude onion peroxidase extract catalyzed the reaction of catechol and different heterocyclic 1,3-dicarbonyl compounds as studied by Angeleska *et al.*¹⁴³ When 4-hydroxycoumarin was employed, domino reaction occur leading to the formation of coumestan product which exhibited potent antioxidant activity (Scheme 42).



Neog *et al.*¹⁴⁴ developed a palladium catalyzed cascade reaction of 4-hydroxycoumarins and *in situ* generated arynes. It involves C-H bond activation and C-O and C-C bond formation. The methodology was applied for the synthesis of flemichapparin C (Scheme 43).



Scheme 43

4-Hydroxycoumarin was coupled readily with *p*-benzoquinone and its derivatives to afford the corresponding 3-(p-benzoquinonyl)-4-hydroxycoumarins by Wagh and Usgaonkar¹⁴⁵ They were successfully reduced with ascorbic acid to the corresponding 3-(2,5dihydroxyphenyl)-4-hydroxycoumarins which were cyclodehydrated to give 8hydroxycoumestans (Scheme 44).



Scheme 44

Another photochemical study was carried out by Rodríguez and Baumgartner¹⁴⁶ wherein 4hydroxycoumarin on treatment with *o*-dihalobenzenes resulted in biaryl coupling followed by intramolecular heterocyclization affording coumestan and reduced product 4-hydroxy-3phenylcoumarin. In addition, 4-hydroxy-3-(2-chlorophenyl)coumarin was also obtained when 2-chloroiodobenzene was used as starting (Scheme 45).



Scheme 45

Darbarwar *et al.*¹⁴⁷ carried out a condensation reaction of 4-hydroxycoumarin and 2chlorocyclohexanone in the presence of anhyd. potassium carbonate and xylene to form an intermediate which on intramolecular cyclization resulted in a hemiketal. This was then easily dehydrated giving rise to tetrahydrocoumestan. Finally its dehydrogenation using Pd/C resulted in coumestan formation (Scheme 46). The disadvantage includes the inability to prepare halo substituted coumestans due to dehalogenation occurring during dehydrogenation.



Scheme 46

Various 4-hydroxycoumarin derivatives were treated with vinyl sulfide in the presence of silver carbonate on celite giving rise to furocoumarins by Lee *et al.*¹⁴⁸ These compounds were then dehydrogenated using Pd/C in diphenyl ether to provide coumetstans (Scheme 47).



Scheme 47

4.3.4: From Flavonoids (Route D)

Conversion of 2',4',7-trimethoxyisoflavanone into coumestrol using pyridine hydrochloride at 180-200 °C was reported by Dewick *et al.*¹⁴⁹ (Scheme 48).





Krishna Prasad *et al.*¹⁵⁰ reported an efficient three step synthesis of tuberostan from benzyloxyisoflavone. This isoflavone was converted to pterocarpan *via* hydrogenative cyclization involving debenzylation, reduction of double bond and carbonyl function and cyclization steps. Condensation of it with 2-methylbut-3-en-2-ol in presence of boron-trifluoride-diethyl ether gave prenylated compound which on treatment with DDQ afforded tuberostan. Also, acetylation of pterocarpan followed by DDQ oxidation resulted in the formation of 9-acetoxy-3-methoxy coumestan (Scheme 49).



Scheme 49

Oxidation of the pterocarpan and pterocarpene systems to coumestanes has been carried out by Ferreira *et al.*¹⁵¹ using DDQ (Scheme 50).



Scheme 50

Gunning *et al.*¹⁵² synthesized coursestrol dimethyl ether from the corresponding pterocarpan by employing DDQ (Scheme 51).



Scheme 51

Rukmani Iyer and co-workers¹⁵³ condensed chromene with 2-chloromercurio-4,5methylenedioxyphenol in presence of lithium chloropalladite to form neorautane which was oxidized using DDQ to render coumestan derivative (Scheme 52). Later using this method many reports have been published wherein various coumestan derivatives have been synthesized.^{84a,154}



Scheme 52

Bowyer *et al.*¹⁵⁵ prepared coumestrol dimethyl ether from corresponding pterocarpene using chromic oxide (CrO_3) (Scheme 53).



Scheme 53

Sant'Ana *et al.*¹⁵⁶ also used DDQ for the synthesis of coumestan from the appropriate pterocarpene which was obtained from the corresponding iodo compound by an intramolecular Heck reaction (Scheme 54).



Scheme 54

A Mitsunobu coupling of 3-hydroxymethylbenzofurans with *o*-iodophenols carried out by Fowler *et al.*¹⁵⁷ resulted in 3-(2-iodophenoxy)methylbenzofurans which on 6-endo Heck cyclization under Jeffery conditions provided access to pterocarpenes. The parent coumestan was obtained when the unsubstituted pterocarpene was oxidized using PCC (Scheme 55).



Scheme 55

Recently Kim and co-workers¹⁵⁸ demonstrated the formal synthesis of coumestrol and plicadin as well as the total synthesis of flemichapparin C using similar strategy wherein the respective starting iodo carbonyl compounds on treatment with BCl₃ delievered the corresponding benzofurans by regioselective ring closure. Further, palladium catalyzed intramolecular direct arylation of these benzofurans resulted in the pterocarpenes which on oxidation with DDQ afforded naturally occurring coumestans (Scheme 56).



Scheme 56

Takeda *et al.*¹⁵⁹ synthesized coumestan by constructing its benzofuran ring first. *O*-Phenylhydroxylamine and 4-chromanone were condensed to give oxime ether which underwent sequential acylation and rearrangement on treatment with trifluoroacetyltriflate (TFAT) furnishing tetracyclic derivative. It was finally converted to the desired coumestan product by PCC as oxidizing reagent (Scheme 57).



Scheme 57

Ghosh *et al.*¹⁶⁰ developed a one pot method which involves the arylation of ethyl acetohydroxamate with diphenyliodonium tetrafluoroborate in absence of any transition metal. The *O*-arylated product thus formed was reacted *in situ* with ketone under acidic medium to form benzofuran through oxime formation, [3,3]-rearrangement and cyclization. The coumestan has been formally synthesized from this benzofuran using PCC oxidation (Scheme 58).



Scheme 58

4.3.5: Miscellaneous routes (Route E)

Chatterjea and Roy¹⁶¹ obtained ketonitrile intermediate on condensing *o*methoxyphenylacetonitrile with ethyl-*o*-methoxybenzoate in the presence of sodium ethoxide in benzene. Ketonitrile was then converted to coumestan by treating with HBr in acetic acid (Scheme 59). Chatterjea later synthesised coumestrol from corresponding ketonitrile using this method.¹⁶²



Scheme 59

Kawase¹⁶³ synthesized coumestrol from ketonitrile intermediate 2,4-dimethoxybenzoyl-2,4dimethoxyphenylacetonitrile by the action of pyridine hydrochloride or hydriodic acid. The ketonitrile intermediate was obtained from the corresponding phenylacetonitrile and ester in the presence of sodium hydride. Kawase later described the action of HBr, HI, Py.HCl and AlCl₃ on ketonitrile and ketoester intermediate giving coumestan derivatives (Scheme 60).¹⁶⁴



Scheme 60

Chatterjea and Prasad¹⁶⁵ synthesized tri-*O*-methylwedelolactone and dihydroerosnin from the corresponding ketonitrile intermediate by employing pyridine hydrochloride (Scheme 61). The ketonitrile intermediate was prepared by reacting the corresponding phenylacetonitrile with ester using sodium hydride.





Zhang *et al.*¹⁶⁶ synthesized coumestans *via* a two-step one-pot tandem demethylationannulation-oxidation reaction from 2,3-*bis*(2-methoxyphenyl)-3-oxopropanals (Scheme 62). The products were obtained in good to excellent yields and halo-substituted products were further functionalized by using transition metal catalyzed cross-coupling reactions.





Kamara *et al.*¹⁶⁷ used chalcones for direct transformation into coumestans in presence of thallium(III)nitrate *via* oxidative rearrangement (Scheme 63). Three natural coumestans namely flemichapparin C, medicagol and sophoracoumestan B have been synthesized. However chalcone was previously converted to coumestan using thallium(III)nitrate with a longer route.¹⁶⁸ Also flemichapparin-B and flemichapparin-C have been synthesized using this approach.¹⁶⁹



Scheme	63
--------	----

Litinas and Stampelos¹⁷⁰ studied the reaction between phosphorous ylide and salicylaldehydes under various reaction conditions. When heated at the reflux temperature of salicylaldehyde the reaction resulted in the formation of coumestan, benzofuranone and coumarin derived compounds. When benzofuranone derivative was refluxed in xylene, coumestan along with coumarin derivative were produced (Scheme 64).



Scheme 64

Authors also prepared coumestan by refluxing coumarin derivative in xylene or with DDQ in toluene. Also, a reaction between another ylide and salicylaldehyde was studied which on refluxing in toluene or diglyme or under neat heating at the boiling point of salicylaldehyde provided coumestan and coumarin derivatives (Scheme 65).



Scheme 65

Chiang *et al.*¹⁷¹ converted 3-diazo-2-oxo-2,3-dihydrobenzofuran photochemically into quinonoid cumulenone which formed coumestan along with other two dimerized products by a remarkably facile addition-cyclization-(elimination) reaction (Scheme 66).



Scheme 66

Tollari *et al.*¹⁷² disclosed a rare example of coumestan synthesis from the rhodium (II) acetate catalyzed decomposition of 3-diazobenzopyran-2,4(3*H*)-dione. Coumestan was obtained as minor product (5-10 %) along with isomeric 2-substituted furo[3,2-*c*]coumarin in 45 % and furo[2,3-*b*]coumarin in 37 % by a formal [3 + 2] cycloaddition (Scheme 67). Coumestan is formed by insertion of Rh-stabilized carbenoid into the solvent followed by intramolecular etherification.



Scheme 67

A short coumestrol synthesis was achieved by Al-Maharik and Botting¹⁷³ It involved reaction of methyl 2-hydroxy-4-methoxy-phenylacetate with protected hydroxyl group and 2,4dimethoxybenzoyl chloride to afford methyl ester of 3-oxo propanoates. Among these, the methoxy protected intermediate on treating with excess of BBr₃ directly led to the formation of coumestrol via tandem demethylation and intramolecular cyclization. Also the authors have synthesized multiple ¹³C labelled coumestrols *viz*. [6,6a,11a-¹³C₃]coumestrol by adapting this method from suitable starting materials (Scheme 68).



Scheme 68

Recently, Pahari *et al.*¹⁷⁴ reported synthesis of various psoralidin derivatives using similar strategy *via* BBr₃ mediated one pot demethylation and cyclization sequence. Authors have also reported the first synthesis of lespeflorin I_1 .

Liu *et al.*¹⁷⁵ investigated a tin tetrachloride catalyzed synthesis of substituted benzofurans by highly regioselective allylic substitution of quinone monoketals with α -oxoketene dithioacetals *via* a formal [3 + 2] cycloaddition process. Accordingly coumestan derivatives

were obtained from quinone monoketals and vinylogous thioester by employing SnCl₄ as catalyst (Scheme 69).



Scheme 69

4.4: Results and Discussion

Literature studies show various routes for the synthesis of coumestans. However many of these methods have their own limitations like multistep syntheses, expensive reagents/catalysts usage, hazardous metal catalysts, difficulty in handling of reagents and/or its excessive requirement, troublesome reaction work up and product isolation. Hence continuous search for new method/reagent/catalyst for coumestan synthesis is pursued. In order to overcome the aforementioned limitations we devised a retrosynthetic route for coumestan formation which would require simple substrates and reagents.



Scheme 70: Retrosynthetic analysis of coumestan 1.

Initially a one pot retrosynthetic pathway was thought from simple substrates such as 2'hydroxybenzaldehydes **50** and 2-coumaranone **51** (Scheme 70). We thought that any suitable reagent could directly provide us the target molecule **1** *via* intermediate 3-(2hydroxyphenyl)coumarins **52** through condensation followed by oxidative cyclization.

4.4.1: One pot approach

Thus a one pot synthesis of coumestan **1a** from 2-coumaranone **51** and 2'hydroxybenzaldehyde **50a** as the starting materials was visualized. Various reaction conditions were tried for its synthesis (Table 10). FeCl₃ is already known for C-O bond formation (Scheme 18),¹¹² so we envisioned its use in coumestan synthesis. Firstly, the reaction was carried out using FeCl₃ (2.5 equiv) in presence of triethylamine in 1,2dichloroethane solvent under reflux condition for 24 h (entry 1). However no coumestan formation was seen. Similarly several other reaction conditions including NEt₃ and I₂ in absence or presence of pyridine under reflux condition (entries 2-3), N,N-dimethylaniline in presence of I₂ at 150 and 170 °C (entries 4-5) and N,N-dimethylaniline in presence of 0.3 equiv of PdCl₂ (entry 6) failed to give product formation. Further stepwise one pot approach was tried wherein the reaction was first refluxed in NEt₃ solvent for 1 h followed by its removal and then the addition of catalyst such as $Pd(PPh_3)_4$ (0.1 equiv) (entry 7) and reagent FeCl₃ (2.5 equiv) in 1,2-dichloroethane under reflux for 24 h was carried out (entry 8). However both these attempts were unsuccessful. Similarly, the stepwise one pot reaction was carried out using FeCl₃ (2.5 equiv) in presence of 230-400 silica gel (1:1 wt./wt. FeCl₃), but no desired product was formed (entry 9). Since 2.5 equiv of FeCl₃ was used in previous reactions, we thought to further increase the amount of FeCl₃ to look for any product formation. Hence directly 10 equiv of $FeCl_3$ and 1,2-dichloroethane solvent were added and refluxed for 24 h after 1 h refluxing in NEt₃. It was delightful to see coumestan formation (characterized later) albeit in trace amount (entry 10). The product formation was encouraging but in unacceptable yield so further standardisation was required. Also some more one pot reaction conditions were tried including 10 wt % Pd/C, 20 wt % Pd/C and 20 wt % Pd/C along with 10 mol% L-proline in diphenyl ether solvent which failed to give the desired product (entries 11-13).

	50a 51		0~<	
Sr. No.	Reaction condition	Time (h)	Temperature	Yield (%)
1)	NEt ₃ , FeCl ₃ (2.5 equiv), 1,2- dichloroethane	24	reflux	0
2)	NEt ₃ , I_2 (1 equiv)	4	reflux	0
3)	NEt ₃ , I ₂ (1 equiv), py	24	reflux	0
4)	N,N-Dimethylaniline, I ₂ (1 equiv)	4	150 °C	0
5)	<i>N</i> , <i>N</i> -Dimethylaniline, I_2 (1 equiv)	4	170 °C	0
6)	<i>N</i> , <i>N</i> -Dimethylaniline, PdCl ₂ (0.3 equiv)	24	150 °C	0
7)	NEt ₃ , Pd(PPh ₃) ₄ (0.1 equiv)	7	reflux	0
8)	NEt ₃ , later FeCl ₃ (2.5 equiv), 1,2- dichloroethane	24	reflux	0

 Table 10: Various reaction conditions attempted for direct synthesis of 1a from 50a & 51.

+

Reaction condition

9)	NEt ₃ , later FeCl ₃ (2.5 equiv), 230- 400 silica gel (1:1 wt./wt. FeCl ₃), 1,2-dichloroethane	24	reflux	0
10)	NEt ₃ , later FeCl ₃ (10 equiv), 1,2- dichloroethane	24	reflux	trace
11)	Pd/C (10 wt %), diphenyl ether	2	reflux	0
12)	Pd/C (20 wt %), diphenyl ether	3	reflux	0
13)	Pd/C (20 wt %), L-proline (10 mol%), diphenyl ether	3	reflux	0

Since one pot approach employing **50a** and **51** as starting materials either failed or resulted in trace amount of the desired coumestan product, a two step protocol was undertaken wherein the intermediate 3-(2-hydroxyphenyl)coumarin **52a** was to be isolated and then oxidatively cyclized.

4.4.2: Two step approach

The required substrates 3-(2-hydroxyphenyl)-coumarin **52a-y** in case of two step approach were prepared from salicylaldehydes **50** and 2-coumaranone **51** (Schemes 71)¹⁷⁶ and salicylaldehydes **50** and 2'-hydroxyphenylacetic acids **53** (Schemes 72)¹⁷⁷ as shown below.



Scheme 71: Synthesis of substrates 52a-o.



Scheme 72: Synthesis of substrates 52p-y.

From the literature survey it was clear that only four reagents are available for the oxidative cyclization of **52** (Scheme 73).^{113,115,117-118} First Pb(OAc)₄ in refluxing anhyd. benzene,¹¹³ later DDQ in refluxing anhyd. benzene,¹¹⁵ then PdCl₂ in presence of sodium acetate in DMF at 150 °C¹¹⁷ and recently iodine in refluxing pyridine.¹¹⁸ These oxidative cyclization methods have some limitations including low product yields, utilization of expensive reagent and limited substrate scope. Hence we thought of exploring some potential reagents for this oxidative cyclization.



1) Pb(OAc)₄ (1.5 mmol), anhyd. benzene (30 mL), reflux, 30 min.

- 2) DDQ (1 mmol), benzene, reflux, 72 h.
- 3) PdCl₂ (1 mmol), NaOAc (13.6 mmol), DMF (10 mL), 150 °C, 24 h.
- 4) I₂ (1 mmol), anhyd. Py (10 mL), reflux, 15 h.

Scheme 73: Reported syntheses of coursetans 1 via oxidative cyclization from 52.

$\begin{array}{c} \begin{array}{c} & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & $				
Sr.	Reaction condition	Time	Temperature	Yield
No.		(h)	(°C)	(%)
1)	Fe_3O_4 (2 equiv), 1,2-dichloroethane	24	rt	0
2)	Fe ₃ O ₄ (2 equiv) 1,2-dichloroethane	24	reflux	0
3)	I ₂ (1 equiv), py	24	110	0
4)	I ₂ (0.3 equiv), DMSO	24	150	0
5)	CuBr ₂ (2.2 equiv), 1,2-dichloroethane	24	rt	0
6)	FeCl ₃ .6H ₂ O (2.5 equiv)	24	150	0
7)	FeCl ₃ (2.5 equiv), TFA	24	reflux	0
8)	FeCl ₃ (10 equiv), 1,2-	24	reflux	20
	dichloroethane			

For our studies, 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** was selected as a model substrate. It was subjected to various reaction conditions as shown in below table 11. In the beginning **52a** was subjected to excess of Fe_3O_4 in DCE solvent. However the reaction failed to provide any product at rt as well as under reflux condition (entries 1-2). Then we changed the reagent to iodine in presence of pyridine (entry 3) but no product formation was seen. Also, a reaction using catalytic amount of iodine in DMSO solvent was attempted (entry 4). $CuBr_2$ also did not provide any product (entry 5). As discussed before, a trace amount of product formation was seen when $FeCl_3$ was employed, we further screened $FeCl_3$ reagent using substrate **52a**. $FeCl_3.6H_2O$ and $FeCl_3$ in TFA failed to show any product formation (entries 6-7) but when the reaction was carried out using large excess of $FeCl_3$ in DCE, 20 %

of the desired coumestan 1a was isolated (entry 8). The product 1a showed band in its IR spectrum at 1732 cm⁻¹ accounting for C=O group and its structure was confirmed by the spectral data given below.

Spectral data of 6*H*-benzofuro[3,2-*c*]chromen-6-one (1a)



colorless solid; m.p. 186-188 °C; lit.¹⁰⁶ 187-188 °C.

IR (KBr): $\tilde{v} = 3078, 3045, 1732$ (C=O), 1498, 1082, 752 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ 7.33-7.45 (m, 4H), 7.52-7.56 (m, 1H), 7.59-7.61 (m, 1H), 7.96 (dd, *J* = 8.0, 1.6 Hz, 1H), 8.06-8.08 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 105.9 (Cq), 111.8 (CH), 112.6 (Cq), 117.5 (CH), 121.8 (CH), 121.9 (CH), 123.4 (Cq), 124.7 (CH), 125.2 (CH), 126.8 (CH), 131.9 (CH), 153.6 (Cq), 155.5 (Cq), 158.1 (Cq), 159.9 (Cq).

Inspired with the product formation, however in low yield, we next went on to standardize the amount of FeCl₃ to improve the yield.

	FeCl ₃	
	1,2-Dichloroethane	0
52a 10	24 h	1a 🖳

Sr. No. FeCl₃ (equiv) 230-400 silica gel **Temperature** (°C) Yield (%) 0.3 reflux 1) -----2) 0.3 1:1 wt./wt. FeCl₃ reflux 3) 2.5 reflux 2.5 4) 1:1 wt./wt. FeCl₃ reflux 5 reflux 5) _____ 8 6) reflux ____ 7) 10 rt ____ reflux 8) 10 ----- $9)^{a}$ 3 120 -----

^a Neat reaction

Table 12: Standardization of FeCl₃.

0

0

0

0

0

0

0

20

11

Since the product formation was observed with 3 equiv of FeCl₃ under neat condition, the reaction was further examined using various conditions (Table 12). We started with catalytic amount of FeCl₃ but no product formation was observed (entry 1). Recently a report had claimed that FeCl₃ in presence of 230-400 mesh sized silica gel gave better results than FeCl₃ used alone for coumestan synthesis from 4-hydroxy-3-phenylcoumarins.¹¹² With this fact we tried next reaction with catalytic amount of FeCl₃ along with 230-400 silica gel (1:1 wt./wt. FeCl₃) but again no product formation was observed (entry 2). Similar results were obtained when the amount of FeCl₃ was increased to 2.5 equiv (entries 3-4). Further increase in amount of FeCl₃ up to 5 and 8 equiv did not help to get any product (entries 5-6). However when 10 equiv of FeCl₃ was employed product formation was not visible when the reaction was carried out at room temperature but at reflux temperature gave 20 % yield of product (entries 7-8). This suggested the requirement of large amount of FeCl₃ for the reaction to occur. Also a neat reaction using 3 equiv of FeCl₃ at 120 °C was tried wherein product was isolated in 11 % yield (entry 9). Since 10 equiv of FeCl₃ is a large amount of FeCl₃.

Table 13: Optimiz	zation of FeCl ₃ with 23	30-400 silica gel uno	der neat condition.
-------------------	-------------------------------------	-----------------------	---------------------



Sr.	FeCl ₃	230-400	Time	Temperature	Yield
No.	(equiv)	silica gel	(h)	(°C)	(%)
1)	2		24	120	0
2)	2.5		24	150	12
3)	3		15	120	11
4)	2.5	1:1 wt./wt.	24	150	19
		FeCl ₃			
5)	2.5,	1:1 wt./wt.	24,	150,	24
	then water	FeCl ₃	24	H ₂ O reflux	
6)	2.5, then sonicated		24	150	33
7)	3, then sonicated	1:1 wt./wt.	24	150	36
		FeCl ₃			
8)	4, then sonicated		24	180	25

9)	4, then sonicated	1:1 wt./wt. FeCl ₃	24	150	42
10)	5, then sonicated	1:1 wt./wt. FeCl ₃	24	150	39

In search of an ideal condition, we went for neat conditions (Table 13). At the outset 2, 2.5 and 3 equiv of $FeCl_3$ was tried at different temperatures but the product was isolated in low yields in only the latter 2 cases (entries 1-3). Further 2.5 equiv in presence of 230-400 silica gel gave slightly increased product yield of 19 % (entry 4). Next, the reaction was carried out with 2.5 equiv of FeCl₃ for 24 h followed by the addition of water and refluxing for 24 more hours, however only 24 % of product could be isolated (entry 5). Since the product was not getting isolated in appreciable yield, next we thought to sonicate the reaction mixture after neat heating. Accordingly, a neat reaction was carried out in 2.5 equiv of FeCl₃ in absence of silica gel for 24 and later sonicated after addition of ethyl acetate. This resulted in slight increased product yield of 33 % (entry 6). Similarly, 3 equiv of FeCl₃ in presence of silica gel following similar procedure delivered 36 % yield of coumestan (entry 7). Further increasing FeCl₃ amount to 4 equiv in absence of silica gel at 180 °C delivered 25 % yield (entry 8) whereas when the reaction was carried out in presence of silica gel at lower temperature (150 °C) gave maximum yield of 42 % (entry 9). On using higher amount of FeCl₃ (5 equiv) in presence of silica gel at same temperature resulted in lowered product yield (entry 10). In spite of carrying out various reactions, improvement in the yield of the desired coumestan

was unsuccessful. So, next we tried screening some other common reagents as shown below.

	Reaction condition	
но		
52a		1a

Table 14:	Various	reaction	conditions	from 52a .	
-----------	---------	----------	------------	-------------------	--

Sr.	Reaction condition	Time	Temperature	Yield
No.		(h)	(° C)	(%)
1)	10 wt % Pd/C, 1,4-dioxane	12	reflux	0
2)	10 wt % Pd/C, 0.1 equiv	16	reflux	0
	tetrabutylammonium iodide, 1,4-			
	dioxane			

3)	10 wt % Pd/C, 1 equiv	5	reflux	0
	tetrabutylammonium iodide, 1,4-			
	dioxane			
4)	10 wt % Pd/C, 1 equiv	5	reflux	0
	tetrabutylammonium iodide, 1			
	equiv K ₂ CO ₃ , 1,4-dioxane			
5)	$0.3 \text{ equiv Cu(OTf)}_2,$	24	rt	0
	dichloromethane			
6)	$0.3 \text{ equiv Cu(OTf)}_2,$	24	reflux	0
	dichloromethane			
7)	$0.3 \text{ equiv Cu(OTf)}_2,$	24	reflux	0
	dichloroethane			

Table 14 shows various reaction conditions examined for coumestan formation from **52a**. In the beginning 10 wt % Pd/C was tested in 1,4-dioxane under reflux condition, however no product formation was observed (entry 1). So Pd/C was used in addition with tetrabutylammonium iodide in catalytic as well as stoichiometric amount in 1,4-dioxane under reflux condition but could not give any product (entries 2-3). Above reaction was then attempted in presence of mild base K₂CO₃ but again no product was formed (entry 4). Next we moved on to use copper triflate. Catalytic amount of Cu(OTf)₂ in dichloromethane as well as dichloroethane proved to be ineffective (entries 5-7).

Copper salts have been widely used in organic reactions owing to its cheap availibility and low toxicity. Several reviews have appeared on its role either as reagent and/or catalyst.¹⁷⁸ In particular Cu(OAc)₂ is a mild reagent/catalyst known for the synthesis of several heterocycles.¹⁷⁹ It has gained considerable attention for its role in the intamolecular C-O cyclization *via* C-H functionalization^{179c-d} for the construction of heterocyclic compounds, however the area is not fully explored.

From the above literature discussion the effective role of $Cu(OAc)_2$ in the C-O cyclization of some compounds is clearly seen. However its use in the oxidative cyclization is limited and needs to be explored further. Also it is clear that $Cu(OAc)_2$ has not been employed for coumestan synthesis. With regard to this we envisioned the role of $Cu(OAc)_2$ in the C-O cyclization of 3-(2-hydroxyphenyl)coumarin **52** which may lead to important biologically active coumestan **1** compounds.

	Į	52a HO solvent	-> 1a		
Sr.	Cu(OAc) ₂ .H ₂ O	Solvent	Time	Temperature	Yield
No.	(equiv)		(h)	(°C)	(%)
1)	0.5	dichloromethane	24	rt	0
2)	0.5	dichloromethane	24	reflux	0
3)	0.5	dichloroethane	24	reflux	0
4)	0.2	acetic acid	24	reflux	0
5)	1	acetic acid	24	reflux	0
6)	1	diphenyl ether	5	reflux	55
7) ^a	1	diphenyl ether	6	reflux	mixture of
					products
8) ^b	1	diphenyl ether	6	reflux	76

Table 15: Screening of Cu(OAc)₂ in the coumestan formation.

^a 2 equiv K₂CO₃ was used.

^b Anhyd. Cu(OAc)₂ was used.

In order to check the feasibility of this reaction 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** was treated with 0.5 equiv of Cu(OAc)₂.H₂O. The reactions carried out in dichloromethane at rt as well as reflux condition failed to provide any product (Table 15, entries 1-2). Similarly changing the solvent to dichloroethane did not prove to be effective (entry 3). Attempts to carry out the reaction in acetic acid in presence of catalytic as well as equivalent amounts of Cu(OAc)₂.H₂O failed to provide any product (entries 4-5). The solvent was then changed to diphenyl ether. When the reaction was carried out in this high boiling solvent we were delighted to see the coumestan **1a** formation which occurred within 5 h in 55 % yield (entry 6). To see the effect of base on this reaction, a reaction was attempted in presence of K₂CO₃ but it ended in mixture of products (entry 7). Finally, anhyd. Cu(OAc)₂ proved to be efficient reagent for this oxidative cyclization (entry 8).

Table 16: Screening of various reagents in the coumestan formation.



Sr. No.	Reagent	Time (h)	Yield (%)
1)	Cu(OAc) ₂ .H ₂ O	5	55
2)	Cu(OAc) ₂	6	76
3) ^a	Cu(OAc) ₂ :Zn(OTf) ₂	24	7
4)	CuCl ₂	24	69
5)	CuBr ₂	24	48
6)	CuI	24	19
7)	Cu ₂ O	24	75
8)	CuO	16	60
9) ^b	Cu(OTf) ₂	24	40
10)	Cu(OTf) ₂	16	63
11)	Cu (nanopowder)	24	74
12)	Cu (metal powder)	24	72
13)	Ag.OAc	2	complex mixture
14) ^c	Ag.OAc	24	57
15)	Mn(OAc) ₃ .2H ₂ O	14	18
16)	Zn(OAc) ₂ .2H ₂ O	24	30
17)	ZnCl ₂	24	15
18)	ZnO	24	30
19)	MgCl ₂	24	41
20)	TiO ₂	24	32
21)	Fe ₃ O ₄ (nanopowder)	15	68
22)	Pd(OAc) ₂	18	86
23) ^d	Pd(OAc) ₂	24	RI
24) ^d	PdCl ₂	24	RI
25) ^e	10 % Pd/C	12	RI
26) ^f	10 % Pd/C	12	RI
27) ^g	10 % Pd/C	12	RI
28) ^h	10 % Pd/C	12	78
29)		6	00

^a Toluene:DMSO (20:1) was used as solvent. ^b *p*-xylene solvent was used. ^c 0.5 equiv of reagent was used. ^d 0.1 equiv of reagent was used. ^{e,f,g,h} 10, 20, 30, 50 wt % of reagent was used respectively. RI: Reaction Incomplete.

Encouraged with the coursestan formation using easily available inexpensive Cu reagents (Table 16, entries 1-2) we went ahead with the screening of other copper and various metal reagents (entries 3-28).

Recently, Hong and co-workers^{179c} have demonstrated the synthesis of 6H-benzofuro[2,3c]chromen-6-one using a combination of 1.2 equiv Cu(OAc)₂ and 0.2 equiv of Zn(OTf)₂ in toluene:DMSO (20:1) *via* C-H functionalization/C-O cyclization. Interestingly, it was not explored for the synthesis of isomeric 6H-benzofuro[3,2-c]chromen-6-one (coumestans) *via* insertion of oxygen into aromatic rings. Hence we tried to see coumestan formation using Cu(OAc)₂ and Zn(OTf)₂ combination. However, we could isolate only 7 % coumestan product using this reaction condition (entry 3).

Employing CuCl₂ and CuBr₂ diminished the product yield (entries 4-5). Other Cu reagents such as CuI, Cu₂O and CuO also showed the formation of product among which Cu₂O gave highest yield of 75 % (entries 6-8). However Cu(OTf)₂ was reactive enough to give 40 % yield in refluxing *p*-xylene (entry 9) but in diphenyl ether the yield was increased to 63 % (entry 10). Employing Cu nanopowder and metal powder also proved effective for this cyclization (entries 11-12).

Ag.OAc was found to give a complex mixture when used as a reagent (entry 13). However, reducing its amount by half resulted in 57 % of coumestan formation (entry 14). Mn(OAc)₃.2H₂O was ineffective under above reaction condition and provided coumestan in only 18 % due to poor conversion (entry 15). Zinc metal compounds such as $Zn(OAc)_2.2H_2O$, ZnCl₂ and ZnO were less effective for the cyclization reaction providing low yields of product (entries 16-18). Similarly MgCl₂, TiO₂ and Fe₃O₄ (nanopowder) were also less effective (entries 19-21). Application of $Pd(OAc)_2$ under the present reaction condition afforded maximum yield of 86 % (entry 22). On using catalytic amount of Pd(OAc)₂ and $PdCl_2$ reaction was found to be incomplete even after refluxing for 24 h (entries 23-24). Similarly we tried 10, 20 & 30 wt % of 10 % Pd/C which also resulted in incomplete reaction (entries 25-27). Further, when the amount was increased to 50 wt %, 78 % of coursestan was isolated (entry 28). However due to stoichiometric requirement we did not continued with expensive Pd reagents further. Among all the above reagents, anhyd. Cu(OAc)₂ was concluded to be more efficient and best source for the present oxidative cyclization with respect to cost, availability and reaction time giving yield of 76 % (entry 2). It must be noted that the absence of any reagent did not show any product formation (entry 29).

Table	17:	Solvent	screening.
		~~~~	Seree and

		DAc) ₂ (1 equiv)		
Sr. No.	Solvent	Temperature (°C)	Time (h)	Yield (%)
1)	Toluene	110	24	7
2)	AcOH	118	24	0
3)	<i>p</i> -Xylene	138	24	20
4)	DMF	153	24	15
5)	DMA	165	24	6
6)	o-DCB	180	24	30
7)	DMSO	189	24	ND
8)	Diphenyl ether	258	6	76

ND: Not Determined

To check the feasibility of this reaction in other solvents, solvent screening was done (Table 17). At the outset a reaction with anhyd.  $Cu(OAc)_2$  (1 equiv) in toluene under refluxing condition for 24 h delivered product albeit in poor yield of 7 % (entry 1). Acetic acid did not show any product formation even after 24 h (entry 2). Examining different solvents with varying boiling points such as, *p*-xylene, DMF, DMA and *o*-DCB proved to be inefficient to provide desired product in good yields (entries 3-6). Reaction in DMSO resulted in charring and no coumestan product was isolated (entry 7). Among these solvents, diphenyl ether proved to be the optimum solvent as substantial product yield of 76 % was isolated (entry 8).

Table 1	18:	Standardization	of reaction	temperature.
---------	-----	-----------------	-------------	--------------

	O     O     Cu(OAc) ₂ (1 equiv) Diphenyl ether       Temperature     Temperature		
Sr. No.	Temperature (°C)	Time (h)	Yield (%)
1)	100	24	25
2)	150	24	50
3)	170-180	12	62
4)	200	10	64
5)	258	6	76

Temperature studies were done by carrying out the reactions at different temperatures starting from 100 °C to reflux temperature in diphenyl ether solvent (Table 18). It was observed that at 100 °C only 25 % yield was isolated (entry 1). Increasing the reaction temperature to 150 °C allowed increase in product yield to 50 % (entry 2). Further increase up to 62 % was observed when the reaction temperature was maintained between 170-180 °C (entry 3). At 200 °C no much increase in product yield was observed (entry 4). However maximum yield of 76 % was observed at 258 °C (entry 5). Hence refluxing temperature was found to be the optimum temperature as lowering in temperature showed decreased product yield.

	HO 52a HO Cu(OA Diphenyl eth	Ac) ₂ her, reflux	
Sr. No.	Cu(OAc) ₂ (equiv)	Time (h)	Yield (%)
1)	0.2	24	66
2)	0.5	12	70
3)	1	6	76
4)	1.2	11	70
5)	2	14	56

Table 19: Standardization of Cu(OAc)₂ concentration.

Standardization of  $Cu(OAc)_2$  concentration was done by carrying out different reactions with varying amount of  $Cu(OAc)_2$  from catalytic (0.2 equiv) to excess (2 equiv) (Table 19). It was endearing to see that even catalytic amount of  $Cu(OAc)_2$  was effective enough to provide product yield of 66 % (entry 1). Similarly 0.5 equiv of  $Cu(OAc)_2$  worked well to deliver increased yield of 70 % (entry 2). Best quantity was found to be 1 equiv of  $Cu(OAc)_2$  giving highest yield of 76 % in just 6 h (entry 3). Slight increase in reagent quantity resulted in yield drop (entry 4). Excess of the reagent caused further drop to 56 % (entry 5).

Table 20: Synthesis of coursetans using optimized reaction condition.



Sr.	Substituted 3-(2-	Coumestan	Time	Yield
No.	hydroxyphenyl)-2H-		( <b>h</b> )	(%)
	chromen-2-one			
1)	HO		6	76
	52a	<b>1</b> a		
2)	MeO O O HO	MeO O O	13	66
	52b	1b		
3)	MeO HO	MeO O O	6	72
	52c	1c		
4)	HO		13	70
	52d	1d		
5)	HO		8	80
	52e	1e		
6)	HO		10	65
	52f	1f		
7)	HO		17	67
	52g	1g		
8)	MeO HO	OMe MeO O O	6	80
	52h	1h		

	MeO	MeO		
9)	MeO	MeO	8	77
,	но	0-		
	52i	1i		
10)	0	0	14	61
	HO			
	52j	1j		
	OH	OH O O		
11)			16	62
	НО	0-		
	52k	1k		
	HO	HO		
12)			14	68
	HO	0-()		
	521	11		
			24	30
13)	Br	Br		
	HO ~		$18^a$	53 ^a
	52111	Im		
14)	CI	CI O	24	67
	52n	1n		
	0,00			
15)	O-N	O-N	13	54
15)	НО	0	15	54
	520	10		
16)			5	70
	HOOMe			
	52p	1p		
	MeO V O V	MeO		
17)			5	65
	HOHOME	0-		_
	52q	OMe <b>7</b> £		



^a Reaction was carried out in 1.5 equiv of Cu(OAc)₂.

After obtaining the optimum reaction conditions further studies to check the substrate scope were undertaken (Table 20). Substituents on both the phenyl rings A and B were evaluated. Parent coumestan was obtained in 76 % yield. Study on ring A revealed that the electron releasing methoxy and ethoxy substituents reacted smoothly to give the desired products **1b**-**1d** in good yields. Naphthol group was very reactive to provide the expected coumestan **1e** in 80 % yield. Monomethyl and dimethyl substituents were also successfully converted into the desired products **1f** and **1g**. Coumestans **1h-1j** were formed when dimethoxy and methylenedioxy substitutions were examined on ring A. Hydroxy substituents also reacted to produce the required coumestans **1k** and **1l** without any protection thus exhibiting good efficiency and practicability of this method. Coumestan bearing electron withdrawing bromo group **1m** was isolated in low yield when 1 equiv of  $Cu(OAc)_2$  was employed. However when the amount of  $Cu(OAc)_2$  was increased to 1.5 equiv the product yield also increased to 53 %. Similarly electron deficient coumestan with chloro substituent **1n** was synthesised in 67 % yield. Strong electron withdrawing nitro group was quite reactive enough to offer the desired product **1o**.

On successfully synthesising above derivatives, we went on to explore the substitution pattern on ring B. When methoxy substituent was employed on ring B without any substituent on ring A, reaction went on smoothly to afford coumestan **1p** in 70 % yield. Similarly various substituents on ring A provided diverse coumestans **2f**, **1q-1t** in moderate to good yields in presence of methoxy substituent on ring B. Among these the dimethyl ether of natural compounds coumestrol **2f**⁹ and sativol **1s** were isolated. Moreover trimethyl ether of lucernol **1t** was also successfully formed. Various reports exhibiting the conversion of compound **2f** to naturally occurring coumestrol **2c** have been demonstrated.^{111b,132} With the introduction of methyl group on ring B also successfully delivered coumestans **1u** and **1v** in 65 and 60 % yields respectively without affecting the side chain.

Encouraged by the formation of hydroxyl coumestans **1k** and **1l** we thought to apply this methodology towards the protective group free synthesis of naturally occurring 4'-O-methylcoumestrol/9-methoxycoumestrol **2e**⁸ and coumestrol **2c**.^{6,8a-c,14a,23a,26a-b,31a,33a-c,34,49} Several syntheses of these coumestans have been reported but an efficient method without any protection-deprotection strategy is still in high demand. On subjecting the necessary starting materials to the above reaction condition it was endearing to see the formation of both coumestans **2e** and **2c** in 59 and 55 % yields respectively thus eliminating the need of protection-deprotection steps as reported in literature methods. As most of the naturally
occurring coumestans contain hydroxyl and/or methoxy group/s, our methodology provides a broad scope for synthesis of more natural members of coumestan family.

#### Spectral data of all compounds

#### 3-(2-Hydroxyphenyl)-2H-chromen-2-one (52a)



yield (0.869 g, 98 %); colorless solid; m.p. 212-214 °C; lit.¹⁷⁶ 212-213 °C.

**IR (KBr):**  $\tilde{v} = 3340$  (OH), 3062, 1697 (C=O), 1604, 1350, 750 cm⁻¹.

¹**H NMR** (**DMSO**-*d*₆, **400 MHz**): δ 6.84-6.90 (m, 2H), 7.21-7.26 (m, 2H), 7.35 (td, *J* = 7.6, 0.8 Hz, 1H), 7.41 (d, *J* = 8.4 Hz, 1H), 7.61 (td, *J* = 8.4, 1.6 Hz, 1H), 7.73 (dd, *J* = 7.6, 1.2 Hz, 1H), 7.98 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 115.6 (CH), 115.8 (CH), 118.8 (CH), 119.2 (Cq), 122.1 (Cq), 124.6 (CH), 125.9 (Cq), 128.3 (CH), 129.8 (CH), 130.7 (CH), 131.6 (CH), 141.9 (CH), 152.9 (Cq), 154.9 (Cq), 159.5 (Cq).

#### 6H-Benzofuro[3,2-c]chromen-6-one (1a)



yield (0.075 g, 76 %); colorless solid; m.p. 186-188 °C; lit.¹⁰⁶ 187-188 °C.

**IR** (**KBr**):  $\tilde{v} = 3078$ , 3045, 1732 (C=O), 1498, 1082, 752 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 7.33-7.45 (m, 4H), 7.52-7.56 (m, 1H), 7.59-7.61 (m, 1H), 7.96 (dd, *J* = 8.0, 1.6 Hz, 1H), 8.06-8.08 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 105.9 (Cq), 111.8 (CH), 112.6 (Cq), 117.5 (CH), 121.8 (CH), 121.9 (CH), 123.4 (Cq), 124.7 (CH), 125.2 (CH), 126.8 (CH), 131.9 (CH), 153.6 (Cq), 155.5 (Cq), 158.1 (Cq), 159.9 (Cq).

## 3-(2-Hydroxyphenyl)-7-methoxy-2H-chromen-2-one (52b)



yield (0.950 g, 95 %); colorless solid; m.p. 160-162 °C. **Rf:** 0.56 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3346$  (OH), 1685 (C=O), 1622, 1508, 1280, 738 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 3.87 (s, 3H), 6.86 (td, *J* = 7.6, 1.2 Hz, 1H), 6.92 (dd, *J* = 8.0, 0.8 Hz, 1H), 6.97 (dd, *J* = 8.4, 2.4 Hz, 1H), 7.03 (d, *J* = 2.4 Hz, 1H), 7.22 (ddd, *J* = 8.0, 7.6, 1.6 Hz, 1H), 7.26 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.65 (d, *J* = 8.8 Hz, 1H), 7.96 (s, 1H), 9.57 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.9 (OCH₃), 100.3 (CH), 112.3 (CH), 112.8 (Cq), 115.6 (CH), 118.7 (CH), 122.2 (Cq), 122.4 (Cq), 129.3 (CH), 129.4 (CH), 130.8 (CH), 142.1 (CH), 154.8 (Cq), 155.0 (Cq), 159.6 (Cq), 162.1 (Cq).

**HRMS (ESI):** for  $C_{16}H_{12}O_4Na: m/z [M + Na]^+$ , calcd: 291.0633, found: 291.0632.

## <u>3-Methoxy-6H-benzofuro[3,2-c]chromen-6-one (1b)</u>



yield (0.065 g, 66 %); colorless solid; m.p. 196-198 °C; lit.¹⁸⁰ 195-197 °C.

**IR (KBr):**  $\tilde{v} = 2985$ , 1735 (C=O), 1627, 1274, 754 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.85 (s, 3H), 6.91-6.93 (m, 2H), 7.35-7.39 (m, 2H), 7.55-7.59 (m, 1H), 7.86 (d, *J* = 9.2 Hz, 1H), 8.01-8.05 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.8 (OCH₃), 101.4 (CH), 103.2 (Cq), 105.7 (Cq), 111.5 (CH), 112.9 (CH), 121.4 (CH), 122.8 (CH), 123.6 (Cq), 125.0 (CH), 126.1 (CH), 155.2 (Cq), 155.5 (Cq), 158.2 (Cq), 160.5 (Cq), 162.9 (Cq).

# <u>3-(2-Hydroxyphenyl)-6-methoxy-2H-chromen-2-one (52c)</u>



yield (0.960 g, 96 %); colorless solid; m.p. 144-146 °C; lit.¹¹⁷ 144-145 °C.

**IR (KBr):**  $\tilde{v} = 3327$  (OH), 1689 (C=O), 1577, 1487, 1031, 750 cm⁻¹.

¹**H NMR** (**DMSO**-*d*₆, **400 MHz**): δ 3.82 (s, 3H), 6.86 (t, *J* = 7.6 Hz, 1H), 6.92 (d, *J* = 8.0 Hz, 1H), 7.19-7.28 (m, 3H), 7.31 (d, *J* = 2.8 Hz, 1H) 7.38 (d, *J* = 8.8 Hz, 1H), 7.98 (s, 1H), 9.62 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.7 (OCH₃), 110.5 (CH), 115.7 (CH), 116.9 (CH), 118.7 (CH), 118.9 (CH), 119.7 (Cq), 122.2 (Cq), 126.2 (Cq), 129.7 (CH), 130.8 (CH), 141.7 (CH), 147.4 (Cq), 155.0 (Cq), 155.6 (Cq), 159.5 (Cq).

#### 2-Methoxy-6H-benzofuro[3,2-c]chromen-6-one (1c)



yield (0.071 g, 72 %); colorless solid; m.p. 156-158 °C; lit.^{111a} 155-157 °C.

**IR** (**KBr**):  $\tilde{v} = 3089, 2941, 1735$  (C=O), 1566, 1074, 777 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.83 (s, 3H), 7.03 (dd, *J* = 9.2, 2.8 Hz, 1H), 7.25-7.29 (m, 2H), 7.32-7.39 (m, 2H), 7.54 (dd, *J* = 6.8, 2.0 Hz, 1H), 8.00-8.02 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.9 (OCH₃), 103.2 (CH), 105.9 (Cq), 111.6 (CH), 112.7 (Cq), 118.6 (CH), 120.2 (CH), 121.8 (CH), 123.4 (Cq), 125.2 (CH), 126.7 (CH), 148.1 (Cq), 155.4 (Cq), 156.3 (Cq), 158.2 (Cq), 159.8 (Cq).

## 8-Ethoxy-3-(2-hydroxyphenyl)-2H-chromen-2-one (52d)



yield (0.988 g, 94 %); colorless solid; m.p. 170-172 °C.

**Rf:** 0.48 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3300$  (OH), 1695 (C=O), 1469, 1278, 1111, 779 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 1.42 (t, *J* = 7.2 Hz, 3H), 4.19 (q, *J* = 14.0, 7.2 Hz, 2H), 6.87 (td, *J* = 7.6, 1.2 Hz, 1H), 6.91 (d, *J* = 8.0 Hz, 1H), 7.21-7.28 (m, 5H), 8.01 (s, 1H) 9.60 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 14.6 (CH₃), 64.3 (OCH₂), 114.6 (CH), 115.6 (CH), 118.7 (CH), 119.5 (CH), 119.9 (Cq), 122.1 (Cq), 124.4 (CH), 126.0 (Cq), 129.7 (CH), 130.7 (CH), 142.1 (CH), 142.4 (Cq), 145.5 (Cq), 155.0 (Cq), 159.1 (Cq).

**HRMS (ESI):** for  $C_{17}H_{14}O_4Na: m/z [M + Na]^+$ , calcd: 305.0790, found: 305.0790.

# <u>4-Ethoxy-6H-benzofuro[3,2-c]chromen-6-one (1d)</u>



yield (0.069 g, 70 %); colorless solid; m.p. 172-174 °C.

**Rf:** 0.52 (30 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3084$ , 2972, 1722 (C=O), 1566, 1284, 1058, 746 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz):  $\delta$  1.48 (t, *J* = 7.2 Hz, 3H), 4.17 (q, *J* = 14.0, 6.8 Hz, 2H), 7.06 (d, *J* = 8.0 Hz, 1H), 7.24 (t, *J* = 8.0 Hz, 1H), 7.36-7.43 (m, 2H), 7.52 (d, *J* = 8.0 Hz, 1H), 7.59 (dd, *J* = 6.8, 2.4 Hz, 1H), 8.08-8.10 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 14.8 (CH₃), 65.1 (CH₂), 105.9 (Cq), 111.7 (CH), 113.1 (CH), 113.4 (Cq), 114.9 (CH), 121.9 (CH), 123.5 (Cq), 124.7 (CH), 125.2 (CH), 126.7 (CH), 143.6 (Cq), 147.1 (Cq), 155.5 (Cq), 157.7 (Cq), 160.2 (Cq).

**HRMS (ESI):** for  $C_{17}H_{12}O_4H$ : m/z  $[M + H]^+$ , calcd: 281.0814, found: 281.0814.

#### 2-(2-Hydroxyphenyl)-3H-benzo[f]chromen-3-one (52e)



yield (0.990 g, 92 %); colorless solid; m.p. 216-218 °C; lit.¹⁸¹ 216-217 °C.

**IR (KBr):**  $\tilde{v} = 3282$  (OH), 1707 (C=O), 1454, 1355, 808, 734 cm⁻¹.

¹**H** NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  6.91 (td, J = 7.6, 1.2 Hz, 1H), 6.95 (d, J = 8.0 Hz, 1H), 7.25-7.29 (m, 1H), 7.38 (dd, J = 7.6, 1.6 Hz, 1H), 7.61-7.64 (m, 2H), 7.71 (t, J = 7.2 Hz, 1H), 8.08 (d, J = 8.0 Hz, 1H), 8.20 (d, J = 8.8 Hz, 1H), 8.61 (d, J = 8.8 Hz, 1H), 8.86 (s, 1H), 9.65 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 113.2 (Cq), 115.7 (CH), 116.5 (CH), 118.7 (CH), 122.5 (CH), 122.6 (Cq), 125.5 (Cq), 125.9 (CH), 128.2 (CH), 128.8 (CH), 128.8 (Cq), 129.7 (CH), 129.9 (Cq), 131.1 (CH), 132.6 (CH), 138.0 (CH), 152.6 (Cq), 155.2 (Cq), 159.3 (Cq).

## 8H-Benzo[f]benzofuro[3,2-c]chromen-8-one (1e)



yield (0.079 g, 80 %); colorless solid; m.p. 210-212 °C.

Rf: 0.5 (20 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3066$ , 1741 (C=O), 1560, 1008, 746 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ 7.43-7.50 (m, 2H), 7.56-7.60 (m, 2H), 7.73-7.78 (m, 2H),
7.91 (d, *J* = 8.4 Hz, 1H), 7.99 (d, *J* = 8.8 Hz, 1H), 8.15-8.18 (m, 1H), 9.21 (d, *J* = 7.6 Hz, 1H).
¹³C NMR (CDCl₃, 100 MHz): δ 105.9 (Cq), 106.8 (Cq), 111.6 (CH), 117.2 (CH), 121.7 (CH), 122.7 (Cq), 125.3 (2 X CH), 126.3 (CH), 126.7 (CH), 127.2 (Cq), 128.7 (2 X CH),
130.2 (Cq), 132.9 (CH), 153.7 (Cq), 155.3 (Cq), 157.9 (Cq), 161.1 (Cq).

**HRMS (ESI):** for  $C_{19}H_{10}O_3H$ : m/z  $[M + H]^+$ , calcd: 287.0708, found: 287.0708.

#### 3-(2-Hydroxyphenyl)-6-methyl-2H-chromen-2-one (52f)



yield (0.845 g, 90 %); colorless solid; m.p. 166-168 °C; lit.¹⁸² 167-168 °C.

**IR (KBr):**  $\tilde{v} = 3398$  (OH), 3068, 1710 (C=O), 1452, 1228, 742 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 2.38 (s, 3H), 6.86 (t, *J* = 7.6 Hz, 1H), 6.91 (d, *J* = 8.4 Hz, 1H), 7.21-7.28 (m, 2H), 7.33 (d, *J* = 8.4 Hz, 1H), 7.43 (d, *J* = 8.4 Hz, 1H), 7.53 (s, 1H), 7.96 (s, 1H), 9.61 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 20.2 (CH₃), 115.6 (2 X CH), 118.7 (CH), 119.0 (Cq), 122.3 (Cq), 125.8 (Cq), 127.9 (CH), 129.6 (CH), 130.8 (CH), 132.3 (CH), 133.6 (Cq), 141.8 (CH), 151.2 (Cq), 155.0 (Cq), 159.5 (Cq).

## 2-Methyl-6H-benzofuro[3,2-c]chromen-6-one (1f)



yield (0.064 g, 65 %); colorless solid; m.p. 156-158 °C; lit.^{111a} 157-158 °C.

**IR (KBr):**  $\tilde{v} = 2924$ , 1730 (C=O), 1570, 1444, 748 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz):  $\delta$  2.52 (s, 3H), 7.44 (s, 2H), 7.47-7.53 (m, 2H), 7.69 (dd, J = 6.8, 3.2 Hz, 1H), 7.86 (s, 1H), 8.16-8.18 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 20.9 (CH₃), 105.7 (Cq), 111.7 (CH), 112.2 (Cq), 117.2 (CH), 121.5 (CH), 121.8 (CH), 123.5 (Cq), 125.1 (CH), 126.6 (CH), 133.0 (CH), 134.6 (Cq), 151.8 (Cq), 155.4 (Cq), 158.3 (Cq), 159.9 (Cq).

## 3-(2-Hydroxyphenyl)-5,7-dimethyl-2*H*-chromen-2-one (52g)



yield (0.850 g, 86 %); colorless solid; m.p. 166-168 °C.

**Rf:** 0.63 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3348$  (OH), 3016, 1728 (C=O), 1448, 1232, 846 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 2.36 (s, 3H), 2.45 (s, 3H), 6.84-6.89 (m, 2H), 7.02 (s, 1H), 7.06 (s, 1H), 7.22 (td, *J* = 7.6, 1.6 Hz, 1H), 7.27 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.99 (s, 1H), 9.70 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 17.8 (CH₃), 21.1 (CH₃), 113.9 (CH), 115.4 (Cq), 115.5 (CH), 118.8 (CH), 122.6 (Cq), 124.1 (Cq), 126.8 (CH), 129.6 (CH), 130.9 (CH), 136.1 (Cq), 138.9 (CH), 141.9 (Cq), 153.6 (Cq), 154.9 (Cq), 159.5 (Cq).

**HRMS (ESI):** for  $C_{17}H_{14}O_3Na: m/z [M + Na]^+$ , calcd: 289.0841, found: 289.0841.

#### 1,3-Dimethyl-6H-benzofuro[3,2-c]chromen-6-one (1g)



yield (0.066 g, 67 %); colorless solid; m.p. 252-254 °C.

Rf: 0.59 (20 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 2924$ , 1737 (C=O), 1606, 1446, 1083, 783 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 2.42 (s, 3H), 2.86 (s, 3H), 6.98 (s, 1H), 7.12 (s, 1H), 7.42-7.47 (m, 2H), 7.62-7.66 (m, 1H), 8.11-8.15 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 21.2 (CH₃), 21.7 (CH₃), 105.0 (Cq), 109.5 (Cq), 111.6 (CH), 115.4 (CH), 121.7 (CH), 123.1 (Cq), 125.1 (CH), 126.2 (CH), 128.0 (CH), 134.9 (Cq), 142.5 (Cq), 154.6 (Cq), 155.3 (Cq), 158.4 (Cq), 161.4 (Cq).

**HRMS (ESI):** for  $C_{17}H_{12}O_3H [M + H]^+$ , calcd: 265.0865, found: 265.0865.

#### <u>3-(2-Hydroxyphenyl)-7,8-dimethoxy-2*H*-chromen-2-one (52h)</u>



yield (0.890 g, 80 %); colorless solid; m.p. 162-164 °C.

**Rf:** 0.42 (50 % ethyl acetate/petroleum ether).

**IR** (**KBr**):  $\tilde{v} = 3286$  (OH), 1689 (C=O), 1604, 1284, 1107, 781 cm⁻¹.

¹**H** NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  3.87 (s, 3H), 3.93 (s, 3H), 6.85 (td, J = 7.6, 1.2 Hz, 1H), 6.91 (dd, J = 8.0, 1.2 Hz, 1H), 7.13 (d, J = 8.8 Hz, 1H), 7.19-7.27 (m, 2H), 7.47 (d, J = 8.8 Hz, 1H), 7.95 (s, 1H), 9.56 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 56.3 (OCH₃), 60.8 (OCH₃), 109.2 (CH), 113.9 (Cq), 115.6 (CH), 118.7 (CH), 122.3 (Cq), 122.6 (Cq), 123.5 (CH), 129.5 (CH), 130.8 (CH), 134.9 (Cq), 142.3 (CH), 146.8 (Cq), 154.8 (Cq), 155.1 (Cq), 159.2 (Cq).

**HRMS (ESI):** for  $C_{17}H_{14}O_5Na: m/z [M + Na]^+$ , calcd: 321.0739, found: 321.0739.

#### 3,4-Dimethoxy-6H-benzofuro[3,2-c]chromen-6-one (1h)



yield (0.079 g, 80 %); colorless solid; m.p. 188-190 °C; lit.¹⁰⁶ 189-190 °C.

**IR (KBr):**  $\tilde{v} = 2947$ , 1743 (C=O), 1292, 1087, 1031, 748 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.94 (s, 3H), 3.99 (s, 3H), 6.96 (d, *J* = 9.2 Hz, 1H), 7.37-7.39 (m, 2H), 7.56-7.58 (m, 1H), 7.67 (d, *J* = 8.8 Hz, 1H), 8.05-8.07 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 56.5 (OCH₃), 61.6 (OCH₃), 103.6 (Cq), 107.2 (Cq), 109.0 (CH), 111.5 (CH), 116.8 (CH), 121.6 (CH), 123.6 (Cq), 125.1 (CH), 126.3 (CH), 136.9 (Cq), 147.8 (Cq), 155.3 (Cq), 155.9 (Cq), 157.8 (Cq), 160.5 (Cq).

## 3-(2-Hydroxyphenyl)-6,7-dimethoxy-2H-chromen-2-one (52i)



yield (0.910 g, 82 %); colorless solid; m.p. 164-166 °C.

Rf: 0.5 (50 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3523$  (OH), 3078, 1703 (C=O), 1571, 1292, 758 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 3.78 (s, 3H), 3.85 (s, 3H), 6.82-6.89 (m, 2H), 7.05 (s, 1H), 7.18-7.23 (m, 2H), 7.24 (s, 1H), 7.87 (s, 1H), 9.75 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.8 (OCH₃), 56.1 (OCH₃), 99.5 (CH), 108.7 (CH), 111.6 (Cq), 115.5 (CH), 118.8 (CH), 122.3 (Cq), 122.4 (Cq), 129.4 (CH), 130.8 (CH), 142.2 (CH), 145.8 (Cq), 148.8 (Cq), 152.1 (Cq), 154.9 (Cq), 160.1 (Cq).

**HRMS (ESI):** for  $C_{17}H_{14}O_5Na: m/z [M + Na]^+$ , calcd: 321.0739, found: 321.0739.

## 2,3-Dimethoxy-6H-benzofuro[3,2-c]chromen-6-one (1i)

MeO
MeO
1i 0

yield (0.059 g, 60 %); colorless solid; m.p. 256-258 °C.

Rf: 0.55 (40 % ethyl acetate/petroleum ether).

**IR** (**KBr**):  $\tilde{v} = 2941$ , 1730 (C=O), 1517, 1273, 775 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 3.93 (s, 3H), 3.97 (s, 3H), 6.96 (s, 1H), 7.32 (s, 1H), 7.37-7.41 (m, 2H), 7.56-7.59 (m, 1H), 8.04-8.06 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 56.5 (2 X OCH₃), 100.7 (CH), 101.9 (CH), 103.7 (Cq), 104.7 (Cq), 111.5 (CH), 121.6 (CH), 123.7 (Cq), 125.1 (CH), 126.2 (CH), 146.9 (Cq), 149.8 (Cq), 153.1 (Cq), 155.3 (Cq), 158.5 (Cq), 160.7 (Cq).

**HRMS (ESI):** for  $C_{17}H_{12}O_5H$ : m/z  $[M + H]^+$ , calcd: 297.0763, found: 297.0763.

## 7-(2-Hydroxyphenyl)-6H-[1,3]dioxolo[4,5-g]chromen-6-one (52j)

52j	но

yield (0.735 g, 70 %); colorless solid; m.p. 242-244 °C.

**Rf:** 0.48 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3388$  (OH), 1683 (C=O), 1492, 1278, 734 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 6.18 (s, 2H), 6.84 (td, *J* = 7.6, 1.2 Hz, 1H), 6.89 (dd, *J* = 7.6, 0.8 Hz, 1H), 7.16 (s, 1H), 7.19-7.26 (m, 3H), 7.91 (s, 1H), 9.56 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 97.6 (CH), 102.4 (CH₂), 105.3 (CH), 112.9 (Cq), 115.6 (CH), 118.6 (CH), 122.3 (Cq), 122.4 (Cq), 129.4 (CH), 130.8 (CH), 142.3 (CH), 144.4 (Cq), 150.0 (Cq), 150.5 (Cq), 154.9 (Cq), 159.6 (Cq).

**HRMS (ESI):** for  $C_{16}H_{10}O_5Na: m/z [M + Na]^+$ , calcd: 305.0426, found: 305.0425.

## 6H-Benzofuro[3,2-c][1,3]dioxolo[4,5-g]chromen-6-one (1j)

1j •

yield (0.069 g, 70 %); colorless solid; m.p. 272-274 °C.

Rf: 0.42 (20 % ethyl acetate/petroleum ether).

**IR** (**KBr**):  $\tilde{v} = 2924$ , 1751 (C=O), 1458, 1261, 773 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 6.06 (s, 2H), 6.92 (s, 1H), 7.29 (s, 1H), 7.37-7.39 (m, 2H), 7.55-7.57 (m, 1H), 8.02-8.04 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 98.1 (CH), 98.4 (CH), 101.5 (CH₂), 102.5 (Cq), 104.9 (Cq), 110.5 (CH), 120.6 (CH), 122.5 (Cq), 124.1 (CH), 125.3 (CH), 144.3 (Cq), 149.9 (Cq), 150.6 (Cq), 154.2 (Cq), 157.3 (Cq), 159.7 (Cq).

**HRMS (ESI):** for  $C_{16}H_8O_5H$ : m/z  $[M + H]^+$ , calcd: 281.0450, found: 281.0450.

#### 8-Hydroxy-3-(2-hydroxyphenyl)-2H-chromen-2-one (52k)



yield (0.715 g, 75 %); colorless solid; m.p. 210-212 °C.

**Rf:** 0.48 (50 % ethyl acetate/petroleum ether).

**IR** (**KBr**):  $\tilde{v} = 3452$  (OH), 3305 (OH), 3047, 1674 (C=O), 1604, 1483, 1184, 758 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 6.87 (td, *J* = 7.6, 1.2 Hz, 1H), 6.92 (dd, *J* = 8.4, 0.8 Hz, 1H), 7.08-7.12 (m, 1H), 7.14-7.19 (m, 2H), 7.24 (ddd, *J* = 8.0, 7.6, 1.6 Hz, 1H), 7.28 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.97 (s, 1H), 9.62 (br s, 1H), 10.23 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 115.6 (CH), 117.7 (CH), 118.3 (CH), 118.7 (CH), 120.2 (Cq), 122.3 (Cq), 124.4 (CH), 125.8 (Cq), 129.6 (CH), 130.8 (CH), 141.7 (Cq), 142.3 (CH), 144.3 (Cq), 155.0 (Cq), 159.3 (Cq).

**HRMS (ESI):** for  $C_{15}H_{10}O_4Na: m/z [M + Na]^+$ , calcd: 277.0477, found: 277.0477.

## <u>4-Hydroxy-6*H*-benzofuro[3,2-*c*]chromen-6-one (1k)</u>



yield (0.061 g, 62 %); colorless solid; m.p. 246-248 °C.

Rf: 0.55 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3230$  (OH), 2924, 1703 (C=O), 1566, 1184, 740 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 7.22 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.33 (t, *J* = 8.0 Hz, 1H), 7.51-7.60 (m, 3H), 7.91 (d, *J* = 8.0 Hz, 1H), 8.01 (dd, *J* = 7.2, 1.6 Hz, 1H), 10.53 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 104.9 (Cq), 111.6 (CH), 112.2 (CH), 112.9 (Cq), 118.7 (CH), 120.8 (CH), 122.9 (Cq), 125.2 (CH), 125.4 (CH), 126.9 (CH), 141.9 (Cq), 145.4 (Cq), 154.9 (Cq), 157.0 (Cq), 160.0 (Cq).

**HRMS (ESI):** for  $C_{15}H_8O_4H$ : m/z  $[M + H]^+$ , calcd: 253.0501, found: 253.0501.

# 7-Hydroxy-3-(2-hydroxyphenyl)-2H-chromen-2-one (52l)



yield (0.736 g, 78 %); colorless solid; m.p. 200-202 °C; lit.¹¹⁷ 201-202 °C. **IR (KBr):**  $\tilde{v} = 3232$  (OH), 2746, 1676 (C=O), 1585, 1228, 756 cm⁻¹. ¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 6.76-6.91 (m, 4H), 7.20 (ddd, *J* = 8.0, 7.2, 1.6 Hz, 1H), 7.24 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.56 (d, *J* = 8.4 Hz, 1H), 7.90 (s, 1H), 9.52 (br s, 1H), 10.56 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 101.8 (CH), 111.7 (Cq), 113.1 (CH), 115.6 (CH), 118.6 (CH), 121.2 (Cq), 122.6 (Cq), 129.3 (CH), 129.6 (CH), 130.9 (CH), 142.4 (CH), 154.9 (Cq), 155.0 (Cq), 159.7 (Cq), 160.9 (Cq).

#### <u>3-Hydroxy-6H-benzofuro[3,2-c]chromen-6-one (11)</u>



yield (0.067 g, 68 %); colorless solid; m.p. 270-272 °C; lit.¹²⁵ 270-272 °C.

**IR (KBr):**  $\tilde{v} = 3317$  (OH), 2924, 1724 (C=O), 1438, 1093, 748 cm⁻¹.

¹H NMR (DMSO-*d*₆, 400 MHz): δ 6.65 (d, J = 1.6 Hz, 1H), 6.75 (dd, J = 8.8, 1.6 Hz, 1H), 7.26-7.33 (m, 2H), 7.51 (d, J = 7.6 Hz, 1H), 7.58 (d, J = 8.8 Hz, 1H), 7.63 (d, J = 6.8 Hz, 1H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 101.7 (Cq), 103.0 (CH), 103.8 (Cq), 111.9 (CH), 113.9 (CH), 120.3 (CH), 123.1 (Cq), 123.3 (CH), 125.2 (CH), 126.2 (CH), 154.5 (Cq), 155.2 (Cq), 157.4 (Cq), 160.6 (Cq), 161.9 (Cq).

## 6-Bromo-3-(2-hydroxyphenyl)-2H-chromen-2-one (52m)



yield (1.090 g, 92 %); pale yellow solid; m.p. 234-236 °C; lit.¹⁷⁶ 235-236 °C.

**IR** (**KBr**):  $\tilde{v} = 3350$  (OH), 3049, 1691 (C=O), 1483, 1197, 754 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 6.88 (td, *J* = 7.6, 1.2 Hz, 1H), 6.92 (d, *J* = 7.6 Hz, 1H), 7.23-7.28 (m, 2H), 7.42 (d, *J* = 8.8 Hz, 1H), 7.77 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.99 (s, 1H), 8.01 (d, *J* = 2.4 Hz, 1H), 9.76 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 115.6 (CH), 116.0 (Cq), 118.2 (CH), 118.8 (CH), 121.1 (Cq), 121.8 (Cq), 127.1 (Cq), 129.9 (CH), 130.3 (CH), 130.7 (CH), 133.8 (CH), 140.6 (CH), 152.0 (Cq), 154.9 (Cq), 158.9 (Cq).

#### 2-Bromo-6H-benzofuro[3,2-c]chromen-6-one (1m)



yield (0.030 g, 30 %; 0.052 g, 53 %); colorless solid; m.p. 242-244 °C.

**Rf:** 0.64 (20 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3074$ , 1749 (C=O), 1550, 1446, 1103, 979, 750 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 7.32 (d, *J* = 8.8 Hz, 1H), 7.39-7.47 (m, 2H), 7.59-7.63 (m, 2H), 8.06-8.08 (m, 1H), 8.09 (d, *J* = 2.4 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 106.6 (Cq), 111.9 (CH), 114.2 (Cq), 117.5 (Cq), 119.2 (CH), 122.0 (CH), 123.2 (Cq), 124.4 (CH), 125.5 (CH), 127.3 (CH), 134.7 (CH), 152.4 (Cq), 155.7 (Cq), 157.4 (Cq), 158.5 (Cq).

**HRMS (ESI):** for  $C_{15}H_7BrO_3Na: m/z [M + Na]^+$ , calcd: 336.9476, found: 336.9477.

## 6-Chloro-3-(2-hydroxyphenyl)-2H-chromen-2-one (52n)



yield (0.880 g, 86 %); pale brown solid; m.p. 230-232 °C; lit.¹⁷⁶ 230-231 °C.

**IR (KBr):**  $\tilde{v} = 3344$  (OH), 3061, 1689 (C=O), 1483, 1199, 754 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 6.87 (td, *J* = 7.6, 1.2 Hz, 1H), 6.92 (dd, *J* = 8.0, 0.8 Hz, 1H), 7.22-7.28 (m, 2H), 7.49 (d, *J* = 8.8 Hz, 1H), 7.66 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.89 (d, *J* = 2.4 Hz, 1H), 8.01 (s, 1H), 9.67 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 115.7 (CH), 117.9 (CH), 118.7 (CH), 120.7 (Cq), 121.8 (Cq), 127.1 (Cq), 127.3 (CH), 128.2 (Cq), 129.9 (CH), 130.7 (CH), 130.9 (CH), 140.6 (CH), 151.7 (Cq), 155.0 (Cq), 158.9 (Cq).

## 2-Chloro-6H-benzofuro[3,2-c]chromen-6-one (1n)



yield (0.066 g, 67 %); colorless solid; m.p. 236-238 °C.

**Rf:** 0.5 (10 % ethyl acetate/petroleum ether).

**IR** (**KBr**):  $\tilde{v} = 3089$ , 1755 (C=O), 1556, 1111, 983, 821 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 7.38 (d, *J* = 8.8 Hz, 1H), 7.43 (td, *J* = 8.0, 2.0 Hz, 2H), 7.48 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.61 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.94 (d, *J* = 2.4 Hz, 1H), 8.06-8.08 (m, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 106.6 (Cq), 111.9 (CH), 113.7 (Cq), 118.9 (CH), 121.4 (CH), 122.0 (CH), 123.2 (Cq), 125.5 (CH), 127.3 (CH), 130.3 (Cq), 131.9 (CH), 151.9 (Cq), 155.7 (Cq), 157.5 (Cq), 158.7 (Cq).

**HRMS (ESI):** for C₁₅H₇ClO₃H: m/z [M + H]⁺, calcd: 271.0162, found: 271.0193.

#### 3-(2-Hydroxyphenyl)-6-nitro-2H-chromen-2-one (520)



yield (0.999 g, 95 %); colorless solid; m.p. 236-238 °C; lit.¹⁷⁶ 236-237 °C.

**IR (KBr):**  $\tilde{v} = 3417$  (OH), 3078, 1724 (C=O), 1537, 1348, 761 cm⁻¹.

¹H NMR (DMSO-*d₆*, 400 MHz): δ 6.87-6.92 (m, 2H), 7.24-7.29 (m, 2H), 7.64 (d, *J* = 9.2 Hz, 1H), 8.17 (s, 1H), 8.41 (dd, *J* = 9.2, 2.8 Hz, 1H), 8.72 (d, *J* = 2.8 Hz, 1H), 9.87 (s, 1H).
¹³C NMR (DMSO-*d₆*, 100 MHz): δ 115.6 (CH), 117.4 (CH), 118.9 (CH), 119.5 (Cq), 121.4 (Cq), 124.1 (CH), 126.1 (CH), 127.8 (Cq), 130.2 (CH), 130.7 (CH), 140.8 (CH), 143.6 (Cq), 154.9 (Cq), 156.6 (Cq), 158.4 (Cq).

## <u>2-Nitro-6H-benzofuro[3,2-c]chromen-6-one (10)</u>



yield (0.053 g, 54 %); colorless solid; m.p. 264-266 °C.

**Rf:** 0.5 (35 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3091$ , 1757 (C=O), 1533, 1348, 974, 750 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 7.43-7.53 (m, 2H), 7.58 (d, *J* = 9.2 Hz, 1H), 7.67 (d, *J* = 8.0 Hz, 1H), 8.08 (dd, *J* = 7.6, 1.2 Hz, 1H), 8.40 (dd, *J* = 9.2, 2.8 Hz, 1H), 8.88 (d, *J* = 2.8 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 107.3 (Cq), 112.1 (CH), 113.1 (Cq), 118.2 (CH), 118.7 (CH), 122.1 (CH), 122.8 (Cq), 125.8 (CH), 126.4 (CH), 127.9 (CH), 144.2 (Cq), 155.9 (Cq), 156.5 (Cq), 156.6 (Cq), 158.2 (Cq).

**HRMS (ESI):** for C₁₅H₇NO₅H: m/z [M + H]⁺, calcd: 282.0397, found: 282.0397.

#### <u>3-(2-Hydroxy-4-methoxyphenyl)-2*H*-chromen-2-one (52p)</u>



yield (0.215 g, 80 %); colorless solid; m.p. 170-172 °C; lit.¹¹³ 171-172 °C.

**IR** (**KBr**):  $\tilde{v} = 3192$  (OH), 2960, 1724 (C=O), 1602, 1203, 761 cm⁻¹.

¹**H** NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  3.72 (s, 3H), 6.44-6.46 (m, 2H), 7.19 (d, J = 8.8 Hz, 1H), 7.33 (td, J = 8.4, 1.2 Hz, 1H), 7.39 (d, J = 8.0 Hz, 1H), 7.58 (td, J = 8.4, 1.6 Hz, 1H), 7.69 (dd, J = 8.0, 1.6 Hz, 1H), 7.93 (s, 1H), 9.87 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.0 (OCH₃), 101.3 (CH), 104.5 (CH), 114.8 (Cq), 115.8 (CH), 119.3 (Cq), 124.5 (CH), 125.5 (Cq), 128.2 (CH), 131.3 (CH), 131.5 (CH), 141.4 (CH), 152.8 (Cq), 156.1 (Cq), 159.7 (Cq), 160.5 (Cq).

## 9-Methoxy-6H-benzofuro[3,2-c]chromen-6-one (1p)



yield (0.069 g, 70 %); colorless solid; m.p. 216-218 °C; lit.¹¹² 216 °C.

**IR (KBr):**  $\tilde{v} = 2993$ , 1743 (C=O), 1504, 1282, 750 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz):  $\delta$  3.94 (s, 3H), 7.09 (dd, J = 8.4, 2.0 Hz, 1H), 7.21 (d, J = 2.0 Hz, 1H), 7.42 (td, J = 8.4, 1.2 Hz, 1H), 7.52 (dd, J = 8.4, 0.8 Hz, 1H), 7.58-7.62 (m, 1H), 7.99-8.02 (m, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.9 (OCH₃), 96.8 (CH), 106.1 (Cq), 112.9 (Cq), 113.6 (CH), 116.5 (Cq), 117.4 (CH), 121.5 (CH), 122.0 (CH), 124.6 (CH), 131.3 (CH), 153.2 (Cq), 156.8 (Cq), 158.3 (Cq), 159.3 (Cq), 159.7 (Cq).

## 3-(2-Hydroxy-4-methoxyphenyl)-7-methoxy-2*H*-chromen-2-one (52q)



yield (0.167 g, 56 %); colorless solid; m.p. 160-162 °C; lit.¹¹³ 160-161 °C.

**IR** (**KBr**):  $\tilde{v} = 3304$  (OH), 2956, 1691 (C=O), 1604, 1510, 1278, 835 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 3.72 (s, 3H), 3.84 (s, 3H), 6.43-6.45 (m, 2H), 6.93 (d, *J* = 8.8 Hz, 1H), 6.97 (s, 1H), 7.16 (d, *J* = 8.8 Hz, 1H), 7.61 (d, *J* = 8.4 Hz, 1H), 7.87 (s, 1H), 9.76 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 54.9 (OCH₃), 55.8 (OCH₃), 100.2 (CH), 101.3 (CH), 104.4 (CH), 112.3 (CH), 112.8 (Cq), 115.0 (Cq), 121.9 (Cq), 129.2 (CH), 131.5 (CH), 141.7 (CH), 154.6 (Cq), 156.0 (Cq), 159.9 (Cq), 160.2 (Cq), 161.9 (Cq).

## 3,9-Dimethoxy-6H-benzofuro[3,2-c]chromen-6-one (2f)



yield (0.064 g, 65 %); colorless solid; m.p. 198-200 °C; lit.¹¹² 199-200 °C.

**IR (KBr):**  $\tilde{v} = 2951$ , 1739 (C=O), 1498, 1257, 1099, 840 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ 3.83 (s, 3H), 3.84 (s, 3H), 6.88-6.91 (m, 2H), 6.96 (dd, J = 8.8, 2.4 Hz, 1H), 7.08 (d, J = 2.4 Hz, 1H), 7.79 (d, J = 9.2 Hz, 1H), 7.87 (d, J = 8.4 Hz, 1H). ¹³C NMR (CDCl₃, 100 MHz): δ 55.8 (OCH₃), 55.9 (OCH₃), 96.8 (CH), 101.4 (CH), 103.5 (Cq), 106.1 (Cq), 113.0 (CH), 113.2 (CH), 116.6 (Cq), 121.6 (CH), 122.5 (CH), 155.1 (Cq), 156.5 (Cq), 158.5 (Cq), 159.2 (Cq), 160.1 (Cq), 162.6 (Cq).

## <u>3-(2-Hydroxy-4-methoxyphenyl)-3*H*-benzo[*f*]chromen-3-one (52r)</u>



yield (0.250 g, 79 %); colorless solid; m.p. 208-210 °C.

**Rf:** 0.58 (50 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3315$  (OH), 2993, 1689 (C=O), 1257, 1105, 808 cm⁻¹.

¹**H NMR** (**DMSO**-*d*₆, **400 MHz**): δ 3.74 (s, 3H), 6.49-6.52 (m, 2H), 7.29 (d, *J* = 9.2 Hz, 1H), 7.57 (d, *J* = 8.8 Hz, 1H), 7.61 (dd, *J* = 8.0, 0.8 Hz, 1H), 7.68-7.72 (m, 1H), 8.03 (d, *J* = 7.2 Hz, 1H), 8.14 (d, *J* = 9.2 Hz, 1H), 8.49 (d, *J* = 8.4 Hz, 1H), 8.74 (s, 1H), 9.88 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.1 (OCH₃), 101.3 (CH), 104.6 (CH), 113.2 (Cq), 115.2 (Cq), 116.4 (CH), 122.2 (CH), 124.9 (Cq), 125.9 (CH), 128.2 (CH), 128.6 (Cq), 128.8 (CH), 129.8 (Cq), 131.8 (CH), 132.4 (CH), 137.5 (CH), 152.3 (Cq), 156.2 (Cq), 159.7 (Cq), 160.6 (Cq).

**HRMS (ESI):** for  $C_{20}H_{14}O_4Na: m/z [M + Na]^+$ , calcd: 341.0790, found: 341.0790.

## 11-Methoxy-8*H*-benzo[*f*]benzofuro[3,2-*c*]chromen-8-one (1q)



yield (0.054 g, 55 %); colorless solid; m.p. 238-240 °C.

**Rf:** 0.65 (35 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 2972$ , 1730 (C=O), 1504, 1273, 812 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz):  $\delta$  3.87 (s, 3H), 7.01-7.03 (m, 1H), 7.22 (br s, 1H), 7.49-7.55 (m, 2H), 7.69 (t, *J* = 8.4 Hz, 1H), 7.85 (d, *J* = 8.0 Hz, 1H), 7.89 (d, *J* = 8.8 Hz, 1H), 7.97 (dd, *J* = 8.8, 1.2 Hz, 1H), 9.09 (d, *J* = 7.2 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.9 (OCH₃), 96.6 (CH), 106.3 (Cq), 107.4 (Cq), 113.9 (CH), 115.8 (Cq), 117.4 (CH), 122.0 (CH), 125.5 (CH), 126.3 (CH), 127.4 (Cq), 128.6 (CH), 128.8 (CH), 130.4 (Cq), 132.5 (CH), 153.2 (Cq), 156.7 (Cq), 158.2 (Cq), 159.7 (Cq), 160.7 (Cq).

**HRMS (ESI):** for  $C_{20}H_{12}O_4Na: m/z [M + Na]^+$ , calcd: 339.0633, found: 339.0633.

## 3-(2-Hydroxy-4-methoxyphenyl)-6-methyl-2H-chromen-2-one (52s)



yield (0.220 g, 78 %); colorless solid; m.p. 180-182 °C.

**Rf:** 0.5 (50 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3342$  (OH), 2912, 1678 (C=O), 1502, 1153, 796 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 2.35 (s, 3H), 3.72 (s, 3H), 6.44-6.47 (m, 2H), 7.18 (d, *J* = 8.8 Hz, 1H), 7.28 (d, *J* = 8.4 Hz, 1H), 7.39 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.49 (d, *J* = 1.2 Hz, 1H), 7.87 (s, 1H), 9.82 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 20.2 (CH₃), 55.0 (OCH₃), 101.3 (CH), 104.5 (CH), 114.9 (Cq), 115.5 (CH), 119.0 (Cq), 125.4 (Cq), 127.8 (CH), 131.5 (CH), 132.1 (CH), 133.7 (Cq), 141.4 (CH), 150.9 (Cq), 156.1 (Cq), 159.9 (Cq), 160.4 (Cq).

**HRMS (ESI):** for  $C_{17}H_{14}O_4Na: m/z [M + Na]^+$ , calcd: 305.0790, found: 305.0791.

#### 9-Methoxy-2-methyl-6H-benzofuro[3,2-c]chromen-6-one (1r)



yield (0.065 g, 66 %); colorless solid; m.p. 198-200 °C.

**Rf:** 0.64 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3008$ , 1743 (C=O), 1508, 1273, 812 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 2.42 (s, 3H), 3.85 (s, 3H), 6.99 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.11 (d, *J* = 2.4 Hz, 1H), 7.31 (s, 2H), 7.70 (s, 1H), 7.92 (d, *J* = 8.8 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 20.9 (CH₃), 55.9 (OCH₃), 96.8 (CH), 105.9 (Cq), 112.5 (Cq), 113.5 (CH), 116.6 (Cq), 117.1 (CH), 121.1 (CH), 121.9 (CH), 132.4 (CH), 134.5 (Cq), 151.5 (Cq), 156.7 (Cq), 158.4 (Cq), 159.4 (Cq), 159.6 (Cq).

**HRMS (ESI):** for  $C_{17}H_{12}O_4Na: m/z [M + Na]^+$ , calcd: 303.0633, found: 303.0633.

#### <u>3-(2-Hydroxy-4-methoxyphenyl)-7,8-dimethoxy-2H-chromen-2-one (52t)</u>



yield (0.198 g, 60 %); colorless solid; m.p. 194-196 °C.

**Rf:** 0.45 (50 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3377$  (OH), 2943, 1703 (C=O), 1606, 1284, 802 cm⁻¹.

¹**H NMR** (**DMSO-***d*₆, **400 MHz**): δ 3.71 (s, 3H), 3.83 (s, 3H), 3.89 (s, 3H), 6.43-6.46 (m, 2H), 7.09 (d, *J* = 8.8 Hz, 1H), 7.16 (d, *J* = 8.8 Hz, 1H), 7.43 (d, *J* = 8.8 Hz, 1H), 7.86 (s, 1H), 9.81 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.0 (OCH₃), 56.3 (OCH₃), 60.8 (OCH₃), 101.2 (CH), 104.5 (CH), 109.2 (CH), 113.9 (Cq), 114.9 (Cq), 122.2 (Cq), 123.4 (CH), 131.5 (CH), 134.8 (Cq), 141.9 (CH), 146.6 (Cq), 154.6 (Cq), 156.0 (Cq), 159.6 (Cq), 160.3 (Cq).

**HRMS (ESI):** for  $C_{18}H_{16}O_6Na: m/z [M + Na]^+$ , calcd: 351.0845, found: 351.0845.

## 3,4,9-Trimethoxy-6H-benzofuro[3,2-c]chromen-6-one (1s)



yield (0.059 g, 60 %); colorless solid; m.p. 208-210 °C; lit.⁹⁸ 209-210 °C.

**IR (KBr):**  $\tilde{v} = 2953$ , 1743 (C=O), 1502, 1290, 1097, 798 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz):  $\delta$  3.84 (s, 3H), 3.93 (s, 3H), 3.98 (s, 3H), 6.93 (d, *J* = 8.8 Hz, 1H), 6.97 (dd, *J* = 8.8, 2.0 Hz, 1H), 7.09 (d, *J* = 2.0 Hz, 1H), 7.59 (d, *J* = 8.4 Hz, 1H), 7.89 (d, *J* = 8.4 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.9 (OCH₃), 56.5 (OCH₃), 61.7 (OCH₃), 96.8 (CH), 103.8 (Cq), 107.4 (Cq), 108.9 (CH), 113.2 (CH), 116.4 (CH), 116.6 (Cq), 121.7 (CH), 136.9 (Cq), 147.4 (Cq), 155.4 (Cq), 156.5 (Cq), 157.9 (Cq), 159.3 (Cq), 159.8 (Cq).

#### 3-(2-Hydroxy-4-methoxyphenyl)-6,7-dimethoxy-2H-chromen-2-one (52u)



yield (0.209 g, 64 %); colorless solid; m.p. 200-202 °C; lit.¹¹³ 200-201 °C. **IR (KBr):**  $\tilde{v} = 3313$  (OH), 2989, 1666 (C=O), 1514, 1244, 1008, 821, cm⁻¹. ¹**H NMR (DMSO-***d*₆, 400 MHz): δ 3.69 (s, 3H), 3.76 (s, 3H), 3.82 (s, 3H), 6.43-6.45 (m, 2H), 6.99 (s, 1H), 7.13 (d, *J* = 8.8 Hz, 1H), 7.18 (s, 1H), 7.80 (s, 1H), 9.88 (br s, 1H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 54.9 (OCH₃), 55.8 (OCH₃), 56.1 (OCH₃), 99.5 (CH), 101.2 (CH), 104.5 (CH), 108.6 (CH), 111.7 (Cq), 115.1 (Cq), 122.0 (Cq), 131.5 (CH), 141.9 (CH), 145.8 (Cq), 148.6 (Cq), 151.9 (Cq), 155.9 (Cq), 160.2 (Cq), 160.4 (Cq).

## 2,3,9-Trimethoxy-6H-benzofuro[3,2-c]chromen-6-one (1t)



yield (0.059 g, 60 %); colorless solid; m.p. 252-254 °C; lit.¹¹³ 253-254 °C. **IR (KBr):**  $\tilde{v} = 2951$ , 1726 (C=O), 1517, 1271, 1002, 825 cm⁻¹.

¹**H** NMR (CDCl₃, 400 MHz): δ 3.83 (s, 3H), 3.90 (s, 3H), 3.95 (s, 3H), 6.91 (s, 1H), 6.96 (dd, *J* = 8.4, 2.4 Hz, 1H), 7.07 (d, *J* = 2.0 Hz, 1H), 7.23 (s, 1H), 7.87 (d, *J* = 8.8 Hz, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 55.9 (OCH₃), 56.4 (2 X OCH₃), 96.8 (CH), 100.6 (CH), 101.6 (CH), 103.8 (Cq), 104.9 (Cq), 113.1 (CH), 116.7 (Cq), 121.7 (CH), 146.8 (Cq), 149.2 (Cq), 152.6 (Cq), 156.4 (Cq), 158.6 (Cq), 159.2 (Cq), 159.9 (Cq).

#### <u>3-(2-Hydroxy-5-methylphenyl)-2*H*-chromen-2-one (52v)</u>



yield (0.182 g, 72 %); colorless solid; m.p. 138-140 °C; lit.¹⁷⁶ 138-140 °C.

**IR** (**KBr**):  $\tilde{v} = 3398$  (OH), 2918, 1710 (C=O), 1610, 1510, 744 cm⁻¹.

¹**H NMR** (**DMSO**-*d*₆, **400 MHz**): δ 2.21 (s, 3H), 6.78 (d, *J* = 8.0 Hz, 1H), 7.01-7.05 (m, 2H), 7.35 (td, *J* = 7.6, 1.2 Hz, 1H), 7.41 (d, *J* = 8.4 Hz, 1H), 7.58-7.62 (m, 1H), 7.72 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.96 (s, 1H), 9.50 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 19.9 (CH₃), 115.5 (CH), 115.8 (CH), 119.2 (Cq), 121.9 (Cq), 124.5 (CH), 126.1 (Cq), 127.3 (Cq), 128.3 (CH), 130.2 (CH), 130.9 (CH), 131.5 (CH), 141.8 (CH), 152.6 (Cq), 152.9 (Cq), 159.5 (Cq).

## 8-Methyl-6H-benzofuro[3,2-c]chromen-6-one (1u)



yield (0.064 g, 65 %); colorless solid; m.p. 190-192 °C; lit.¹⁷⁶ 189-191 °C.

**IR (KBr):**  $\tilde{v} = 2924$ , 1737 (C=O), 1631, 1082, 756 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ 2.39 (s, 3H), 7.14 (d, J = 8.4 Hz, 1H), 7.29 (t, J = 7.6 Hz, 1H), 7.39 (t, J = 9.2 Hz, 2H), 7.48 (t, J = 8.4 Hz, 1H), 7.78 (s, 1H), 7.87 (d, J = 7.6 Hz, 1H). ¹³C NMR (CDCl₃, 100 MHz): δ 21.3 (CH₃), 105.6 (Cq), 111.2 (CH), 112.7 (Cq), 117.4 (CH), 121.6 (CH), 121.8 (CH), 123.3 (Cq), 124.6 (CH), 127.9 (CH), 131.7 (CH), 135.1 (Cq), 153.5 (Cq), 153.9 (Cq), 158.2 (Cq), 159.9 (Cq).

#### 3-(2-Hydroxy-5-methylphenyl)-6-methyl-2*H*-chromen-2-one (52w)



yield (0.180 g, 68 %); colorless solid; m.p. 190-192 °C.

**Rf:** 0.58 (40 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3367$  (OH), 2922, 1720 (C=O), 1581, 808 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 2.23 (s, 3H), 2.38 (s, 3H), 6.79 (d, *J* = 8.0 Hz, 1H), 7.03-7.07 (m, 2H), 7.33 (d, *J* = 8.4 Hz, 1H), 7.43 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.53 (d, *J* = 1.6 Hz, 1H), 7.93 (s, 1H), 9.44 (s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 19.9 (CH₃), 20.2 (CH₃), 115.5 (CH), 115.6 (CH), 118.9 (Cq), 121.9 (Cq), 125.9 (Cq), 127.2 (Cq), 127.9 (CH), 130.1 (CH), 130.9 (CH), 132.3 (CH), 133.7 (Cq), 141.7 (CH), 151.1 (Cq), 152.7 (Cq), 159.6 (Cq).

**HRMS (ESI):** for  $C_{17}H_{14}O_3Na: m/z [M + Na]^+$ , calcd: 289.0841, found: 289.0841.

## 2,8-Dimethyl-6H-benzofuro[3,2-c]chromen-6-one (1v)



yield (0.059 g, 60 %); colorless solid; m.p. 202-204 °C.

**Rf:** 0.48 (10 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 2920$ , 1753 (C=O), 1571, 1074, 794 cm⁻¹.

¹**H NMR (CDCl₃, 400 MHz):** δ 2.49 (s, 3H), 2.52 (s, 3H), 7.27 (d, *J* = 7.6 Hz, 1H), 7.39 (s, 2H), 7.53 (d, *J* = 8.4 Hz, 1H), 7.79 (s, 1H), 7.93 (s, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 20.9 (CH₃), 21.3 (CH₃), 105.6 (Cq), 111.1 (CH), 112.4 (Cq), 117.2 (CH), 121.5 (CH), 121.6 (CH), 123.5 (Cq), 127.8 (CH), 132.9 (CH), 134.5 (Cq), 135.1 (Cq), 151.8 (Cq), 153.9 (Cq), 158.5 (Cq), 160.1 (Cq).

**HRMS (ESI):** for  $C_{17}H_{12}O_3H$ : m/z  $[M + H]^+$ , calcd: 265.0865, found: 265.0865.

## 7-Hydroxy-3-(2-hydroxy-4-methoxyphenyl)-2*H*-chromen-2-one (52x)



yield (0.149 g, 53 %); pale yellow solid; m.p. 248-250 °C.

Rf: 0.43 (50 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3414$  (OH), 3257 (OH), 2997, 1676 (C=O), 1610, 1153, 812 cm⁻¹.

¹**H NMR (DMSO-***d***₆, 400 MHz):** δ 3.73 (s, 3H), 6.44-6.46 (m, 2H), 6.75 (d, *J* = 2.4 Hz, 1H), 6.79 (dd, *J* = 8.4, 2.4 Hz, 1H), 7.17 (d, *J* = 8.8 Hz, 1H), 7.55 (d, *J* = 8.8 Hz, 1H), 7.85 (s, 1H), 9.67 (s, 1H), 10.65 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 54.9 (OCH₃), 101.2 (CH), 101.6 (CH), 104.3 (CH), 111.8 (Cq), 112.9 (CH), 115.1 (Cq), 120.8 (Cq), 129.4 (CH), 131.5 (CH), 141.9 (CH), 154.7 (Cq), 155.9 (Cq), 160.0 (Cq), 160.1 (Cq), 160.6 (Cq).

**HRMS (ESI):** for  $C_{16}H_{12}O_5Na: m/z [M + Na]^+$ , calcd: 307.0582, found: 307.0582.

#### <u>3-Hydroxy-9-methoxy-6*H*-benzofuro[3,2-*c*]chromen-6-one (2e)</u>



yield (0.058 g, 59 %); pale yellow solid; m.p. >300 °C; lit.¹¹²>300 °C.

**IR** (**KBr**):  $\tilde{v} = 3267$  (OH), 2920, 1708 (C=O), 1500, 1267, 1101, 840 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 3.86 (s, 3H), 6.92 (d, *J* = 1.6 Hz, 1H), 6.96 (dd, *J* = 8.8, 2.0 Hz, 1H), 7.09 (dd, *J* = 8.4, 2.0 Hz, 1H), 7.47 (d, *J* = 1.6 Hz, 1H), 7.78 (d, *J* = 8.4 Hz, 1H), 7.87 (d, *J* = 8.4 Hz, 1H), 10.89 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 55.8 (OCH₃), 97.2 (CH), 101.8 (Cq), 103.0 (CH), 104.0 (Cq), 113.4 (CH), 113.9 (CH), 115.8 (Cq), 120.5 (CH), 122.8 (CH), 154.7 (Cq), 155.8 (Cq), 157.6 (Cq), 158.8 (Cq), 159.9 (Cq), 161.3 (Cq).

## 3-(2,4-Dihydroxyphenyl)-7-hydroxy-2H-chromen-2-one (52y)



yield (0.132 g, 49 %); pale yellow solid; m.p. >280 °C (decomp).

**Rf:** 0.48 (70 % ethyl acetate/petroleum ether).

**IR (KBr):**  $\tilde{v} = 3410$  (OH), 3315 (OH), 3147 (OH), 2746, 1693 (C=O), 1598, 1463, 1224, 995 cm⁻¹.

¹**H** NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  6.27 (dd, J = 8.4, 2.4 Hz, 1H), 6.35 (d, J = 2.4 Hz, 1H), 6.74 (d, J = 2.4 Hz, 1H), 6.79 (dd, J = 8.4, 2.4 Hz, 1H), 7.04 (d, J = 8.4 Hz, 1H), 7.53 (d, J = 8.4 Hz, 1H), 7.81 (s, 1H), 9.46 (s, 1H), 9.51 (s, 1H), 10.62 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 101.6 (CH), 102.5 (CH), 106.2 (CH), 111.9 (Cq), 112.9 (CH), 113.6 (Cq), 121.2 (Cq), 129.4 (CH), 131.4 (CH), 141.7 (CH), 154.6 (Cq), 155.9 (Cq), 158.3 (Cq), 160.2 (Cq), 160.5 (Cq).

**HRMS (ESI):** for  $C_{15}H_{10}O_5H$ : m/z  $[M + H]^+$ , calcd: 271.0601, found: 271.0601.

# <u>3,9-Dihydroxy-6H-benzofuro[3,2-c]chromen-6-one (2c)</u>



yield (0.054 g, 55 %); pale yellow solid; m.p. >300 °C; lit.¹⁸³>300 °C.

**IR** (**KBr**):  $\tilde{v} = 3398$  (OH), 3082, 2372, 1701 (C=O), 1629, 1300, 1091, 810 cm⁻¹.

¹**H NMR (DMSO-***d*₆, **400 MHz):** δ 6.92-6.98 (m, 3H), 7.19 (d, *J* = 2.0 Hz, 1H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.89 (d, *J* = 8.4 Hz, 1H), 10.20 (br s, 1H), 10.88 (br s, 1H).

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 98.7 (CH), 101.9 (Cq), 102.9 (CH), 104.1 (Cq), 113.8 (CH), 114.0 (CH), 114.5 (Cq), 120.6 (CH), 122.8 (CH), 154.6 (Cq), 155.9 (Cq), 156.9 (Cq), 157.7 (Cq), 159.5 (Cq), 161.1 (Cq).



Scheme 74: Stepwise one pot synthesis of coumestan 1a.

The protocol was then successfully tested for the preparation of **1a** from **52a** on a larger scale (2 mmol) thus demonstrating its utility. Further, a one pot procedure was attempted by mixing 2-coumaranone **51**, salicylaldehyde **50a**, Cu(OAc)₂ and NEt₃ in diphenyl ether as the solvent system. However, no formation of **1a** was observed (Scheme 74-*A*). Hence a stepwise one pot approach was developed wherein NEt₃ was removed under reduced pressure before addition of Cu(OAc)₂ giving product **1a** in good yield (Scheme 74-*B*).

Mechanistic studies were performed to check whether the mechanism is following a radical pathway. Hence the reaction was performed in presence of a radical scavenger TEMPO. It was found to show no effect on the yield of product **1a** suggesting an alternative mechanism. Based on this observation a speculative mechanism is proposed for this intramolecular C-O cyclization (Scheme 75). Cu(II) from Cu(OAc)₂ binds to electronegative hydroxyl oxygen atom of substrate **52** with the liberation of one molecule of acetic acid to form intermediate **54**. Intramolecular oxidative addition of copper to the C-H bond renders **56** *via* the elimination of another molecule of acetic acid from **55**. Finally the reductive elimination of **56** furnished coumestan **1** and metallic copper which was supported from recorded XRD (Figure 12) of the residue left after reaction and also from the copper mirror deposits on the walls of the reaction flask. The Cu₂O visible from XRD could be due to the aerobic oxidation of Cu at high reaction temperature.



Scheme 75: Proposed mechanism for the formation of coumestans using Cu(OAc)₂.



Figure 12: (A) XRD of Cu, Cu₂O and the residue left after reaction (B) Overlay.

It is known that the presence of other metal impurity or "homeopathic" metal can also be responsible for such results.¹⁸⁴ But, it was essential to confirm that presence of Pd impurity was not responsible for this Cu(OAc)₂ mediated cylization. Hence Cu(OAc)₂ sample was tested for the presence of Pd by ICP-MS analysis which showed the presence of 1817.97 ppb of Pd. So accordingly to gain some clarity in whether Pd has any role in this oxidative cyclization we conducted two experiments in presence of 1 equiv of Cu(OAc)₂ along with 0.01 equiv of Pd(OAc)₂ or PdCl₂ (Scheme 76). It was observed that on addition of external Pd source there was no much difference visible in terms of reaction time or product yield. Also the incomplete conversion of starting resulted in partial product formation when 0.1 equiv of Pd(OAc)₂ or PdCl₂ for coumestan synthesis over longer duration of time (24 h) which support our observation. Also, a reaction using catalytic PdCl₂ under Wacker condition in presence of CuCl₂ in DMF:DMA (1:1) solvent system at 150 °C was attempted giving trace amount of product. All these results suggests that Cu is playing the major role in the present

#### **CHAPTER 4**

oxidative cyclization reaction and not the Pd impurity present in copper. However the probable role of Pd present in ppb level in contact with copper and their synergestic effect cannot be ruled out completely.



Scheme 76: Reactions in Pd source in presence/absence of Cu(OAc)₂.

#### 4.5: Conclusion

We developed an efficient methodology for the synthesis of coumestans by implementing economical  $Cu(OAc)_2$  as the sole reagent in absence of any additional reagent/additive in diphenyl ether solvent *via* C-H activation.

Several advantages are associated such as simple reaction procedure, large substrate scope, effortless product isolation & good yields which makes this method superior over reported methods.

Method was applied successfully for the synthesis of hydroxy substituted naturally occurring coumestans *viz*. coumestrol and 4'-*O*-methylcoumestrol without protection/deprotection strategies making this method attractive.

Also one pot synthesis and possible use of catalytic amount of Cu(OAc)₂ is demonstrated.

#### 4.6: Experimental



**4.6.1:** A general procedure for the synthesis of substrates 52a-o:¹⁷⁶ Substituted salicylaldehyde derivative 50 (3.7 mmol) and 2-coumaranone 51 (0.5 g, 3.7 mmol) were mixed together in a round bottom flask. To it triethylamine (15 mL) was added and refluxed for 1 h. After 1 h the solvent was removed under vacuum and the crude solid was recrystallized from ethanol to afford pure product 3-(2-hydroxyphenyl)-2*H*-chromen-2-one 52a-o.



**4.6.2:** A general procedure for the synthesis of substituted 2-hydroxyphenylacetic acid **53a-c:**¹⁸⁵ Substituted salicylaldehyde derivative **50** (29 mmol) along with morpholine (87 mmol), sulphur (58 mmol) and *p*-toluenesulphonic acid (1 mmol) were refluxed with stirring at 120-130 °C for 8 h. After reaction completion the mixture was cooled to room temperature. Hydrolysis was carried out by heating in presence of 20 % aq. NaOH (41 mL) and tetrabutylammonium bromide (0.46 mmol) at 100 °C for 8 h. The reaction mixture was cooled and filtered. The filtrate was acidified with HCl to pH 2. It was then extracted with ethyl acetate (3 times) and combined organic layer was washed with water and dried over sodium sulphate. The crude product obtained was purified by column chromatography using petroleum ether-ethyl acetate as eluents.



**4.6.3:** A general procedure for the synthesis of substrates 52p-y:¹⁷⁷ Substituted salicylaldehyde derivative **50** (1 mmol), substituted 2-hydroxyphenylacetic acid **53** (1 mmol), sodium acetate (5.0 mmol) and acetic anhydride (2.4 mmol) were mixed together in a round bottom flask. To it acetic acid (4 mL) was added and refluxed for 24 h. After 24 h the solvent was removed under vacuum and water was added to it. The crude solid obtained was filtered and then loaded on column (eluent: petroleum ether-ethyl acetate) to afford pure product 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52p-y**.



**4.6.4:** A procedure for the attempted one pot synthesis of coumestan 1a: 2-coumaranone **51** (0.1 g, 0.75 mmol) and 2'-hydroxybenzaldehyde **50a** (0.091 g, 0.75 mmol) were mixed together and refluxed in 10 mL of triethylamine in a 50 mL round bottom flask. After 1 h refluxing, the solvent was removed under reduced pressure and to the same vessel, anhyd. FeCl₃ (1.209 g, 7.5 mmol) was added along with 5 mL of 1,2-dichloroethane solvent. The reaction mixture was then refluxed for 24 h and showed trace amount of product **1a** on TLC.



**4.6.5:** A procedure for the synthesis of coumestan 1a using FeCl₃ in 1,2-dichloroethane solvent: Anhyd. FeCl₃ (0.68 g, 4.2 mmol) or (0.21 g, 1.26 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.42 mmol) in a 25 mL round bottom flask. To it 10 mL of 1,2-dichloroethane solvent was added. The resulting mixtures were then heated to reflux/120 °C respectively. After 24 h, the solvent was removed under reduced pressure and was loaded on column to afford product 6*H*-benzofuro[3,2-*c*]chromen-6-one **1a** as colorless solid (0.020 g, 20 %) or (0.011 g, 11 %) respectively with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.6:** A procedure for the synthesis of coumestan 1a using FeCl₃ under neat condition in absence of 230-400 mesh silica gel: Anhyd. FeCl₃ (0.17 g, 1.05 mmol) or (0.21 g, 1.26 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.42 mmol) in a 25 mL round bottom flask. The resulting mixtures were then heated in absence of solvent (neat) at 150/120 °C for 24/15 h respectively. After the specified time, the reaction mixture was loaded on column to afford product **1a** (0.012 g, 12 %) or (0.011 g, 11 %) respectively with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.7:** A procedure for the synthesis of coumestan 1a using FeCl₃ under neat condition in presence of 230-400 mesh silica gel: Anhyd. FeCl₃ (0.17 g, 1.05 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.42 mmol) and 230-400 silica gel (0.17 g) in a 25 mL round bottom flask. The resulting mixture was then heated in absence of solvent (neat) at 150 °C for 24 h. Then the reaction mixture was loaded on column to afford product 6*H*-benzofuro[3,2-*c*]chromen-6-one **1a** as colorless solid (0.019 g, 19 %) with petroleum ether-ethyl acetate (8.5:1.5) as an eluent. In other case, similar reaction was carried out to which after 24 h water was added and refluxed for 24 h. The water was then removed under

reduced pressure and the reaction mixture was loaded on column to afford product **1a** (0.024 g, 24 %) with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.8:** A procedure for the synthesis of coumestan 1a using FeCl₃ under neat condition: Anhyd. FeCl₃ (0.17 g, 1.05 mmol) or (0.27 g, 1.68 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.42 mmol) in a 25 mL round bottom flask. The resulting mixtures were then heated in absence of solvent (neat) at 150/180 °C for 24 h respectively. Then the reaction mixture was sonicated for 30 minutes with a gap after every 5 minutes and then loaded on column to afford product **1a** (0.033 g, 33 %)/(0.025 g, 25 %) respectively with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.9:** A procedure for the synthesis of coumestan 1a using FeCl₃ under neat condition in presence of 230-400 mesh silica gel: Anhyd. FeCl₃ (0.20 g, 1.26 mmol) or (0.27 g, 1.68 mmol) or (0.34 g, 2.1 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.42 mmol) and 230-400 silica gel (0.20/0.27/0.34 g) respectively in a 25 mL round bottom flask. The resulting mixtures were then heated in absence of solvent (neat) at 150 °C for 24 h. Then the reaction mixtures were sonicated for 30 minutes with a gap after every 5 minutes and loaded on column to afford **1a** (0.036 g, 36 %)/(0.042 g, 42 %)/(0.039 g, 39 %) respectively with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.





further work up it was directly loaded on column to afford pure product **1a** in 7-86 % yield with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.11:** A general procedure for the synthesis of coumestans 1a-v,2c,2e-f using  $Cu(OAc)_2$ :  $Cu(OAc)_2$  (0.4 mmol) was added to substituted 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a-y** (0.4 mmol) in a 25 mL round bottom flask. To it 10 mL of diphenyl ether was added. The resulting mixture was then heated to reflux for 4-24 h. After completion of the reaction (monitored by TLC) the reaction mass was cooled to room temperature. Without any further work up it was directly loaded on column to afford pure product 6*H*-benzofuro[3,2-*c*]chromen-6-one **1a-v,2c,2e-f** in 30-80 % yield with petroleum ether-ethyl acetate as an eluent.



**4.6.12:** Procedure for the synthesis of coumestan 1a:  $Cu(OAc)_2$  (0.076 g, 0.42 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.42 mmol) in a 50 mL round bottom flask. To it 10 mL of diphenyl ether was added. The resulting mixture was then heated to reflux for 6 h. After completion of the reaction (monitored by TLC) the reaction mass was cooled to room temperature. Without any further work up it was directly loaded on column to afford product **1a** (0.075 g, 76 %) with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.13:** Procedure for the synthesis of coumestan 1a on 0.5 g scale:  $Cu(OAc)_2 (0.382 \text{ g}, 2.1 \text{ mmol})$  was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.5 g, 2.1 mmol) in a 50 mL round bottom flask. To it 20 mL of diphenyl ether was added. The resulting mixture was then heated to reflux for 21 h. After completion of the reaction (monitored by TLC) the reaction mass was cooled to room temperature. Without any further work up it was directly loaded on column to afford product **1a** (0.296 g, 60 %) with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.14: Procedure for stepwise one pot synthesis of coumestan 1a:** 2-coumaranone **51** (0.057 g, 0.4 mmol) and salicylaldehyde **50a** (0.052 g, 0.4 mmol) were mixed together in a 25 mL round bottom flask. To it triethylamine (5 mL) was added and refluxed for 1 h. After reaction solvent was removed under vacuum and to the product formed 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a**, Cu(OAc)₂ (0.077 g, 0.4 mmol) and 10 mL of diphenyl ether were added. The resulting mixture was then heated to reflux for 6 h. After completion of the reaction (monitored by TLC) the reaction mass was cooled to room temperature. Without any further work up it was directly loaded on column to afford product **1a** (0.070 g, 71 %) with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.



**4.6.15: Procedure for synthesis of coumestan 1a in presence of radical scavenger TEMPO:** TEMPO (0.131 g, 0.8 mmol) was added to 3-(2-hydroxyphenyl)-2*H*-chromen-2-one **52a** (0.1 g, 0.4 mmol) and Cu(OAc)₂ (0.077 g, 0.4 mmol) in a 25 mL round bottom flask. To it 10 mL of diphenyl ether was added. The resulting mixture was flushed with argon gas few times and then heated with stirring at 100 °C for 24 h in argon atmosphere. After 24 h the reaction mass was cooled to room temperature. Without any further work up it was directly loaded on column to afford product **1a** (0.027 g, 27 %) with petroleum ether-ethyl acetate (8.5:1.5) as an eluent.

#### 4.7: References

- Ollis, W. D. Recent advances in Phytochemistry, Edited by Mabry, T. A.; Alston, R. E.; Runeckles, V. C. (Appleton – Century- Crofts, New York), 1966, 345.
- a) Darbarwar, M.; Sundaramurthy, V.; Subba Rao, N. V. J. Sci. Ind. Res. 1976, 35, 297. b) Jain, A. C.; Tuli, D. K. J. Sci. Ind. Res. 1978, 37, 287.
- a) Upadhyay, R. K.; Singh, S.; Pandey, V. B. Orient. J. Chem. 2001, 17, 369. b) Helferich, W. G.; Allred, C. D.; Ju, Y-H. Food Toxicology 2001, 37. c) Al-Hazimi, H. M. G.; Alkhathlan, H. Z. J. King Saud Univ. Sci. 2000, 12, 93. d) Dean, F. M. Total Synth. Nat. Prod. 1973, 1,

467. e) Verdeal, K.; Ryan, D. S. J. Food Prot. **1979**, 42, 577. f) Cornwell, T.; Cohick, W.; Raskin, I. *Phytochemistry* **2004**, 65, 995.

- 4. Govindachari, T. R.; Nagarajan, K.; Pai, B. R. J. Chem. Soc. 1956, 629.
- a) Liu, Q-M.; Zhao, H-Y.; Zhong, X-K.; Jiang, J-G. Food Chem. Toxicol. 2012, 50, 4016. b) Bhargava, K. K.; Krishnaswamy, N. R.; Seshadri, T. R. Indian J. Chem. 1970, 8, 664. c) Banu, H. R.; Nagarajan, N. J. Chem. Pharm. Res. 2013, 5, 279. d) Sarg, T. M.; Salam, N. A. A.; El-Domiaty, M.; Khafagy, S. M. Sci. Pharm. 1981, 49, 262. e) Lee, K. Y.; Ha, N. R.; Kim, T. B.; Kim, Y. C.; Sung, S. H. Nat. Prod. Sci. 2010, 16, 164.
- 6. a) Bickoff, E. M.; Booth, A. N.; Lyman, R. L.; Livingston, A. L.; Thompson, C. R.; Deeds, F. Science 1957, 126, 969. b) Lyman, R. L.; Bickoff, E. M.; Booth, A. N.; Livingston, A. L. Arch. Biochem. Biophys. 1959, 80, 61. c) Livingston, A. L.; Bickoff, E. M.; Guggolz, J.; Thompson, C. R. J. Agric. Food Chem. 1961, 9, 135. d) Guggolz, J.; Livingston, A. L.; Bickoff, E. M. J. Agric. Food Chem. 1961, 9, 330. e) Millington, A. J.; Francis, C. M.; McKeown, N. R. Austral. Jour. Agr. Res. 1964, 15, 520. f) Francis, C. M.; Millington, A. J. Austral. Jour. Agr. Res. 1965, 16, 927. g) Bennett, D.; Morley, F. H. W.; Axelsen, A. Austral. Jour. Agr. Res. 1967, 18, 495. h) Wong, E. Jour. Sci. Food Agr. 1962, 13, 304. i) Knuckles, B. E.; deFremery, D.; Kohler, G. O. J. Agric. Food Chem. 1976, 24, 1177. j) Knuckles, B. E.; Miller, R. E.; Bickoff, E. M. J. Assoc. Off. Anal. Chem. 1975, 58, 983. k) Wada, H.; Yuhara, M. Jap. Jour. Zootec. Sci. 1964, 35, 87. 1) Fritsche, S.; Steinhart, H. Eur. Food Res. Technol. 1999, 209, 153. m) Price, K. R.; Fenwick, G. R. Food Addit. Contam. 1985, 2, 73. n) Franke, A. A.; Custer, L. J.; Cerna, C. M.; Narala, K. Proc. Soc. Exp. Biol. Med. 1995, 208, 18. o) Wang, G.; Kuan, S. S.; Francis, O. J.; Ware, G. M.; Carman, A. S. J. Agric. Food Chem. 1990, 38, 185. p) Chansakaow, S.; Ishikawa, T.; Sekine, K.; Okada, M.; Higuchi, Y.; Kudo, M.; Chaichantipyuth, C. Planta Med. 2000, 66, 572. q) Zoghbi, M. das G. B.; Marques, M. de F. dos S.; Cabral, J. A. da S.; Braz Filho, R. Acta Amazon. 1988, 18, 57. r) Lookhart, G. L.; Jones, B. L.; Finney, K. F. Cereal chem. 1978, 55, 967. s) Hutabarat, L.S.; Greenfield, H.; Mulholland, M. J. Chromatogr. A 2000, 886, 55. t) Reinli, K.; Block, G. Nutr. Cancer 1996, 26, 123. u) Ingham, J. L.; Tahara, S.; Dziedzic, S. Z. Z. Naturforsch. C 1986, 41, 403. v) Takeya, K.; Itokawa, H. Chem. Pharm. Bull. 1982, 30, 1496. w) Bickoff, E. M.; Booth, A. N.; Lyman, R. L.; Livingston, A. L.; Thompson, C. R.; Kohler, G. O. J. Agric. Food Chem. 1958, 6, 536. x) Koshino, H.; Masaoka, Y.; Ichihara, A. Phytochemistry 1993, 33, 1075. y) Saxena, V. K.; Nigam, S. J. Inst. Chem. 1996, 68, 122. z) Zilg, H.; Grisebach, H. Phytochemistry 1968, 7, 1765.
- a) Fu, H. W.; Zhang, H. L.; Pei, Y. H. Chin. Chem. Lett. 2005, 16, 918. b) Chang, L. C.; Gerhäuser, C.; Song, L.; Farnsworth, N. R.; Pezzuto, J. M.; Kinghorn, A. D. J. Nat. Prod. 1997, 60, 869.

- a) Wong, E.; Latch, G. C. M. Phytochemistry 1971, 10, 466. b) Macías, F. A.; Simonet, A. M.; Galindo, J. C. G.; Castellano, D. Phytochemistry 1999, 50, 35. c) Bickoff, E. M.; Spencer, R. R.; Knuckles, B. E.; Lundin, R. E. J. Agric. Food Chem. 1966, 14, 444. d) Bickoff, E. M.; Livingston, A. L.; Witt, S. C.; Lundin, R. E.; Spencer, R. R. J. Agric. Food Chem. 1965, 13, 597. e) Zilg, H.; Grisebach, H. Phytochemistry 1969, 8, 2261. f) Donnelly, D. M. X.; Kavanagh, P. J. Phytochemistry 1974, 13, 2587. g) Donnelly, D. M. X.; Thompson, J. C.; Whalley, W. B.; Ahmad, S. J. Chem. Soc., Perkin Trans 1 1973, 1737. h) Stevenson, P. C.; Veitch, N. C. Phytochemistry 1998, 48, 995. i) Dewick, P. M. Phytochemistry 1977, 16, 93. j) Francis, C. M.; Millington, A. J. Austral. Jour. Agr. Res. 1971, 22, 75. k) Jurd, L.; Wong, R. Y. Aust. J. Chem. 1984, 37, 1127. l) Lu, D.; He, H.; Wu, B.; Yao, S. Nat. Prod. Commun. 2009, 4, 809. m) Chan, S-C.; Chang, Y-S.; Wang, J-P.; Chen, S-C.; Kuo, S-C. Planta Med. 1998, 64, 153.
- 9. De Alencar, R.; Braz Filho, R.; Gottlieb, O. R. Phytochemistry 1972, 11, 1517.
- a) Shirataki, Y.; Tsuzuku, T.; Yokoe, I.; Hirano, R. T.; Komatsu, M. Chem. Pharm. Bull.
   **1990**, 38, 1712. b) Ferreira, D.; Kamara, B. I.; Brandt, E. V.; Joubert, E. J. Agric. Food Chem.
   **1998**, 46, 3406. c) Fukunaga, T.; Nishiya, K.; Takeya, K.; Itokawa, H. Chem. Pharm. Bull.
   1987, 35, 1610. d) Shiao, Y-J.; Wang, C-N.; Wang, W-Y.; Lin, Y-L. Planta Med. 2005, 71,
   835. e) Tang, Y-P.; Hu, J.; Wang, J-H.; Lou, F-C. J. Asian Nat. Prod. Res. 2002, 4, 1.
   f) Komatsu, M.; Yokoe, I.; Shirataki, Y. Chem. Pharm. Bull. 1978, 26, 1274. g) Shirataki, Y.;
   Komatsu, M.; Yokoe, I.; Manaka, A. Chem. Pharm. Bull. 1981, 29, 3033. h) Livingston, A.
   L.; Witt, S. C.; Lundin, R. E.; Bickoff, E. M. J. Org. Chem. 1965, 30, 2353. i) Takai, M.;
   Yamaguchi, H.; Saitoh, T.; Shibata, S. Chem. Pharm. Bull. 1972, 20, 2488.
- 11. Spencer, R. R.; Bickoff, E. M.; Lundin, R. E.; Knuckles, B. E. J. Agric. Food. Chem. 1966, 14, 162.
- 12. a) Rajani, P.; Sarma, P. N. *Phytochemistry* 1988, 27, 648. b) Mahabusarakam, W.; Deachathai, S.; Phongpaichit, S.; Jansakul, C.; Taylor, W. C. *Phytochemistry* 2004, 65, 1185.
  c) Adityachaudhury, N.; Gupta, P. K. *Phytochemistry* 1973, 12, 425. d) Chaudhury, N. A.; Gupta, P. K. *Chem. Ind.* 1970, 34, 1113.
- a) Donnelly, D. M. X.; Fitzgerald, M. A. *Phytochemistry* **1971**, *10*, 3147. b) Burns, D.T.;
   Dalgarno, B. G.; Gargan, P. E.; Grimshaw, J. *Phytochemistry* **1984**, *23*, 167.
- 14. a) O'Neill, M. J. Z. Naturforsch. 1983, 38c, 698. b) Wang, W.; Zhao, Y-Y.; Liang, H.; Jia, Q.;
  Chen, H-B. J. Nat. Prod. 2006, 69, 876. c) Yang M.; Wang, W.; Sun, J.; Zhao, Y.; Liu, Y.;
  Liang, H.; Guo, D. Rapid Commun. Mass Spectrom. 2007, 21, 3833.
- 15. Spencer, R. R.; Knuckles, B. E.; Bickoff, E. M. J. Org. Chem. 1966, 31, 988.
- 16. Li, L.; Deng, X-Y.; Zhang, L-X.; Shu, P.; Qin, M-J. Fitoterapia 2011, 82, 615.

- 17. a) Hatano, T.; Aga, Y.; Shintani, Y.; Ito, H.; Okuda, T.; Yoshida, T. *Phytochemistry* 2000, 55, 959. b) Tao, W-W.; Duan, J-A.; Yang, N-Y.; Tang, Y-P.; Liu, M-Z.; Qian, Y-F. *Fitoterapia* 2012, 83, 422.
- 18. Komatsu, M.; Yokoe, I.; Shirataki, Y. Chem. Pharm Bull. 1981, 29, 2069.
- 19. Yadav, P. P.; Ahmad, G.; Maurya, R. Phytochemistry 2004, 65, 439.
- 20. Nkengfack, A. E.; Kouam, J.; Vouffo, T. W.; Meyer, M.; Tempesta, M. S.; Fomum, Z. T. *Phytochemistry* **1994**, *35*, 521.
- 21. Abdel-Kader, M. S. Planta Medica 2001, 67, 388.
- 22. Biggs, D. R.; Shaw, G. J. Phytochemistry 1980, 19, 2801.
- 23. Rao, P. P.; Srimannarayana, G. Phytochemistry 1980, 19, 1272.
- a) Bickoff, E. M.; Livingston, A. L.; Witt, S. C.; Knuckles, B. E.; Guggolz, J.; Spencer, R. R. J. Pharm. Sci. 1964, 53, 1496. b) Livingston, A. L.; Bickoff, E. M.; Lundin, R. E.; Jurd, L. Tetrahedron 1964, 20, 1963.
- 25. a) Flores, Y.; Rodrigo, G.; Moffinedo, P.; Akesson, B.; Sterner, O.; Almanza, G. R. *Rev. Boliv. Quim.* 2009, 26, 21. b) Daily, A.; Seligmann, O.; Nonnenmacher, G.; Fessler, B.; Wong, S.; Wagner, H. *Planta Med* 1988, 54, 50. c) Zdero, C.; Bohlmann, F.; Solomon, J. *Phytochemistry* 1988, 27, 891.
- 26. a) Keen, N. T.; Ingham, J. L. Z. Naturforsch. 1980, 35c, 923. b) O'Neill, M. J.; Adesanya, S. A.; Roberts, M. F.; Pantry, I. R. Phytochemistry 1986, 25, 1315. c) Chakravarti, K. K.; Bose, A. K.; Siddiqui, S. J. Sci. Ind. Res. 1948, 7B, 24. d) Rao, G. V.; Annamalai, T.; Kavitha, K.; Mukhopadhyay, T. Res. J. Chem. Sci. 2012, 2, 50.
- a) Shul'ts, E. E.; Petrova, T. N.; Shakirov, M. M.; Chernyak, E. I.; Tolstikov, G. A. *Chem. Nat. Compd.* 2000, *36*, 362. b) Saitoh, T.; Shibata, S. *Chem. Pharm. Bull.* 1969, *17*, 729. c) Shiozawa, T.; Urata, S.; Kinoshita, T.; Saitoh, T. *Chem. Pharm. Bull.* 1989, *37*, 2239. d) Fukai, T.; Wang, Q. H.; Kitagawa, T.; Kusano, K.; Nomura, T.; Iitaka, Y. *Heterocycles* 1989, *29*, 1761. e) Ryu, Y. B.; Kim, J. H.; Park, S-J.; Chang, J. S.; Rho, M-C.; Bae, K-H.; Park, K. H.; Lee, W. S. *Bioorg. Med. Chem. Lett.* 2010, *20*, 971. f) Shibano, M.; Henmi, A.; Matsumoto, Y.; Kusano, G.; Miyase, T.; Hatakeyama, Y. *Heterocycles* 1997, *45*, 2053.
- 28. a) O'Neill, M. J.; Adesanya, S. A.; Roberts, M. F. *Phytochemistry* 1984, 23, 2704. b) Nguyen,
  P-H.; Nguyen, T-N-A.; Dao, T-T.; Kang, H-W.; Ndinteh, D-T.; Mbafor, J-T.; Oh, W-K. J. *Nat. Prod.* 2010, 73, 598.
- Nkengfack, A. E.; Vouffo, T. W.; Vardamides, J. C.; Fomum, Z. T.; Bergendorff, O.; Sterner, O. *J. Nat. Prod.* **1994**, *57*, 1172.
- 30. Ramakrishna, K. V.; Khan, R. A.; Kapil, R. S. Indian J. Chem. 1988, 27B, 285.
- 31. a) Ingham, J. L.; Tahara, S.; Dziedzic, S. Z. Z. Naturforsch. 1988, 43, 5. b) Kemertelidze, É. P.; Syrov, V. N.; Alaniya, M. D.; Kavtaradze, N. S.; Khushbaktova, Z. A. Pharm. Chem. J. 2008, 42, 340.

- 32. Jain, A. C.; Gupta, G. K.; Rao, P. R. Indian J. Chem. 1974, 12, 659.
- 33. a) Sharma, H. C.; Norris, D. M. J. Sci. Food Agric. 1991, 55, 353. b) Keen, N. T.; Taylor, O. C. Plant Physiol. 1975, 55, 731. c) Berlin, J.; Barz, W. Planta 1971, 98, 300. d) Kouam, J.; Tane, P.; Alain, M. L.; Noundou, X. S.; Choudhary, M. I.; Fomum, Z. T. Nat. Prod. Commun. 2007, 2, 835.
- 34. Zhang, D-W.; Yang, Y.; Yao, F.; Yu, Q-Y.; Dai, S-J. J. Nat. Med. 2012, 66, 362.
- 35. Gupta, G. K.; Dhar, K. L.; Atal, C. K. Phytochemistry 1977, 16, 403.
- 36. Gupta, S.; Jha, B. N.; Gupta, G. K.; Gupta, B. K.; Dhar, K. L. Phytochemistry 1990, 29, 2371.
- 37. Chen, Y.; Wei, X.; Xie, H.; Deng, H. J. Nat. Prod. 2008, 71, 929.
- 38. Gupta, B. K.; Gupta, G. K.; Dhar, K. L.; Atal, C. K. Phytochemistry 1980, 19, 2232.
- 39. Eisenbeiss, J.; Schmid, H. Helv. Chim. Acta 1959, 42, 61.
- 40. Rasool, N.; Khan, A. Q.; Ahmad, V. U.; Malik, A. Phytochemistry 1991, 30, 2800.
- 41. a) Komatsu, M.; Yokoe, I.; Shirataki, Y. *Chem. Pharm. Bull.* 1981, 29, 532. b) Ruan, B.;
  Kong, L-Y.; Takaya, Y.; Niwa, M. J. Asian Nat. Prod. Res. 2007, 9, 41.
- 42. Prasad, A. V. K.; Kapil, R. S.; Popli, S. P. Indian J. Chem. 1985, 24B, 236.
- 43. a) Ya, J.; Zhang, X-Q.; Wang, Y.; Zhang, Q-W.; Chen, J-X.; Ye, W-C. *Nat. Prod. Res.* 2010, 24, 621. b) Shou, Q. Y.; Tan, Q.; Shen, Z. W. *Bioorg. Med. Chem. Lett.* 2009, 19, 3389. c) Zheng, S.; Fu, R.; Shen, Z. *Chin. J. Chem.* 2012, 30, 1405.
- 44. Kishore, P. H.; Reddy, M. V. B.; Gunasekar, D.; Murthy, M. M.; Gaux, C.; Bodo, B. *Chem. Pharm. Bull.* **2003**, *51*, 194.
- 45. Ganapaty, S.; Srilakshmi, G. V. K.; Pannakal, S. T.; Rahman, H.; Laatsch, H.; Brun, R. *Phytochemistry* **2009**, *70*, 95.
- 46. Choi, Y-H.; Hong, S. S.; Shin, Y. S.; Hwang, B. Y.; Park, S-Y.; Lee, D. Arch. Pharm. Res. **2010**, *33*, 1651.
- 47. Lee, S. J.; Baek, H. J.; Lee, C. H.; Kim, H. P. Arch. Pharm. Res. 1994, 17, 31.
- 48. Zhang, C-F.; Sun, Z-H.; Zhang, D.; Zhang, M. Biochem. Syst. Ecol. 2010, 38, 1253.
- 49. Le-Van, N. Phytochemistry 1984, 23, 1204.
- 50. Bhandari, P.; Rastogi, R. P. Phytochemistry 1981, 20, 2044.
- 51. Nchancho, K.; Kouam, J.; Tane, P.; Kuete, V.; Watchueng, J.; Fomum, Z. T. *Nat. Prod. Commun.* **2009**, *4*, 931.
- 52. Bhargava, K. K.; Krishnaswamy, N. R.; Seshadri, T. R. Indian J. Chem. 1972, 10, 810.
- 53. Kouam, J.; Mabeku, L. B. K.; Kuiate, J. R.; Tiabou, A. T.; Fomum, Z. T. *Int. J. Chem.* **2011**, *3*, 23.
- 54. a) Nehybova, T.; Smarda, J.; Benes, P. Anticancer Agents Med. Chem. 2014, 14, 1351. b)
  Poluzzi, E.; Piccinni, C.; Raschi, E.; Rampa, A.; Recanatini, M.; De Ponti, F. Current Med. Chem. 2014, 21, 417. c) Bedell, S.; Nachtigall, M.; Naftolin, F. J. Steroid Biochem. Mol. Biol. 2014, 139, 225. d) Mostrom, M.; Evans, T. J. Reproductive and Developmental

Toxicology 2011, 707. e) Sammartino, A.; Gargano, V.; Di Carlo, C.; Tommaselli, G. A.;
Nappi, C. Recent Progress in Medicinal Plants 2008, 20, 367. f) Prakash, D.; Suri, S. Indian
J. Agric. Biochem. 2005, 18, 1. g) Maekelae, S.; Gustafsson, J-A. Phytoestrogens and Health
2002, 235. h) Mithun, N. M.; Shashidhara, S.; Vivek, K. R. Pharmacology. 2011, 1, 345.

- 55. a) Wang, P-L.; Yao, Z-H.; Zhang, F-X.; Shen, X-Y.; Dai, Y.; Qin, L.; Yao, X-S. J. Pharm. Biomed. Anal. 2015, 112, 23. b) Yuan, F.; Chen, J.; Sun, P-P.; Guan, S.; Xu, J. J. Biomed. Sci. 2013, 20, 84. c) Prakash, K. M. M. S.; Naidu, P. V. S.; Muralidhar, P. Int. J. Pharm. Tech. 2011, 3, 2868.
- 56. a) Idris, A. I.; Libouban, H.; Nyangoga, H.; Landao-Bassonga, E.; Chappard, D.; Ralston, S. H. *Mol. Cancer Ther.* 2009, *8*, 2339. b) Vender, J. R.; Laird, M. D.; Dhandapani, K. M.; *Neurosurgery* 2008, *62*, 1122; discussion 1027.
- 57. Wagner, H.; Fessler, B. Planta Med. 1986, 52, 374.
- Pôças, E. S. C.; Lopes, D. V. S.; da Silva, A. J. M.; Pimenta, P. H. C.; Leitão, F. B.; Netto, C. D.; Buarque, C. D.; Brito, F. V.; Costa, P. R. R.; Noël, F. *Bioorg. Med. Chem.* 2006, *14*, 7962.
- Kaushik-Basu, N.; Bopda-Waffo, A.; Talele, T. T.; Basu, A.; Costa, P. R.; da Silva, A. J.; Sarafianos, S. G.; Noël, F. *Nucleic Acids Res.* 2008, *36*, 1482.
- Syed, S. D.; Deepak, M.; Yogisha, S.; Chandrashekar, A. P.; Muddarachappa, K. A.; D'Souza,
   P.; Agarwal, A.; Venkataraman, B. V. *Phytother. Res.* 2003, *17*, 420.
- 61. Dalal, S.; Rana, S.; Sastry, K.; Kataria, S. Internet J. Microbiol. 2009, 7, 1.
- 62. Dalal, S.; Kataria, S. K. Asian J. Chem. 2010, 22, 7336.
- 63. a) Wagner, H.; Geyer, B.; Kiso, Y.; Hikino, H.; Rao, G. S. *Planta Med.* 1986, 52, 370. b)
  Singh, B.; Saxena, A. K.; Chandan, B. K.; Agarwal, S. G.; Anand, K. K. *Indian J. Physiol. Pharmacol.* 2001, 45, 435. c) Sagar, B. P. S.; Panwar, R.; Goswami, A.; Kadian, K.; Tyagi,
  K.; Chugh, M.; Dalal, S.; Zafar, R. *Pharm. Biol.* 2006, 44, 554.
- 64. a) Melo, P. A.; Ownby, C. L. *Toxicon* 1999, *37*, 199. b) Mors, W. B.; do Nascimento, M. C.; Parente, J.; da Silva, M. H.; Melo, P. A.; Suarez-Kurtz, G. *Toxicon* 1989, *27*, 1003. c) Melo, P. A.; Do Nascimento, M. C.; Mors, W.B.; Suarez-Kurtz, G. *Toxicon* 1994, *32*, 595. d) Diogo, L.C.; Fernandes, R. S.; Marcussi, S.; Menaldo, D. L.; Roberto, P. G.; Matrangulo, P. V. F.; Pereira, P. S.; França, S. C.; Giuliatti, S.; Soares, A. M.; Lourenço, M. V. *Basic Clin. Pharmacol. Toxicol.* 2009, *104*, 293. e) Pereira, N. A.; Pereira, B. M. R.; do Nascimento, M. C.; Parente, J. P.; Mors, W. B. *Planta Med.* 1994, *60*, 99.
- 65. a) Kobori, M.; Yang, Z.; Gong, D.; Heissmeyer, V.; Zhu, H.; Jung, Y-K.; Angelica, M.; Gakidis, M.; Rao, A.; Sekine, T.; Ikegami, F.; Yuan, C.; Yuan, J. *Cell Death Differ.* 2004, *11*, 123. b) Lin, F-M.; Chen, L-R.; Lin, E-H.; Ke, F-C.; Chen, H-Y.; Tsai, M-J.; Hsiao, P-W. *Carcinogenesis*, 2007, *28*, 2521. c) Tsai, C-H.; Lin, F-M; Yang, Y-C.; Lee, M-T.; Cha, T-L.; Wu, G-J.; Hsieh, S-C.; Hsiao, P-W. *Clin. Cancer Res.* 2009, *15*, 5435. d) Benes, P.; Knopfova, L.; Trcka, F.; Nemajerova, A.; Pinheiro, D.; Soucek, K.; Fojta, M.; Smarda, J.

*Cancer Lett.* **2011**, *303*, 29. e) Benes, P.; Alexova, P.; Knopfova, L.; Spanova, A.; Smarda, J. *Environ. Mol. Mutagen.* **2012**, *53*, 515.

- 66. a) Upadhyay, K.; Gupta, N. K.; Dixit, V. K. *Drug Dev. Ind. Pharm.* **2012**, *38*, 1152. b) Emmanuel, S.; Amalraj, T.; Ignacimuthu, S. *Indian J. Exp. Biol.* **2001**, *39*, 1305.
- 67. Sarveswaran, S.; Gautam, S. C.; Ghosh, J. Int. J. Oncol. 2012, 41, 2191.
- 68. Červeň, J.; Havran, L.; Pečinka, P.; Fojta, M. Electroanalysis 2015, 27, 2268.
- Chen, Z.; Sun, X.; Shen, S.; Zhang, H.; Ma, X.; Liu, J.; Kuang, S. Yu, Q. J. Biol. Chem. 2013, 288, 14417.
- 70. Prajapati, H.; Patel, M. B. Chem. Bio. Interf. 2012, 2, 38.
- 71. Li, L.; Huang, X-J.; Peng, J-L.; Zheng, M-Y.; Zhong, D-F.; Zhang, C-F.; Chen, X-Y. *Phytomedicine* **2016**, *23*, 340.
- 72. a) Whitten, P. L.; Naftolin, F. *Baillieres Clin. Endocrinol. Metab.* 1998, *12*, 667. b) Kuiper, G. G. J. M.; Lemmen, J. G.; Carlsson, B.; Corton, J. C.; Safe, S. H.; Van Der Saag, P. T.; Van Der Burg, B.; Gustafsson, J-Å. *Endocrinology* 1998, *139*, 4252. c) Kuiper, G. G. J. M.; Carlsson, B.; Grandien, K.; Enmark, E.; Häeggblad, J.; Nilsson, S.; Gustafsson, J-Å. *Endocrinology* 1997, *138*, 863. d) Gélinas, S.; Martinoli, M-G. *J. Neurosci. Res.* 2002, *70*, 90. e) Hess-Wilson, J. K.; Boldison, J.; Weaver, K. E.; Knudsen, K. E. *Breast Cancer Res. Treat.* 2006, *96*, 279.
- 73. Tsutsumi, N. Biol. Pharm. Bull. 1995, 18, 1012.
- 74. a) Kostelac, D.; Rechkemmer, G.; Briviba, K. J. Agric. Food Chem. 2003, 51, 7632. b) Bickoff, E. M.; Livingston, A. L.; Hendrickson, A. P.; Booth, A. N. J. Agric. Food Chem. 1962, 10, 410. c) Folman, Y.; Pope, G. S. J. Endocrinol. 1969, 44, 213. d) Shemesh, M.; Lindner, H. R.; Ayalon, N. J. Reprod. Fert. 1972, 29, 1.
- 75. Wagner, H.; Geyer, B.; Kiso, Y.; Hikino, H.; Rao, G. Planta Med. 1986, 52, 370.
- 76. Kim, Y. C.; Oh, H.; Kim, B. S.; Kang, T-H.; Ko, E-K.; Han, Y. M.; Kim, B. Y.; Ahn, J. S. Planta Med. 2005, 71, 87.
- 77. a) Nehybova, T.; Smarda, J.; Benes, P. Anticancer Agents Med. Chem. 2014, 14, 1351. b)
  Lee, Y-J.; Lin, W-L.; Chen, N-F.; Chuang, S-K.; Tseng, T-H. Eur. J. Med. Chem. 2012, 56, 361.
- 78. Xu, M-Y.; Kim, Y. S. Food Chem. Toxicol. 2014, 74, 311.
- 79. Shin, E. M.; Zhou, H. Y.; Guo, L. Y.; Kim, J. A.; Lee, S. H.; Merfort, I.; Kang, S. S.; Kim, H. S.; Kim, S.; Kim, Y. S. *Int. Immunopharmacol.* **2008**, *8*, 1524.
- 80. a) Rich, J. R.; Keen, N. T.; Thomason, I. J. *Physiol. Plant Pathol.* 1977, 10, 105. b) Gnanamanickam, S. S.; Patil, S. S. *Physiol. Plant Pathol.* 1977, 10, 159. c) Simons, R.; Vincken, J-P.; Roidos, N.; Bovee, T. F. H.; van Iersel, M.; Verbruggen, M. A.; Gruppen, H. J. *Agric. Food Chem.* 2011, 59, 6748. d) Durango, D.; Pulgarin, N.; Echeverri, F.; Escobar, G.; Quinones, W. *Molecules* 2013, 18, 10609. e) Morandi, D.; Le Quere, J. L. *New Phytol.* 1991,

117, 75. f) Lyon, F. M.; Wood, R. K. S. *Physiol. Plant Pathol.* **1975**, *6*, 117. g) Olah, A. F.; Sherwood, R. T. *Phytopathology* **1971**, *61*, 65.

- a) Bickoff, E. M.; Livingston, A. L.; Booth, A. N. Arch. Biochem. Biophys. 1960, 88, 262. b)
   Bickoff, E. M.; Booth, A. N. U.S. Patent 2,890,116 1959. c) Shlyankevich, M. US. Patent 5,569,459 1996.
- 82. Tanaka, Y.; Kikuzaki, H.; Fukuda, S.; Nakatani, N. J. Nutr. Sci. Vitarninol 2001, 47, 270.
- 83. a) Farahani, M. S.; Bahramsoltani, R. F.; Mohammad, H.; Abdollahi, M.; Rahimi, R. *Rev. Neurosci.* 2015, 26, 305. b) Chen, Y.; Cheung, Y-T.; Kong, L-D.; Ng, T. B.; Qiao, C.; Mo, S-F.; Xu, H-X.; Kung, H-F. *Life Sci.* 2008, 82, 1117. c) Yi, L-T.; Li, Y-C.; Pan, Y.; Li, J-M.; Xu, Q.; Mo, S-F.; Qiao, C-F.; Jiang, F-X.; Xu, H-X.; Lu, X-B.; Kong, L-D.; Kung, H-F. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 2008, *32*, 510.
- 84. a) Da Silva, A. J. M.; Melo, P. A.; Silva, N. M. V.; Brito, F. V.; Buarque, C. D.; de Souza, D. V.; Rodrigues, V. P.; Poças, E. S. C.; Noël, F.; Albuquerque, E. X.; Costa, P. R. R. *Bioorg. Med. Chem. Lett.* 2001, *11*, 283. b) Pôças, E. S. C.; Lopes, D. V. S.; da Silva, A. J. M.; Pimenta, P. H. C.; Leitão, F. B.; Netto, C. D.; Buarque, C. D.; Brito, F. V.; Costa, P. R. R.; Noël, F. *Bioorg. Med. Chem. Lett.* 2006, *14*, 7962.
- 85. a) Lopes, D. V. S.; Caruso, R. R. B.; Castro, N. G.; Costa, P. R. R. da Silva, A. J. M.; Noël, F. *Eur. J. Pharmacol.* 2004, 495, 87. b) Pôças, E. S. C.; Costa, P. R. R.; da Silva, A. J. M.; Noël, F. *Biochem. Pharmacol.* 2003, 66, 2169. c) Nichols, D. B.; Leão, R. A. C.; Basu, A.; Chudayeu, M.; de Moraes, P. de F.; Talele, T. T.; Costa, P. R. R.; Kaushik-Basu, N. *Chem. Biol. Drug Des.* 2013, 81, 607. d) Pôças, E. S. C.; Touza, N. A.; Pimenta, P. H. C.; Leitão, F. B.; Neto, C. D.; da Silva, A. J. M.; Costa, P. R. R.; Noël, F. *Bioorg. Med. Chem.* 2008, 16, 8801.
- 86. Xi, G-L.; Liu, Z-Q. J. Agric. Food Chem. 2014, 62, 5636.
- 87. a) Grozinger, C. M.; Chao, E. D.; Blackwell, H. E.; Moazed, D.; Schreiber, S. L. J. Biol. Chem. 2001, 276, 38837. b) Angeleska, S.; Kefalas, P.; Detsi, A. Tetrahedron Lett. 2013, 54, 2325.
- 88. Wong, S. M.; Antus, S.; Gottsegen, A.; Fessler, B.; Rao, G. S.; Sonnenbichler, J.; Wagner, H. *Arzneimittelforschung* **1988**, *38*, 661.
- Kowalski, K.; Szczupak, Ł.; Oehninger, L.; Ott, I.; Hikisz, P.; Koceva-Chyla, A.; Therien, B. J. Organomet. Chem. 2014, 772-773, 49.
- 90. Soman, S. S.; Soni, J. N.; Inamdar, G. S.; Robertson, G. P. Der Pharma Chemica 2013, 5, 201.
- 91. a) Tuskaev, V. A. Pharm. Chem. J. 2013, 47, 1. b) Stadlbauer, W.; Kappe, T. Heterocycles 1993, 35, 1425.
- 92. Bowyer, W. J.; Robertson, A.; Whalley, W. B. J. Chem. Soc. 1957, 542.
- 93. Govindachari, T. R.; Nagarajan, K.; Parthasarathy, P. C. J. Chem. Soc. 1957, 548.

- 94. Deschampo-Vallet, C.; Mentzer, C. Compt. Rend. 1960, 251, 736.
- 95. Govindachari, T. R.; Nagarajan, K.; Parthasarathy, P. C. Tetrahedron 1961, 15, 129.
- 96. a) Emerson, O. H.; Bickoff, E. M. J. Am. Chem. Soc. 1958, 80, 4381. b) Emerson, O. H.; Bickoff, E. M. U. S. Pat. 2,863,915, 1958. c) Emerson, O. H.; Bickoff, E. M. U. S. Pat. 2,884,427, 1959.
- 97. Nasipuri, D.; Pyne, G. J. Chem. Soc. 1962, 3105.
- 98. Kalra, V. K.; Kukla, A. S.; Sheshadri, T. R. Tetrahedron Lett. 1967, 8, 2153.
- 99. Uma Rani, B. S.; Darbarwar, M. J. Indian Chem. Soc. 1986, 63, 1060.
- 100. Kappe, T.; Korbuly, G.; Stadlbauer, W. Chem. Ber. 1978, 111, 3857.
- 101. Laschober, R.; Kappe, T. Synthesis 1990, 5, 387.
- 102. Hong, F.; Chen, Y.; Lu, B.; Cheng, J. Adv. Synth. Catal. 2016, 358, 353.
- 103. Nolan, M-T.; Pardo, L. M.; Prendergast, A. M.; McGlacken, G. P. J. Org. Chem. 2015, 80, 10904.
- 104. Cheng, C.; Chen, W-W.; Xu, B.; Xu, M-H. Org. Chem. Front. 2016, 3, 1111.
- 105. Mackey, K.; Pardo, L. M.; Prendergast, A. M.; Nolan, M.-T.; Bateman, L. M.; McGlacken, G. P. Org. Lett. 2016, 18, 2540.
- 106. Singh, R. P.; Singh, D. Heterocycles 1985, 23, 903.
- 107. Burns, M. J.; Thatcher, R. J.; Taylor, R. J. K.; Fairlamb, I. J. S. Dalton Trans. 2010, 39, 10391.
- 108. Shah, P.; Santana, M. D.; García, J.; Serrano, J. L.; Naik, M.; Pednekar, S.; Kapdi, A. R. *Tetrahedron* **2013**, 69, 1446.
- 109. Kapdi, A. R.; Karbelkar, A.; Naik, M.; Pednekar, S.; Fischer, C.; Schulzke, C.; Tromp, M. *RSC Adv.* **2013**, *3*, 20905.
- 110. Majumdar, K. C.; Khan, A. T.; Gupta, A. K.; Kundu, A. K.; Choudhury, P. K. *Indian J. Chem.* **1992**, *31B*, 667.
- 111. a) Kappe, T.; Schmidt, H. Org. Prep. Proc. Int. 1972, 4, 233. b) Kappe, T.; Brandner, A. Z. Naturforsch. 1974, 29b, 292.
- 112. Tang, L.; Pang, Y.; Yan, Q.; Shi, L.; Huang, J.; Du, Y.; Zhao, K. J. Org. Chem. 2011, 76, 2744.
- 113. Kurosawa, K. Nogami, K. Bull. Chem. Soc. Jpn. 1976, 49, 1955.
- 114. Phansalkar, M. S.; Deshmukh, K. K.; Kelkar, S. L.; Wadia, M. S. *Indian J. Chem.* **1987**, *26B*, 562.
- 115. a) Mali, R. S.; Tilve, S. G. Indian J. Chem. 1988, 27B, 465. b) Mali, R. S.; Tilve, S. G. Synth. Commun. 1990, 20, 1781.
- 116. Pandit, S. B.; Gadre, S. Y. Synth. Commun. 1988, 18, 157.
- 117. Gong, D-H.; Li, C-Z.; Yuan, C-Y. Chin. J. Chem. 2001, 19, 522.
- 118. Chang, C-F.; Yang, L-Y.; Chang, S-W.; Fang, Y-T.; Lee, Y-J. Tetrahedron 2008, 64, 3661.
- 119. Tricotet, T.; Fleming, P.; Cotter, J.; Hogan, A-M. L.; Strohmann, C.; Gessner, V. H.; O'Shea, D. F. J. Am. Chem. Soc. 2009, 131, 3142.
- 120. Sheng, J.; Xu, T.; Zhang, E.; Zhang, X.; Wei, W.; Zou, Y. J. Nat. Prod. 2016, 79, 2749.
- 121. James, C. A.; Coelho, A. L.; Gevaert, M.; Forgione, P.; Snieckus, V. J. Org. Chem. 2009, 74, 4094.
- 122. Hiroya, K.; Suzuki, N.; Yasuhara, A.; Egawa, Y.; Kasano, A.; Sakamoto, T. J. Chem. Soc. Perkin Trans. 1 2000, 24, 4339.
- 123. McPherson, H. L.; Ponder, B. W. J. Heterocycl. Chem. 1976, 13, 909.
- 124. a) Jurd, L. *Tetrahedron Lett.* 1963, *4*, 1151. b) Jurd, L. *Chem. Ind.* 1963, 1165. c) Jurd, L. U. S. Pat. 3,165,537, 1965. d) Jurd, L. *J. Org. Chem.* 1964, *29*, 2602. e) Jurd, L. *Tetrahedron* 1966, *22*, 2913.
- 125. Jurd, L. J. Org. Chem. 1964, 29, 3036.
- 126. Spencer, R. R.; Knuckles, B. E.; Bickoff, E. M. J. Heterocycl. Chem. 1966, 3, 450.
- 127. Maeda, S.; Masuda, H.; Tokoroyama, T. Chem. Pharm. Bull. 1994, 42, 2536.
- 128. Kshirsagar, U. A.; Parnes, R.; Goldshtein, H.; Ofir, R.; Zarivach, R.; Pappo, D. *Chem. Eur. J.* **2013**, *19*, 13575.
- 129. Larock, R. C.; Harrison, L. W. J. Am. Chem. Soc. 1984, 106, 4218.
- 130. Yao, T.; Yue, D.; Larock, R. C. J. Org. Chem. 2005, 70, 9985.
- 131. Thasana, N.; Worayuthakarn, R.; Kradanrat, P.; Hohn, E.; Young, L.; Ruchirawat, S. J. Org. *Chem.* **2007**, *72*, 9379.
- 132. Kraus, G. A.; Zhang, N. J. Org. Chem. 2000, 65, 5644.
- 133. Liu, J.; Liu, Y.; Du, W.; Dong, Y.; Liu, J.; Wang, M. J. Org. Chem. 2013, 78, 7293.
- 134. Donnelly, D. M. X.; Eades, J. F. K.; Philbin, E. M.; Wheeler, T. S. Chem. Ind. 1961, 1453.
- 135. Wanzlick, H-W.; Gritzky, R.; Heidepriem, H. Chem. Ber. 1963, 96, 305.
- 136. a) Fukui, K.; Nakayama, M. *Tetrahedron Lett.* 1965, *6*, 2559. b) Dholakia, V. N.; Trivedi, K. N. *J. Indian Chem. Soc.* 1971, *48*, 351. c) Shah, K. R.; Trivedi, K. N. *J. Indian Chem. Soc.* 1975, *52*, 224.
- 137. a) Grujić, Z.; Tabaković, I.; Trkovnik, M. *Tetrahedron Lett.* 1976, 17, 4823. b) Tabaković, I.; Grujić, Z.; Bejtović, Z. J. *Heterocycl. Chem.* 1983, 20, 635. c) Golabi, S. M.; Nematollahi, D. J. *Electroanal. Chem.* 1997, 420, 127. d) Golabi, S. M.; Nematollahi, D. J. *Electroanal. Chem.* 1997, 430, 141.
- 138. a) Pandey, G.; Muralikrishna, C.; Bhalerao, U. T. *Tetrahedron* 1989, 45, 6867. b) Bhalerao,
  U. T.; Muralikrishna, C.; Pandey, G. *Synth. Commun.* 1989, 19, 1303.
- 139. Leutbecher, H.; Conrad, J.; Klaiber, I.; Beifuss, U. Synlett 2005, 3126.
- 140. Qwebani-Ogunleye, T.; Kolesnikova, N. I.; Steenkamp, P.; de Koning, C. B.; Brady, D.; Wellington, K. W. *Bioorg. Med. Chem.* 2017, 25, 1172.
- 141. Gong, D-H.; Zhang, L.; Li, J-F.; Yuan, J. Y.; Yuan, C. Y. Chin. J. Chem. 2004, 22, 925.

- 142. Shah, K. R.; Trivedi, K. N. J. Indian Chem. Soc 1979, 56, 995.
- 143. Angeleska, S.; Kefalas, P.; Detsi, A. Tetrahedron Lett. 2013, 54, 2325.
- 144. Neog, K.; Borah, A.; Gogoi, P. J. Org. Chem. 2016, 81, 11971.
- 145. Wagh, U. M.; Usgaonkar, R. N. Indian J Chem. 1976, 14B, 861.
- 146. Rodríguez, S. A.; Baumgartner, M. T. Tetrahedron Lett. 2010, 51, 5322.
- 147. Darbarwar, M.; Sundaramurthy, V.; Subba Rao, N. V. Proc. Indian Acad. Sci. A 1974, 80, 93.
- 148. Lee, Y. R.; Suk, J. Y.; Kim, B. S. Org. Lett. 2000, 2, 1387.
- 149. Dewick, P. M.; Barz, W.; Grisebach, H. J. Chem. Soc. D, Chem. Commun. 1969, 466b.
- 150. Krishna Prasad, A. V.; Kapil, R. S.; Popli, S. P. J. Chem. Soc. Perkin Trans 11986, 1561.
- 151. Ferreira, M. A.; Moir, M.; Thomson, R. H. J. Chem. Soc., Perkin Trans. 1 1974, 2429.
- 152. Gunning, P. J. M.; Kavanagh, P. J.; Meegan, M. J.; Donnelly, D. M. X. J. Chem. Soc. Perkin Trans 11977, 691.
- 153. Narkhede, D. D.; Iyer, P. R.; Rukmani Iyer, C. S. J. Nat. Prod. 1989, 52, 502.
- 154. Da Silva, A. J. M.; Netto, C. D.; Costa, P. R. R. J. Braz. Chem. Soc. 2004, 15, 979.
- 155. Bowyer, W. J.; Chatterjea, J. N.; Dhoubhadel, S. P.; Handford, B. O.; Whalley, W. B. J. *Chem. Soc.* **1964**, 4212.
- 156. Sant'Ana, D. P.; Pinho, V. D.; Maior, M. C. L. S.; Costa, P. R. R. *Tetrahedron Lett.* 2009, 50, 3753.
- 157. Fowler, K. J.; Ellis, J. L.; Morrow, G. W. Synth. Commun. 2013, 43, 1676.
- 158. Nayak, M.; Jung, Y.; Kim, I. Org. Biomol. Chem. 2016, 14, 8074.
- 159. Takeda, N.; Miyata, O.; Naito, T. Eur. J. Org. Chem. 2007, 2007, 1491.
- 160. Ghosh, R.; Stridfeldt, E.; Olofsson, B. Chem. Eur J. 2014, 20, 8888.
- 161. Chatterjea, J. N.; Roy, S. K. J Indian Chem. Soc. 1957, 34, 98.
- 162. Chatterjea, J. N. J. Indian Chem. Soc. 1959, 36, 254.
- 163. Kawase, Y. Bull. Chem. Soc. Jpn. 1959, 32, 690.
- 164. Kawase, Y. Bull. Chem. Soc. Jpn. 1962, 35, 573.
- 165. Chatterjea, J. N.; Prasad, N. Chem. Ber. 1964, 97, 1252.
- 166. Zhang, J.; Qiu, J.; Xiao, C.; Yu, L.; Yang, F.; Tang, J. Eur. J. Org. Chem. 2016, 2016, 3380.
- 167. Kamara, B. I.; Brandt, E. V.; Ferreira, D. Tetrahedron 1999, 55, 861.
- 168. Farkas, L.; Gottsegen, À.; Nóagrádi, M.; Antus, S. J. Chem. Soc., Perkin Trans. 1 1974, 305.
- 169. Farkas, L.; Antus, S.; Nogradi, M. Acta Chim. Acad. Sci. Hung. 1974, 82, 225.
- 170. Litinas, K. E.; Stampelos, X. N. J. Chem. Soc. Perkin Trans. 1 1992, 2981.
- 171. Chiang, Y.; Gaplovsky, M.; Kresge, A. J.; Leung, K. H.; Ley, C.; Mac, M.; Persy, G.; Phillips, D. L.; Popik, V. V.; Rödig, C.; Wirz, J.; Zhu, Y. J. Am. Chem. Soc. 2003, 125, 12872.
- 172. Tollari, S.; Palmisano, G.; Cenini, S.; Cravotto, G.; Giovenzana, G. B.; Penoni, A. *Synthesis* **2001**, *5*, 735.
- 173. Al-Maharik, N.; Botting, N. P. Tetrahedron 2004, 60, 1637.

- 174. Pahari, P.; Saikia, U. P.; Das, T. P.; Damodaran, C.; Rohr, J. Tetrahedron 2016, 72, 3324.
- 175. Liu, Y.; Liu, J.; Wang, M.; Liu, J.; Liu, Q. Adv. Synth. Catal. 2012, 354, 2678.
- 176. Walter, R. Zimmer, H.; Purcell, T. C. J. Org. Chem. 1966, 31, 3854.
- 177. Gong, D-H.; Li, C-Z.; Yuan, C-Y. Chin. J. Chem. 2001, 19, 522.
- 178. For selected reviews: a) Liu, J.; Chen, G.; Tan, Z. Adv. Synth. Catal. 2016, 358, 1174. b) Guo, X-X.; Gu, D-W.; Wu, Z.; Zhang, W. Chem. Rev. 2015, 115, 1622. c) Hirano, K.; Miura, M. Top. Catal. 2014, 57, 878. d) Allen, S. E.; Walvoord, R. R.; Padilla-Salinas, R.; Kozlowski, M. C. Chem. Rev. 2013, 113, 6234. e) Chiba, S. Bull. Chem. Soc. Jpn. 2013, 86, 1400. f) Hirano, K.; Miura, M. Chem. Commun. 2012, 48, 10704.
- 179. For selected reports: a) Guru, M. M.; Punniyamurthy, T. J. Org. Chem. 2012, 77, 5063. b) Huang, A.; Chen, Y.; Zhou, Y.; Guo, W.; Wu, X.; Ma, C. Org. Lett. 2013, 15, 5480. c) Moon, Y.; Kim, Y.; Hong, H.; Hong, S. Chem. Commun. 2013, 49, 8323. d) Gallardo-Donaire, J.; Martin, R. J. Am. Chem. Soc. 2013, 135, 9350. e) Li, X.; He, L.; Chen, H.; Wu, W.; Jiang, H. J. Org. Chem. 2013, 78, 3636. f) Wang, Z.; Ni, J.; Kuninobu, Y.; Kanai, M. Angew. Chem. Int. Ed. 2014, 53, 3496. g) Takamatsu, K.; Hirano, K.; Satoh, T.; Miura, M. Org. Lett. 2014, 16, 2892. h) Chen, F-J.; Liao, G.; Li, X.; Wu, J.; Shi, B-F. Org. Lett. 2014, 16, 5644. i) Takamatsu, K.; Hirano, K.; Satoh, T.; Miura, M. Org. J-J.; Lu, C-J.; Xu, M.; Gao, J-R.; Jia, Y-X. Org. Lett. 2015, 17, 3050.
- 180. Krishnaswamy, N. R.; Seshadri, T. R.; Sharma, B. R. Indian J. Chem. 1966, 4, 120.
- Matos, M. J.; Janeiro, P.; Santana, L.; Uriarte, E.; Oliveira-Brett, A. M. J. Electroanal. Chem. 2014, 726, 62.
- 182. Matos, M. J.; Terán, C.; Pérez-Castillo, Y.; Uriarte, E.; Santana, L.; Viña, D. J. Med. Chem. 2011, 54, 7127.
- 183. Yuk, H. J.; Lee, J. H.; Curtis-Long, M. J.; Lee, J. W.; Kim, Y. S.; Ryu, H. W.; Park, C. G.; Jeong, T.-S.; Park, K. H. *Food Chem.* **2011**, *126*, 1057.
- 184. a) Gonda, Z.; Tolnai, G. L.; Novák, Z. *Chem. Eur. J.* 2010, *16*, 11822. b) Arvela, R. K.; Leadbeater, N. E.; Sangi, M. S.; Williams, V. A.; Granados, P.; Singer, R. D. *J. Org. Chem.* 2005, *70*, 161. c) Bedford, R. B.; Nakamura, M.; Gower, N. J.; Haddow, M. F.; Hall, M. A.; Huwe, M.; Hashimoto, T.; Okopie, R. A. *Tetrahedron Lett.* 2009, *50*, 6110. d) Lauterbach, T.; Livendahl, M.; Rosellón, A.; Espinet, P. Echavarren, A. M. *Org. Lett.* 2010, *12*, 3006.
- Venkateswarlu, S.; Panchagnula, G. K.; Guraiah, M. B.; Subbaraju, G. V. *Tetrahedron* 2006, 62, 9855.






































































































































































































